USER GUIDE

applied biosystems®
by life technologies™

QuantStudio™ 6 and 7 Flex Real-Time PCR System Software

Getting Started Guide

for use with:
QuantStudio™ 6 and 7 Flex Real-Time PCR Systems

Publication Number 4489822
Revision A

life technologies™

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Roadmap

BOOKLET 1 **Getting Started with QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Experiments**

BOOKLET 2 **QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Getting Started Guide for Standard Curve Experiments**

BOOKLET 3 **QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Getting Started Guide for Relative Standard Curve and Comparative C_T Experiments**

- **PART I: Running Relative Standard Curve Experiments**
- **PART II: Running Comparative C_T Experiments**

BOOKLET 4 **QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Getting Started Guide for Genotyping Experiments**

BOOKLET 5 **QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Getting Started Guide for Presence/Absence Experiments**

BOOKLET 6 **QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Getting Started Guide for Melt Curve Experiments**

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Getting Started with QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Experiments

Booklet 1

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Contents

About This Guide

CAUTION! ABBREVIATED SAFETY ALERTS. Hazard symbols and hazard types specified in procedures may be abbreviated in this document. For the complete safety information, refer to the instrument user guide.

IMPORTANT! Before using this product, read and understand the information in the instrument user guide.

Revision history

Revision	Date	Description
A	October 2013	New document

Purpose

The *QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Experiments Getting Started Guide* functions as both a tutorial and as a guide for performing your own experiments on the QuantStudio™ 6 and 7 Flex Instruments.

Note: For differences between the QuantStudio™ 6 System and the QuantStudio™ 7 System, refer to the *QuantStudio™ 6 and 7 Flex Real-Time PCR Systems Maintenance and Administration Guide* (Pub. no. 4489821).

Prerequisites

This getting started guide is intended for personnel who have been specifically trained by Life Technologies. The manufacturer is not liable for damage or injury that results from use of this manual by unauthorized or untrained parties.

This guide uses conventions and terminology that assume a working knowledge of the Microsoft® Windows® operating system, the Internet, and Internet-based browsers.

Note: First-time users of the QuantStudio™ 6 and 7 Flex Real-Time PCR System Software, please read *Getting Started with QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Experiments* thoroughly. The booklet provides information and general instructions that are applicable to all the experiments described in this binder.

How to use these booklets as tutorials

Each booklet in this guide provides a tutorial for running an example experiment using QuantStudio™ 6 and 7 Flex Real-Time PCR System Software and the example data provided on the installation CD. The following booklets are provided:

- *Getting Started with QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Experiments* – introductory information and experiment workflow common to all experiments.
- *QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Getting Started Guide for Standard Curve Experiments* – designing, running, and analyzing a Standard Curve experiment.
- *QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Getting Started Guide for Relative Standard Curve and Comparative C_T Experiments* – designing, running, and analyzing Relative Standard Curve and Comparative C_T experiments.

  Note: This booklet also provides information on setting up, running, and analyzing a gene expression study of two Comparative C_T experiments.
- *QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Getting Started Guide for Genotyping Experiments* – designing, running, and analyzing a Genotyping experiment.
- *QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Getting Started Guide for Presence/Absence Experiments* – designing, running, and analyzing a Presence/Absence experiment.
- *QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Getting Started Guide for Melt Curve Experiments* – designing, running, and analyzing a Melt Curve experiment.
- *QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Experiments - Appendixes* – common information such as ordering information, additional documentation, and glossary.

  Note: In all booklets, the term "experiment" refers to the entire process of performing an experiment, including setup, run, and analysis.

How to use the guides with your own experiments

Each booklet contains instructions specific to an example experiment provided on the installation CD. However, you can use the booklets as guides for your own experiments; tips for running your own experiments are provided at various points in each booklet.

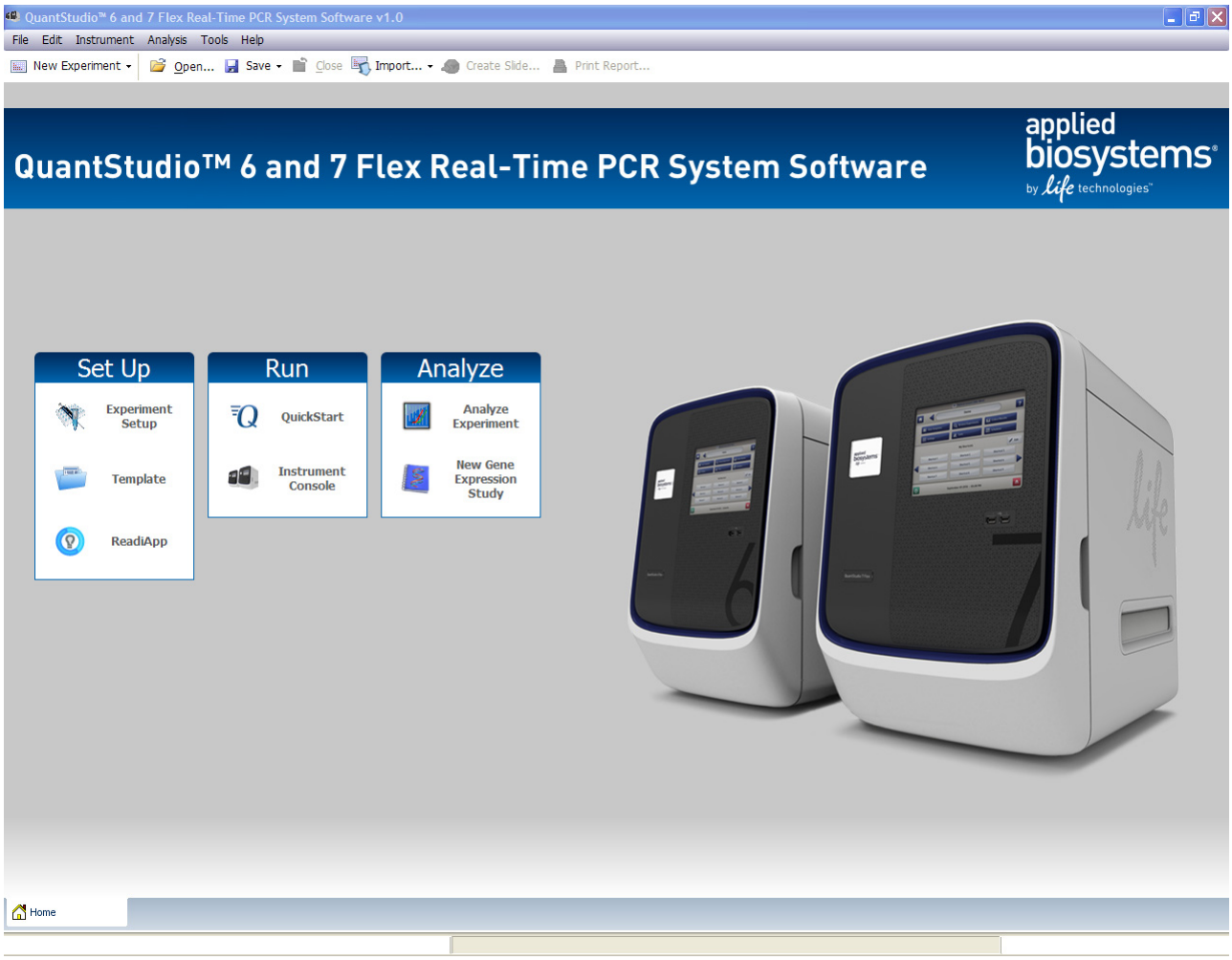
Assumptions

This guide assumes that you have access to the example experiments provided with the software.

How to access an example experiment

Start the QuantStudio™ 6 and 7 Flex Software

Double-click  (QuantStudio™ 6 and 7 Flex Real-Time PCR System Software shortcut) to access the Home screen, shown in the following image.



Open an example experiment

1. In the Home screen, select **Open** from the toolbar.
2. Navigate to the examples folder. The default path is: *<drive>*:\Program Files\Applied Biosystems\QuantStudio 6 and 7 Flex Software\examples\QS6Flex or *<drive>*:\Program Files\Applied Biosystems\QuantStudio 6 and 7 Flex Software\examples\QS7Flex.

   where, *<drive>* is the computer hard drive on which the QuantStudio™ 6 and 7 Flex Real-Time PCR System Software is installed. The default installation drive for the software is the C: drive.

3. Select an example experiment file to open, then click **Open**.

Experiment type	Example experiment file name
Standard Curve	QS6_96-Well Standard Curve Example.eds
	QS6_384-Well_Standard Curve Example.eds
	QS7_TaqMan_Array_Standard Curve Example.eds
	QS7_TaqMan_Array_RNaseP_Example.eds
	QS7_384-Well_Standard Curve Example.eds
	QS7_96-Well Standard Curve Example.eds
Relative Standard Curve	QS6_96-Well Relative Standard Curve Example 2.eds
	QS6_96-Well Relative Standard Curve Example.eds
	QS6_384-Well_Relative Standard Curve Example 2.eds
	QS6_384-Well_Relative Standard Curve Example.eds
	QS7_96-Well Relative Standard Curve Example 2.eds
	QS7_96-Well Relative Standard Curve Example.eds
	QS7_384-Well_Relative Standard Curve Example 2.eds
	QS7_384-Well_Relative Standard Curve Example.eds
Comparative C_T	QS6_96-Well Comparative Ct Example.eds
	QS6_384-Well_Comparative Ct_Example_1.eds
	QS6_384-Well_Comparative Ct_Example_2.eds
	QS6_384-Well_Comparative Ct_Example.eds
	QS7_96-Well Comparative Ct Example.eds
	QS7_384-Well_Comparative Ct_Example_1.eds
	QS7_384-Well_Comparative Ct_Example_2.eds
	QS7_384-Well_Comparative Ct_Example.eds
	QS7_TaqMan_Array_Comparative_Ct_Example.eds
Multiplex	QS6_96-Well Multiplex Example.eds
	QS6_384-Well_Multiplex_Example.eds
	QS7_96-Well Multiplex Example.eds
	QS7_384-Well_Multiplex_Example.eds
Genotyping	QS7_96-Well SNP Genotyping Example.eds
	QS7_384-Well_SNP_Genotyping_Example.eds
	QS6_96-Well SNP Genotyping Example.eds
	QS6_384-Well_SNP_Genotyping_Example.eds
Presence/Absence	QS6_384-Well_Presence-Absence_Example.eds
	QS6_96-Well Presence-Absence Example.eds
	QS7_96-Well Presence-Absence Example.eds
	QS7_384-Well_Presence-Absence_Example.eds

Experiment type	Example experiment file name
Melt Curve	QS6_96-Well SYBR Green PCR w Melt Example.eds
	QS6_384-Well_SYBR_Green_PCR_with_Melt_Example.eds
	QS6_384-Well_SYBR_Green_Melt_Example.eds
	QS6_384-Well_Melt_ Example.eds
	QS7_96-Well SYBR Green PCR w Melt Example.eds
	QS7_384-Well_SYBR_Green_PCR_with_Melt_Example.eds
	QS7_384-Well_SYBR_Green_Melt_Example.eds
	QS7_384-Well_Melt_ Example.eds

Note: In addition to the example experiment files, the following user sample files are located at: C:\Program Files\Applied Biosystems\QuantStudio 6 and 7 Flex Software\examples\User Sample Files

- Barcode template files
- Copy-paste example file
- Custom Sample Properties files
- Sample setup files
- Import custom fields file
- Export files

For more information on using the above files, see Chapter 2, "Experiment Shortcuts" on page 53.

A note on system security

The Security, Auditing, and e-Signature (SAE) feature in QuantStudio™ 6 and 7 Flex Real-Time PCR System Software enables role-based access control to enforce data integrity and authentication of users logging into the system, to strengthen system security. The feature tracks actions performed by users on experiments, templates, and studies, and it tracks changes to the SAE settings. You can enable or disable this feature to accommodate your security needs.

To enable or disable the feature, from the toolbar select **Tools ▸ Security ▸ Settings**.

For more information on the SAE feature, please refer to the instrument user guide.

User attention words

Five user attention words may appear in this document. Each word implies a particular level of observation or action as described below:

Note: Provides information that may be of interest or help but is not critical to the use of the product.

IMPORTANT! Provides information that is necessary for proper instrument operation or accurate chemistry kit use.

CAUTION! Indicates a potentially hazardous situation that, if not avoided, may result in minor or moderate injury. It may also be used to alert against unsafe practices.

WARNING! Indicates a potentially hazardous situation that, if not avoided, could result in death or serious injury.

DANGER! Indicates an imminently hazardous situation that, if not avoided, will result in death or serious injury.

Except for IMPORTANTs, the safety alert words in user documentation appear with an open triangle figure that contains a hazard symbol. These hazard symbols are identical to the hazard symbols that are affixed to the instrument. See the "Safety" appendix for descriptions of the symbols.

1 General Experiment Information and Instructions

This chapter covers:

Note: For more information about any of the topics discussed in this guide, access the Help from within QuantStudio™ 6 and 7 Flex Real-Time PCR System Software by pressing F1, clicking in the toolbar, or selecting **Help ▸ QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Help**.

Set up an experiment

Note: To start the QuantStudio™ 6 and 7 Flex Software, see "Start the QuantStudio™ 6 and 7 Flex Software" on page 7.

Define experiment properties

All experiments require the same general setup tasks; individual booklets supply specific parameters. The following procedures outline general steps to take to set up an experiment.

Access QuantStudio™ 6 and 7 Flex Software and click  (**Experiment Setup**). Click **Experiment Properties** to access the Experiment Properties screen.

Define experiment name and type

1. Enter a unique experiment name in the Experiment Name field. The default is a date and time stamp, which you can change. For example, 2010-04-12 173730.
   - Enter a name that is descriptive and easy to remember. You can enter up to 100 characters.
   - You can only use the alpha-numeric, hyphen (-), underscore (_), and spaces ( ) characters.

   Note: Ensure each experiment name is unique. If you have named two different experiments with the same name, you cannot run them on the same instrument. You will receive the following error message when attempting to start the run:

2. *(Optional)* Enter or scan the barcode on the reaction plate. You can enter up to 100 characters in the Barcode field.
3. *(Optional)* Enter a user name to identify the owner of the experiment. You can enter up to 100 characters in the User Name field.
4. *(Optional)* Enter comments to describe the experiment.
5. Select the instrument type you are using to run the experiment
   - QuantStudio™ 6 Flex System
   - QuantStudio™ 7 Flex System
6. Select the block type you are using to run the experiment
   - 384-Well Block
   - Array Card Block (only applicable to the QuantStudio™ 7 Flex System)

- 96-Well Block (0.2mL)
- Fast 96-Well Block (0.1mL)

7. Select the experiment type:
   - Standard Curve
   - Relative Standard Curve
   - Comparative C_T ($\Delta\Delta C_T$)
   - Melt Curve
   - Genotyping
   - Presence/Absence

Select the reagent

Select the reagent you are using to detect the target sequence:

- TaqMan® Reagents
- SYBR® Green Reagents
- Other

  Note: If you select SYBR® Green as the reagent, then you have the option of including a melt curve for that experiment.

Define the instrument run properties

1. Select the ramp speed for the experiment:
   - Standard
   - Fast
2. For Genotyping and Presence/Absence experiments, select the options for the data collection to include in the experiment run:
   - **Pre-PCR Read** - to include data before amplification occurs. Use the data collected during pre-PCR read to normalize fluorescence data collected during post-PCR read.
   - **Amplification** - to include real-time data.
   - **Post-PCR Read** - to include data after amplification has taken place.
3. For the Melt Curve experiment, select the **Include PCR** check box, to include PCR.

Review the analysis settings

Analysis Settings are different for each experiment type. The software analyzes the data using the default analysis settings. If the default analysis settings in the QuantStudio™ 6 and 7 Flex Software are not suitable for your own experiment, you can change the settings in the Analysis Settings dialog box and save the changed analysis settings to the Analysis Settings Library

Note: For information on Analysis Settings Library, refer to Booklet 7, *QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Experiments - Appendixes.*

Enter the reagent information

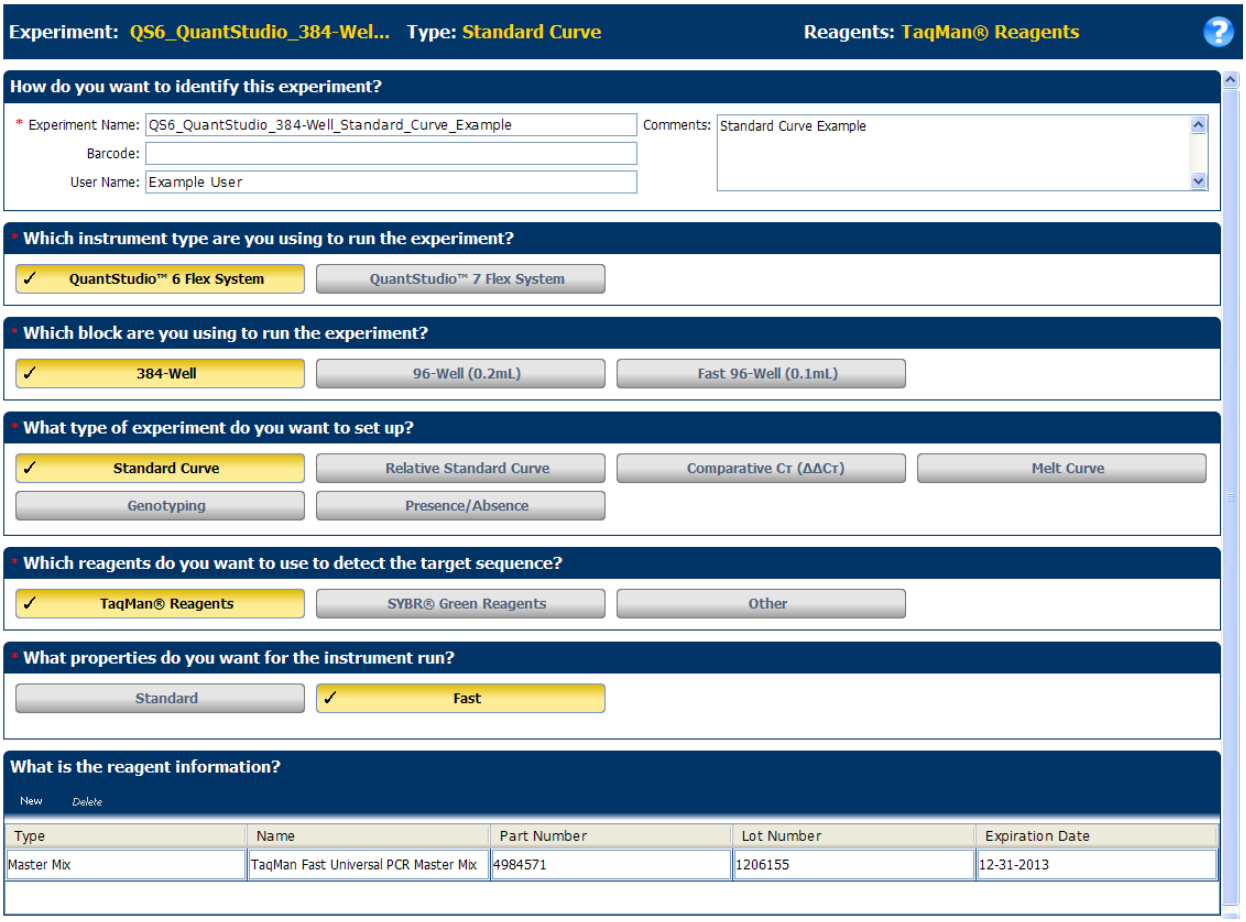
In the Reagent Information panel you can enter detailed reagent information, including the part number, lot number, and expiration date of the reagents you will use in your experiment. This information can be entered before setting up your experiment or starting your calibration run.

1. In the Reagent Information panel, click **New** to add a line for reagent details, or **Delete** to remove an existing one.
2. Click within the first four fields to enter your reagent **Type**, **Name**, **Part Number**, or **Lot Number**, respectively:
3. Click the Expiration Date field, and click the "down arrow" to display the current month's calendar. Select the reagent's expiration date from that month, or click the "forward arrow" to select a future date.

Save the experiment

Save the experiment. The default file name (.eds extension) is the experiment name that you entered when you set up the experiment and saved it for the first time. Changes to the experiment name after the first save do not update the file name. To change the file name, select **File** ▸ **Save As**.

The following is an image of the Experiment Properties screen for a Standard Curve experiment:



Define targets, samples, and biological replicate groups

Use the Define screen to define targets, samples and biological replicates for your experiment.

Note: You can start a run without these definitions, but there will be no real-time data (data will not be visible) in the amplification plots (the amplification plots can be seen only after you have set up the plate).

1. Click **Define** to access the Define screen.
2. Define targets.

Note: For Genotyping experiments, use this screen to specify the number of SNP assays to include in the experiment. For more information on defining SNP assays, refer to Booklet 4, *QuantStudio™ 6 and 7 Real-Time PCR System Software Getting Started Guide for Genotyping Experiments*.

   a. Click **New** to add targets and define them.
   b. In the target table, click a cell in the Target Name column for the target, then enter your target name. The default name is Target 1.
   c. Select the **Reporter** and **Quencher** from the respective drop-down menu.

      Note: The default reporter and quencher dyes used depend on the reagent selected during experiment setup. For example, if TaqMan® is the selected reagent, the default reporter FAM and default quencher is **NFQ-MGB**.

   d. Select the target **Color** from the drop-down menu.
   e. (*Optional*) Click **Save to Library** to save the newly added or existing edited targets to the target library.

      Note: Use the targets from the Target Library to avoid re-entering the information. Refer to Appendix B, Supplemental Information in Booklet 7, *QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Experiments - Appendixes* for information on target libraries.

   f. *(Optional)* Click **Import from Library** to add targets from the target library.

3. Define samples.
   a. Click **New** to add samples and name them.
   b. In the samples table, click a cell in the Sample Name column for the sample to define and enter your sample name. The default sample name is Sample 1.
   c. Select the sample **Color** from the drop-down menu.
   d. (*Optional*) Click **Save to Library** to save the newly added or existing edited samples to the sample library.

      Note: Use the samples from the Sample Library to avoid re-entering the information. Refer to Appendix B, Supplemental Information in Booklet 7, *QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Experiments - Appendixes* for information on sample libraries.

   e. (*Optional*) Click **Import from Library** to add samples from the sample library.

4. (*Optional*) Define biological replicates.
   a. In the Define Biological Replicates Groups table, click **New** to add biological replicate group and name them. You can enter up to 100 characters in this field.

b. Select the **Color** from the drop-down menu.

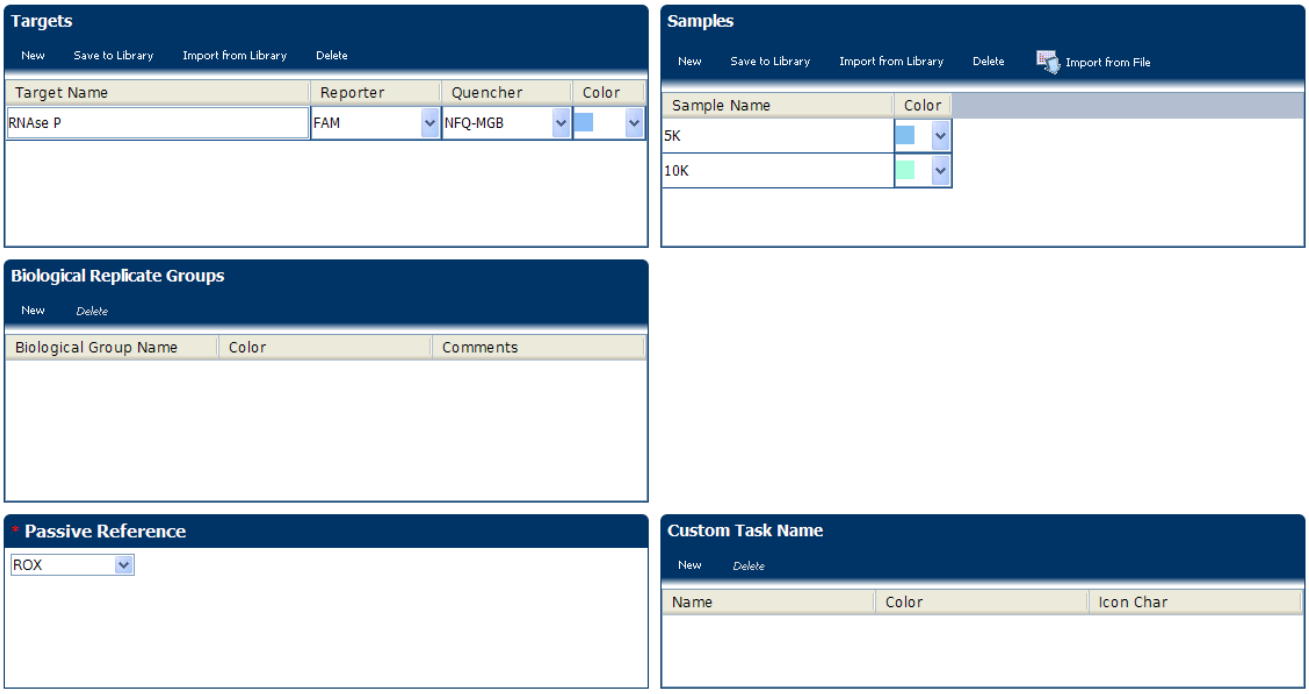
c. Click in the **Comments** column to add comments for that biological replicate group.

5. Select the Passive Reference dye from the drop-down menu.

6. Define custom task name.

   Note: The Custom Task Name panel is visible only when the **Hide the custom task name definition and assignment UI** check box under the Setup tab in the Preferences dialog box is unselected.

The following is an image of the Define screen for a Standard Curve experiment:



Assign targets, samples, and biological replicate groups

Use the Assign screen to assign targets, samples, and biological replicate groups to wells in the reaction plate. For Genotyping experiments, use this screen to assign SNP assays.

Note: You can start a run without these assignments, but there will be no real-time data in the amplification plots (the amplification plots can be seen only after you have set up the plate).

1. Click **Assign** to access the Assign screen.

2. Assign targets.

   a. Select wells using the plate layout or the well table on the Assign screen.

   b. Select a target and assign its task, in the plate, from the drop-down menu. Depending on the experiment type, options are:

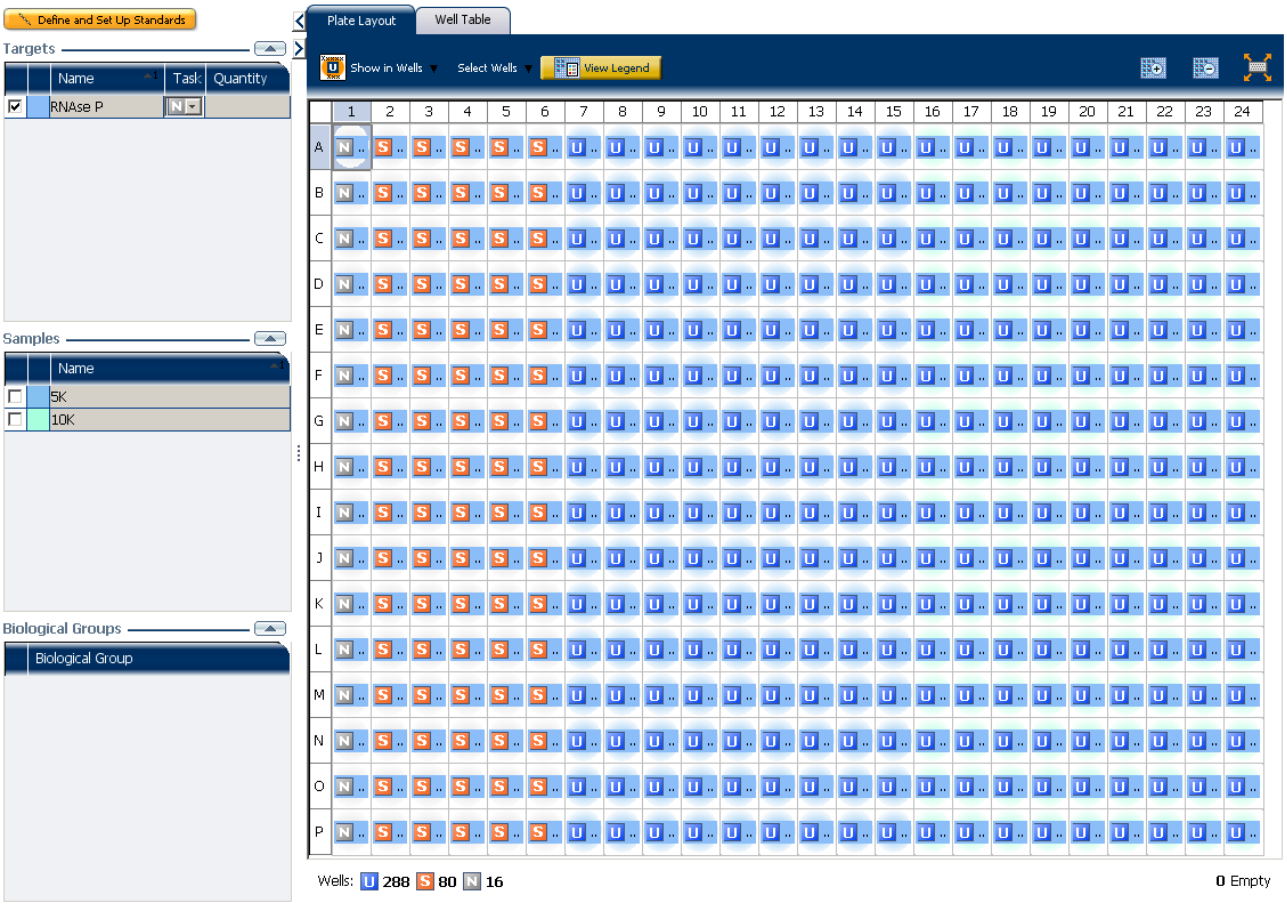
Experiment type	Legend	Tasks
Standard Curve	U	Unknown
	S	Standard
	N	Negative Control
Relative Standard Curve	U	Unknown
	S	Standard
	N	Negative Control
Comparative CT	U	Unknown
	N	Negative Control
Genotyping	U	Unknown
	1/1	Positive Control Allele 1/ Allele 1
	2/2	Positive Control Allele 2/ Allele 2
	1/2	Positive Control Allele 1/ Allele 2
	N	Negative Control
Presence/Absence	U	Unknown
	I	Internal Positive Control
	N	Negative Control
	~~I~~	Blocked Internal Positive Control
Melt Curve	U	Unknown
	N	Negative Control

3. Assign Samples.
   a. Select wells using the plate layout or the well table on the Assign screen.
   b. Select the check box next to the sample to assign to the selected wells.

   Note: You can assign only one sample to a well. If the selected wells contain mixed assignments (indicated by a ▣), remove existing sample assignments before you make the new sample assignment.

4. Assign Biological Replicate Groups.
   a. Select wells using the plate layout or the well table on the Assign screen.
   b. Select the check box next to the biological replicate group to assign to the selected wells.

The following is an image of the Assign screen for a Standard Curve experiment:

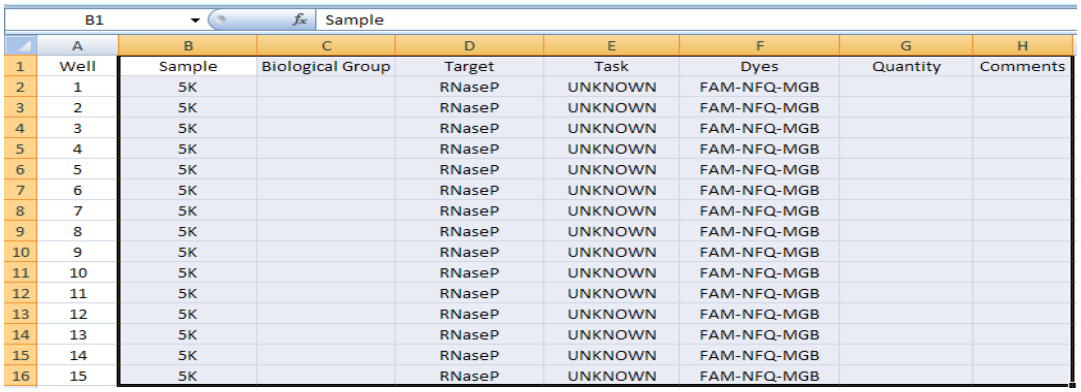


Assign targets, samples, and biological replicate groups - alternate procedure

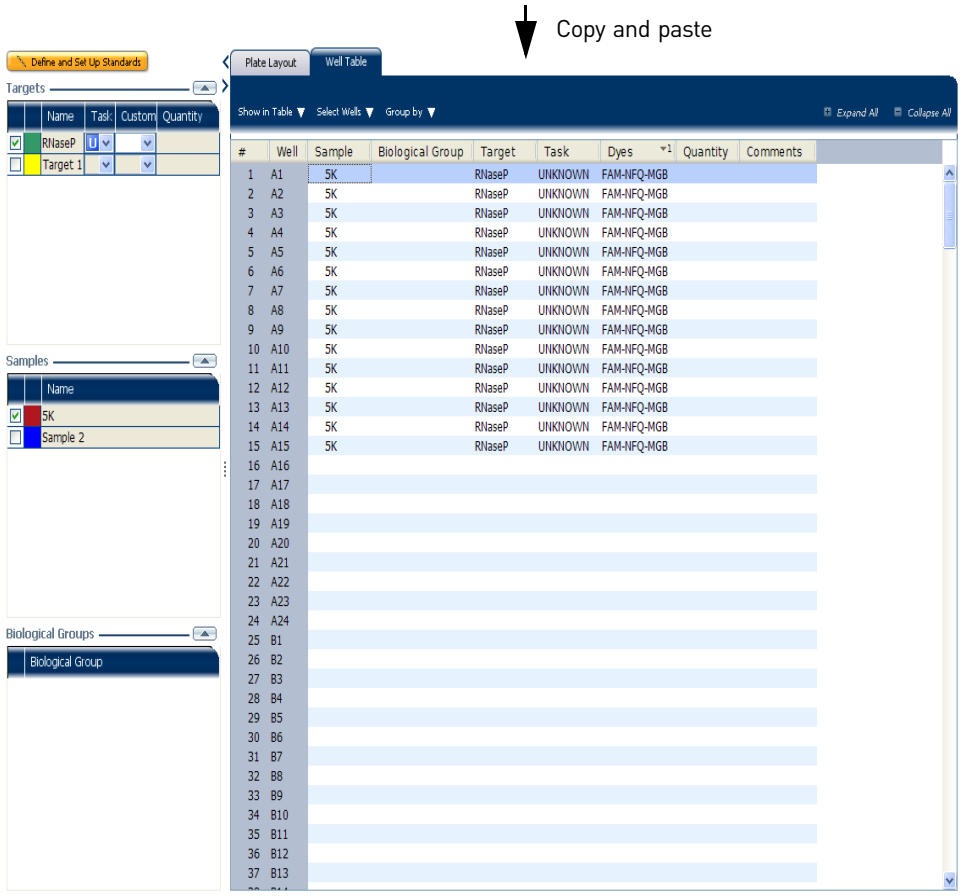
As shown below, you can also paste assignment information from an ***.xls** file into the plate layout of the QuantStudio™ 6 and 7 Flex Software for wells with single targets.

Note: You must select the header and the Well Number column while copying information from the ***.xls** file.

Note: Any of the columns not copied are treated as NULL values for those columns.



	A	B	C	D	E	F	G	H
1	Well	Sample	Biological Group	Target	Task	Dyes	Quantity	Comments
2	1	5K		RNaseP	UNKNOWN	FAM-NFQ-MGB		
3	2	5K		RNaseP	UNKNOWN	FAM-NFQ-MGB		
4	3	5K		RNaseP	UNKNOWN	FAM-NFQ-MGB		
5	4	5K		RNaseP	UNKNOWN	FAM-NFQ-MGB		
6	5	5K		RNaseP	UNKNOWN	FAM-NFQ-MGB		
7	6	5K		RNaseP	UNKNOWN	FAM-NFQ-MGB		
8	7	5K		RNaseP	UNKNOWN	FAM-NFQ-MGB		
9	8	5K		RNaseP	UNKNOWN	FAM-NFQ-MGB		
10	9	5K		RNaseP	UNKNOWN	FAM-NFQ-MGB		
11	10	5K		RNaseP	UNKNOWN	FAM-NFQ-MGB		
12	11	5K		RNaseP	UNKNOWN	FAM-NFQ-MGB		
13	12	5K		RNaseP	UNKNOWN	FAM-NFQ-MGB		
14	13	5K		RNaseP	UNKNOWN	FAM-NFQ-MGB		
15	14	5K		RNaseP	UNKNOWN	FAM-NFQ-MGB		
16	15	5K		RNaseP	UNKNOWN	FAM-NFQ-MGB		



Note: An example copy and paste file is provided with the QuantStudio™ 6 and 7 Flex Real-Time PCR System Software and is located at C:\Program Files\Applied Biosystems\QuantStudio 6 and 7 Flex Software\examples\User Sample Files.
where, *<drive>* is the computer hard drive on which the QuantStudio™ 6 and 7 Flex Software is installed. The default installation drive for the software is the C: drive.

Define the run method

Use the Run Method screen to set up the run method for your own experiments in the QuantStudio™ 6 and 7 Flex Software.

Note: Refer to the Booklet 7, *QuantStudio™ 6 and 7 Real-Time PCR System Software Experiments - Appendixes* for information on analysis settings.

1. Click **Run Method** to access the Run Method screen.

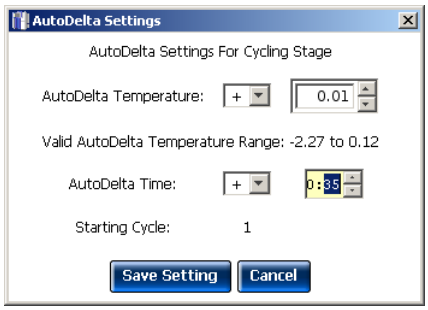
   Note: You can save multiple run methods to the Run Method Library for later use. Refer to Appendix B, Supplemental Information in Booklet 7, *QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Experiments - Appendixes* for information on run method libraries.

2. Enter a number for the reaction volume per well. See "Instrument consumables" on page 17 for maximum reaction volumes for the consumables supported by the QuantStudio™ 6 and 7 Flex Software.

3. In the **Graphical View** tab, review and, if necessary, edit the run method.
   - Make sure that the thermal profile is appropriate for your reagents.
   - Edit the default run method or replace it with one from the run method library included in the QuantStudio™ 6 and 7 Flex Software.
   - Enable data collection by clicking  .

     Note: Enabling data collection is especially useful when you later need to analyze data collected in real-time during the various stages.
   - Edit the ramp rate. You can increase or decrease the ramp rate for a stage.

     Note: Ramp rates are decimal numbers from 0.015—3.4.
   - Edit the PCR Stage.
     - Change the Number of Cycles for the PCR stage.
     - Select the **Enable AutoDelta** check box, to increase or decrease the temperature and/or hold time for each subsequent cycle or to change the Starting Cycle for AutoDelta. Enabling AutoDelta displays the ▲ icon. Click the AutoDelta Off ▲ icon to change the AutoDelta settings for the cycling stage in the AutoDelta Settings dialog box. Then, click **Save Setting** to display the AutoDelta On ▲ icon.

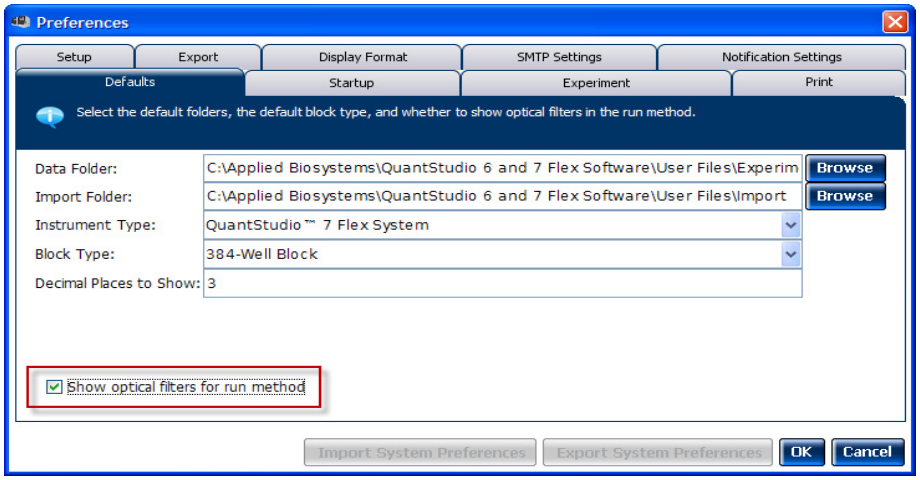


Note: If you selected SYBR® Green as the reagent, the Melt Curve stage automatically appears in the Run Method screen. If you delete the Melt Curve Stage section from the protocol, then the melt curve is active in the Add Stage drop-down menu.

4. *(Optional)* Complete the tasks on the Optical Filters tab:

IMPORTANT! Do not alter the optical filters for system dyes. This feature is optional when you use custom dyes, where you can select a filter set to match the profile of the dye. For more information on how to select the appropriate filter set, contact Life Technologies.

By default, the Optical Filters tab is not visible. To show the Optical Filters tab, go to **Tools** ▸ **Preferences**, and select the Show optical filters for run method check box under the Default tab.



- To add a new filter set to the filter set library, click **Save**.
- To load a saved filter set, click **Load**.
- To go back to the original filter set combinations, click **Revert to Defaults**.

Prepare reactions

Supported reagents

Life Technologies supports the following reagents for experiments performed with the QuantStudio™ 6 and 7 Flex Software.

Reagent	Experiment type
TaqMan® reagents	• Standard Curve • Relative Standard Curve • Comparative C_T ($\Delta\Delta C_T$) • Genotyping • Presence/ Absence
SYBR® Green reagents	• Standard Curve • Relative Standard Curve • Comparative C_T ($\Delta\Delta C_T$) • Melt Curve
Other reagents	• Standard Curve • Relative Standard Curve • Comparative C_T ($\Delta\Delta C_T$) • Genotyping • Presence/ Absence • Melt Curve

Note: The QuantStudio™ 6 and 7 Flex Software can accommodate other reagents, but performance claims have not been tested by Life Technologies.

Precautions while preparing reactions

- Do not prepare the reactions on a wet table. Wet surfaces lead to contamination of your reactions.
- Wear appropriate protective eyewear, clothing, and powder-free gloves.
- Use the appropriate consumables. The quality of pipettors and tips and the care used in measuring and mixing dilutions affect data accuracy.
- Perform dilutions exactly as instructed. Mistakes or inaccuracies in making the dilutions directly affect the quality of results.
- Use a permanent marker or pen to mark a tube and the side of a plate or array card. Do not use fluorescent markers.
- Ensure that the arrangement of the PCR reactions matches the plate layout displayed in the QuantStudio™ 6 and 7 Flex Software.

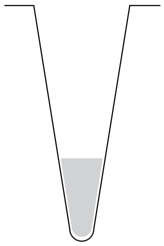
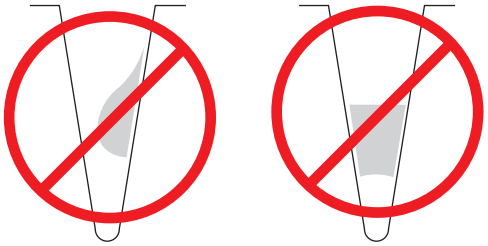
Materials required while preparing the dilutions

- DI water or DEPC water
- Microcentrifuge tubes
- Pipettors
- Pipette tips
- Vortex mixer
- Centrifuge
- Sample stock

- Standard stock
- Reaction mix components
- Plate or array card

Guidelines for preparing the dilutions, reaction mix, and plate

- Include excess volume in your calculations to provide excess volume for the loss that occurs during reagent transfers.
- Use TE buffer or water to dilute the standards and samples.
- Prepare the reagents according to the manufacturer's instructions.
- Keep the dilutions and assay mix protected from light, in the freezer, until you are ready to use it. Excessive exposure to light may affect the fluorescent probes or dyes.
- Prior to use:
  - Mix the master mix thoroughly by swirling the bottle.
  - Resuspend the assay mix by vortexing, then centrifuge the tube briefly.
  - Thaw any frozen samples by placing them on ice. When thawed, resuspend the samples by vortexing, then centrifuge the tubes briefly.
- Do not allow the bottom of the reaction plate to become dirty. Fluids and other contaminants that adhere to the bottom of the reaction plate can contaminate the sample block(s) and cause an abnormally high background signal.

Correct	Incorrect
	
Liquid is at the bottom of the well.	Not centrifuged with enough force *Or* Not centrifuged for enough time

- Place the reaction plate or array card at 4°C and in the dark until you are ready to load it into the instrument.

Seal the reaction plate

If you use optical adhesive film to seal your reaction plates, seal each reaction plate as follows:

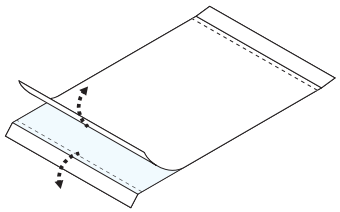
Note: The sealing instructions are applicable to 384-well and 96-well reaction plates.

1. Load the reaction plate using the plate layout described in "Assign targets, samples, and biological replicate groups" on page 16.

   Note: For 96-well reaction plates, place the reaction plate onto the center of the 96-well base, then perform this step. Ensure that the reaction plate is flush with the top surface of the 96-well base.

2. Remove a single optical adhesive film (film) from the box. Bend both end-tabs upward. Hold the film backing side up.

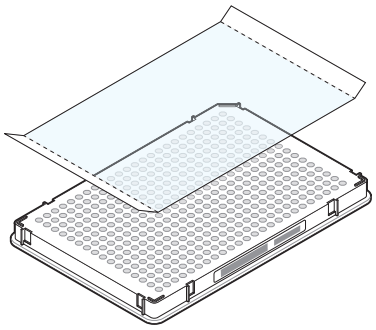
3. In one swift movement, peel back the white protective backing from the center sealing surface. Do not touch the center sealing surface.

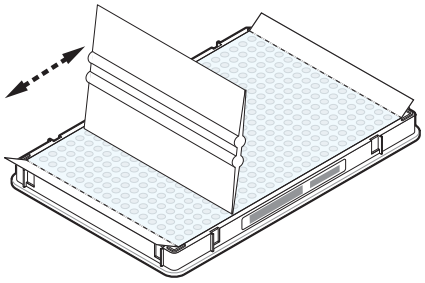


IMPORTANT! Improper peeling of the optical adhesive film may result in haziness, but it will not affect results. Haziness disappears when the film comes into contact with the heated cover in the instrument.

4. Holding the film by the end-tabs, lower the film onto the reaction plate (adhesive side facing the reaction plate). Ensure that the film completely covers all wells of the reaction plate.

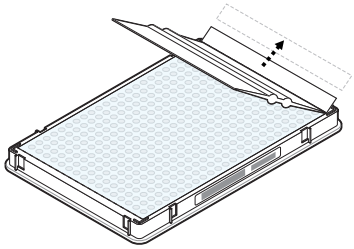
5. Applying firm pressure, move the applicator slowly across the film, horizontally and vertically, to ensure good contact between the film and the entire surface of the reaction plate.





6. Using the applicator to hold the edge of the film in place, grasp one end of the end-tab and pull up and away sharply. Repeat for the other end-tab.

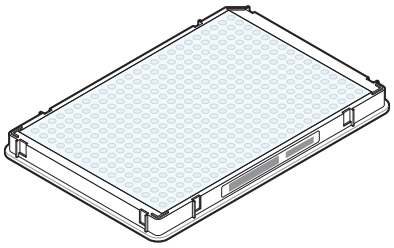
Note: Ensure clean removal of both end-tabs from the dotted line. Improper peeling of the end-tab can cause sticking of plate on the heated cover assembly.



7. To ensure a tight, evaporation-free seal, repeat step 5. Applying firm pressure, run the edge of the applicator along all four sides of the outside border of the film.

Note: Optical adhesive films do not adhere on contact. The films require the application of pressure to ensure a tight, evaporation-free seal.

8. Inspect the reaction plate to ensure that all wells are sealed. You should see an imprint of all wells on the surface of the film. The perforated tab should be completely torn off to avoid plates from sticking to the instrument after a run.



IMPORTANT! Remove all excess adhesive from the perimeter of the optical adhesive cover. When the film is applied, the glue from the optical adhesive cover can adhere to the edges of the plate. If the excess glue is not removed, the plate may adhere to the sample block of the QuantStudio™ 6 or 7 Instrument.

Fill and seal the array card

Fill and spin the array card

IMPORTANT! Wear powder-free gloves while preparing the Arrays.

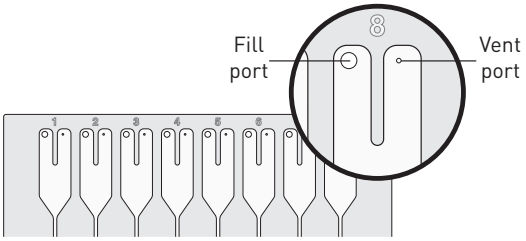
1. Remove an array card from its box and place it on a clean, dry surface.
2. Using a permanent marker, mark the side of the empty array cards.
3. Transfer the experiment-related chemistries and solutions into the port of the array card.

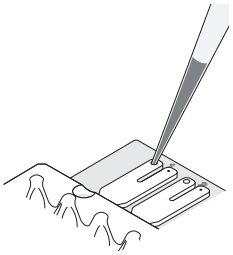
   For each transfer:

   a. Place the array card on a lab bench, with the foil side down.

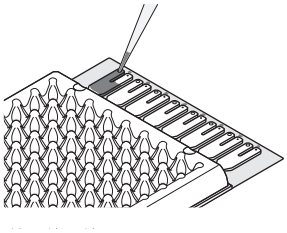
   b. Load 100 µL of fluid into a pipette.

c. Hold the pipette in an angled position (~45 degrees) and place the tip into the fill port. The fill port is the larger of the two holes on the left side of the fill reservoir.



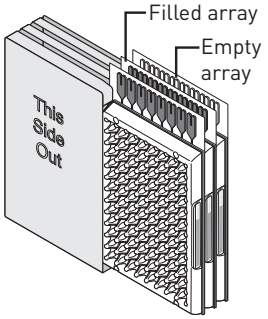


d. Dispense the fluid so that it sweeps in and around the fill reservoir toward the vent port. Pipet fluid into the fill reservoir, but **do not** go past the first stop of pipettor plunger when pipetting the reagents into the array card, or you may blow the solution out of the port.



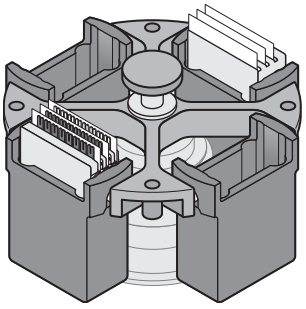
IMPORTANT! Do not allow the tip to contact and possibly damage the coated foil beneath the fill port.

4. Place the filled array card(s) into a centrifuge array card carrier clip and place empty array card(s) in the remaining slots. Confirm that the labels on the buckets and clips are oriented in the same direction.



IMPORTANT! Balance the loads in opposite buckets in the centrifuge.

5. Place the filled carrier clips into the centrifuge buckets. Ensure that the array card fill reservoirs and bucket and clip labels face outward when loaded into the centrifuge. Balance the loads in opposite buckets.



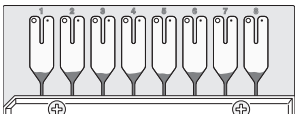
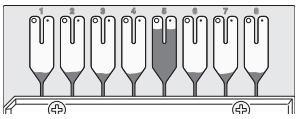
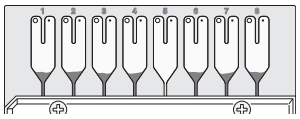
IMPORTANT! You must run the centrifuge with all four buckets in place and each of the two carriers filled with the array card. Place empty array cards into unfilled slots.

6. Close the centrifuge cover, then spin the array card(s) for 1 minute at 1200 rpm.

7. When the run is finished, stop the centrifuge, then spin the array card(s) again for 1 minute at 1200 rpm.

   IMPORTANT! Do not try to save time by doing one spin for 2 minutes. The two sets of ramps are important for a good fill into the array card.

8. When the second run is finished, open the centrifuge and check that the fluid levels in the reservoirs of each array card have decreased by the same amount. Also, check for the formation of bubbles in all wells and note possible problems.

Correct	Incorrect	
		

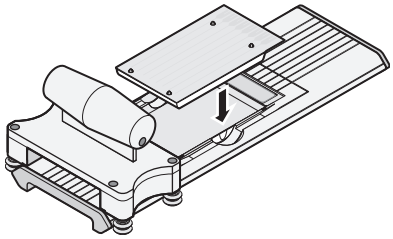
9. If necessary, centrifuge the array card(s) for an additional minute to fill any unfilled wells. Do not exceed three 1-minute runs or centrifuge the array card for longer than 1 minute at a time.

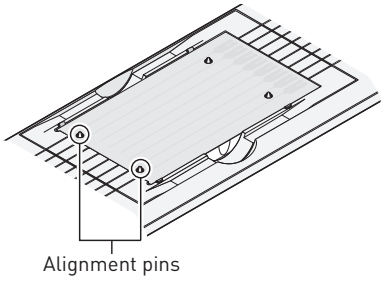
   Note: Contact Life Technologies for more information on loading an array card.

Seal the array card(s)

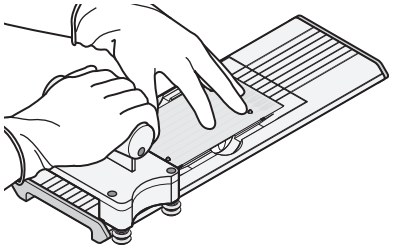
1. With the carriage (roller assembly) of the Array Card Staker/ Sealer in the Start position, place a filled array card into the fixture with the foil side up so that the fill reservoirs are the farthest away from the carriage.

2. Press down on all four corners of the array card to ensure that it is fully seated within the fixture.

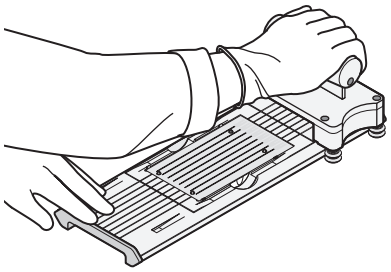




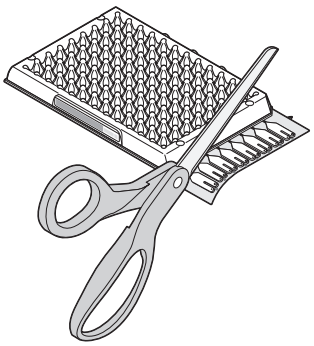
3. Use the two alignment pins in the fixture to position the array card correctly.



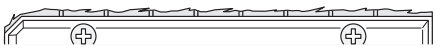
4. Seal the array card by running the carriage slowly over it. Run the carriage over the array card in one direction only. Do not apply downward force on the carriage as you move it forward over the card.



5. Remove the sealed array card from the fixture and trim the fill reservoirs from the array card assembly using scissors. Trim the foil array card so that the edge is even with the plastic carrier.



IMPORTANT! Completely remove the fill reservoirs from the array card so that the edge is free of residual plastic. The plastic from the fill reservoirs that extends beyond the edge of the card can prevent the card from seating properly on the sample block and affect amplification.

Correct	Incorrect
	

IMPORTANT! As you seal the remaining filled array cards, store them in a dark place until you are ready to use them. The fluorescent dyes in the array card are photosensitive. Prolonged exposure to light can diminish the fluorescence of the dye.

Capping and uncapping the 96-well reaction tubes and tube strips

WARNING! Use the flat caps for the 0.2 mL tubes and 0.1 mL tubes. Rounded caps can damage the heated cover.

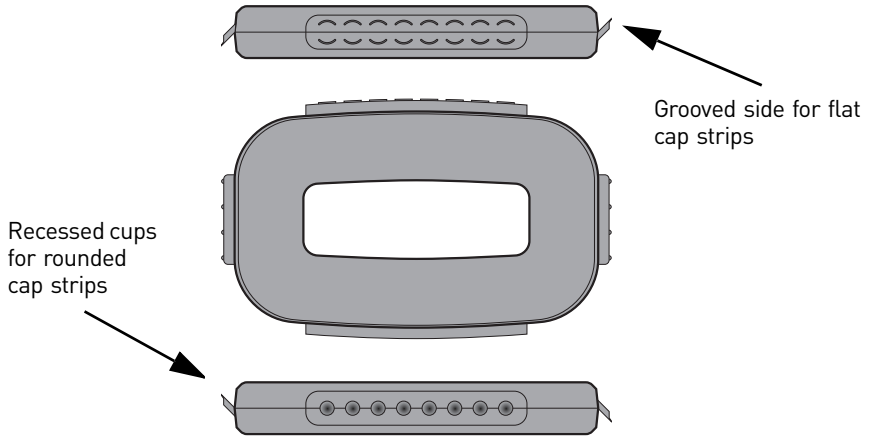
Note: Ensure that you secure the caps on the tubes and tube-strips tightly to avoid sample evaporation.

If you use the 96-well MicroAmp® Optical 8-Tube Strips or MicroAmp® Optical Tubes without Cap, use the MicroAmp® Cap Installing Tool and use the following instructions:

- Applying the MicroAmp® Optical 8-Cap Strip or MicroAmp® Optical Tubes without Cap to the tubes
- Removing a cap string from a plate

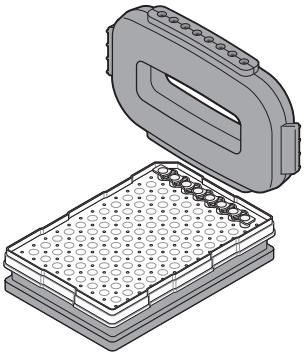
Required materials:

- MicroAmp® Cap Installing Tool
- MicroAmp® Optical 8-Tube Strips or MicroAmp® Optical Tubes without cap
- MicroAmp® Optical 8-Cap Strip



MicroAmp® Cap Installing Tool

Apply the MicroAmp® Optical 8-Cap Strip (flat)

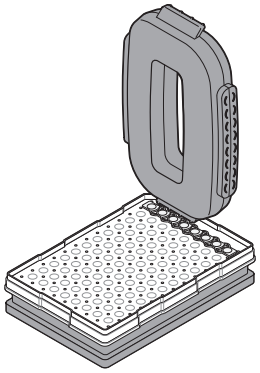


1. Grasp the Cap Installing Tool so that the grooved side is exposed.
2. Hold the strip of caps over the tube strip or the row of tubes.
3. Use the grooved side (shown) of the Cap Installing Tool to push and seat each cap firmly in place. Use a rocking motion to properly seat each cap.

Remove a cap string from a plate

The MicroAmp® Cap Installing Tool is also used for removing the MicroAmp® Optical 8-Cap Strip from the 96-well optical plates and tray/retainer assemblies. To remove the cap or cap strip:

1. Insert the small protrusions on the side of the Cap Installing Tool under the webbing between the caps on a cap strip.
2. Slowly pry the strip from the plate or Tray/Retainer assembly.



Start the experiment

To start an experiment:

1. Prepare the instrument for use as shown below.
2. Load the reaction plate or array card into the instrument, as shown on page 33.
3. Run the experiment as shown on page 34.

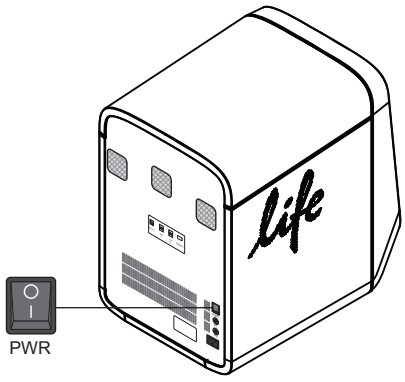
Prepare the instrument for use

Start the QuantStudio™ 6 or 7 Instrument

1. Touch anywhere on the touchscreen to determine if the QuantStudio™ 6 or 7 Instrument is in standby mode.

   Does the touchscreen display the Standby screen after you touch it?

   - **Yes** – The instrument is ready for use. Go to step 3 below.
   - **No** – Go to step 2 to power on the instrument.
2. Toggle the power button on the rear of the QuantStudio™ 6 or 7 Instrument, then wait for it to start.



   The QuantStudio™ 6 or 7 Instrument is ready to use when the touchscreen displays the Main Menu.
3. Power on the monitor.
4. Power on the computer:

a. Press the computer power button, then wait for it to start.

b. When the Login screen appears, enter your user name and password, then click **OK**.

5. Start the QuantStudio™ 6 and 7 Flex Software:

a. From the desktop, double-click  QuantStudio™ 6 and 7 Flex Software.

Note: If the shortcut is not present on the desktop, select **Start ▸ All Programs ▸ Applied Biosystems ▸ QuantStudio™ 6 and 7 Flex Software ▸ QuantStudio™ 6 and 7 Flex Software** to start the software.

IMPORTANT! If the QuantStudio™ 6 and 7 Flex Software will not start, confirm that no other instances of the instrument control software are open. If any instance of the software is open, close it before starting the QuantStudio™ 6 and 7 Flex Software.

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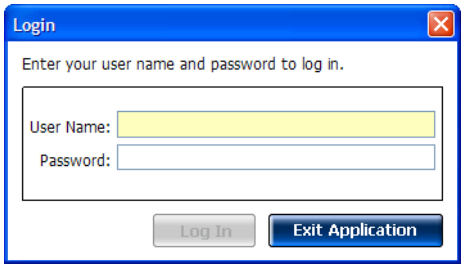
b. From the Login dialog box, enter your user name and password, then click **Log In**.



Note: If the QuantStudio™ 6 and 7 Flex Software displays the License Central screen after you log into the software, your license file may be corrupt. Contact Life Technologies support to obtain a replacement license file.

Add the instrument to the My Instruments group

Before you can use the QuantStudio™ 6 or 7 Instrument, you must add the instrument to the "My Instruments" group in the QuantStudio™ 6 and 7 Flex Software.

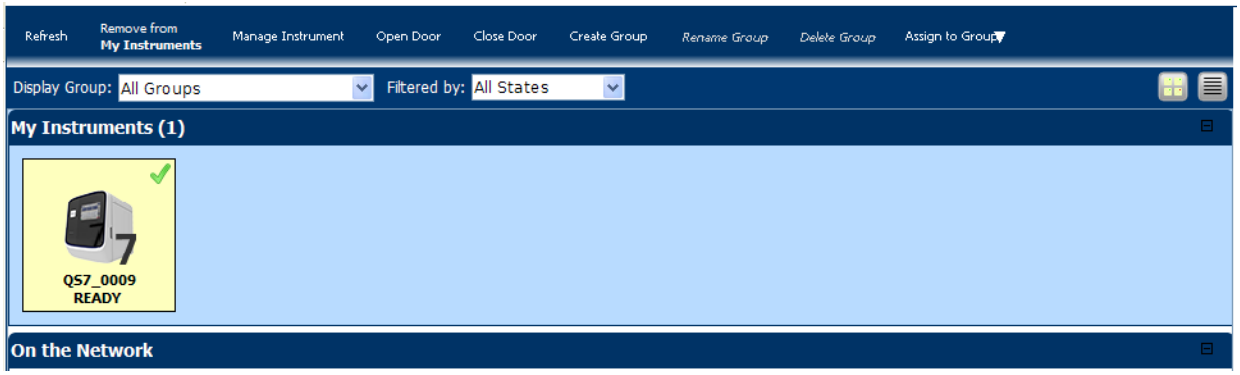
1. Power on the instrument and start the software as explained in "Start the QuantStudio™ 6 or 7 Instrument" on page 30.

2. From the QuantStudio™ 6 and 7 Flex Software Home tab, click  **Instrument Console**.

3. From the Instrument Console, confirm the instrument state:

a. Confirm that the instrument icon appears in the My Instruments group.

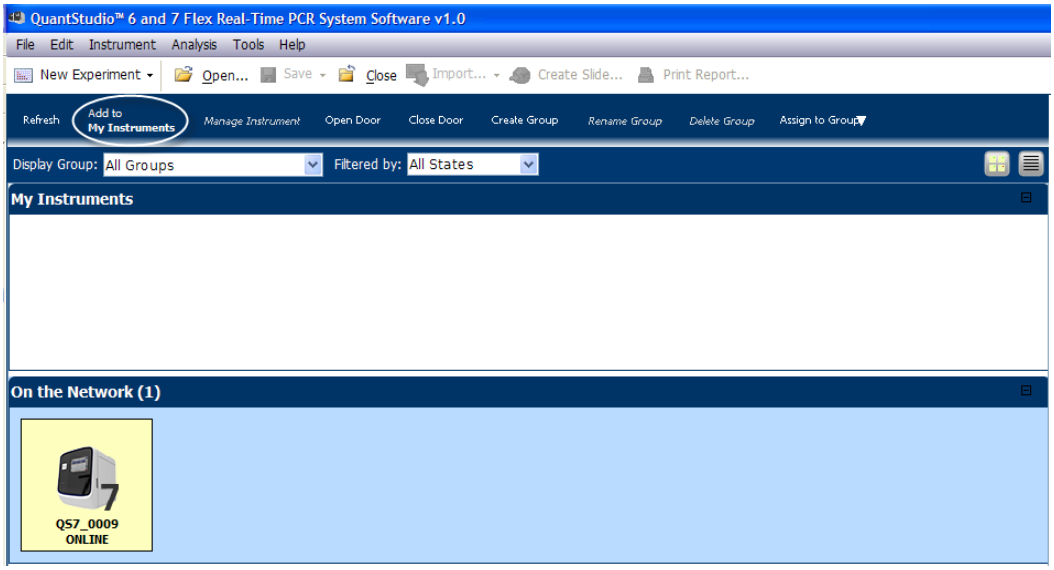
b. Confirm that a green check box appears in the upper-right corner of the instrument icon.



4. If your instrument does not appear within the My Instruments group, add it as follows:

   a. From the Instrument Console, select your QuantStudio™ 6 or 7 Instrument from the list of instruments on the network.

   b. Click **Add to My Instruments**.



Note: The details for a QuantStudio™ 6 or 7 Instrument in the My Preferred list can be exported even if the network connection has been interrupted. The exported details from the disconnected instrument would contain the data most recently downloaded from the instrument before the interruption.

Enable or change the Notification Settings

You can configure the QuantStudio™ 6 and 7 Flex Software to alert you by email when the QuantStudio™ 6 or 7 Instrument begins and completes a run, or if an error occurs during a run.

Note: For details on using the Notification Settings feature, refer to the instrument user guide.

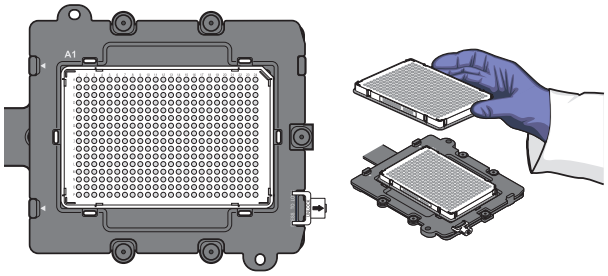
Load the reaction plate or array card into the instrument

CAUTION! PHYSICAL INJURY HAZARD. During instrument operation, the temperature of the sample block(s) can exceed 100°C. Keep your hands away until the sample block(s) reaches room temperature.

IMPORTANT! Wear powder-free gloves when you handle the **reaction plate or array card**.

IMPORTANT! Plates and array cards should be loaded and unloaded by operators who have been warned of the moving parts hazard and have been adequately trained.

1. Eject the QuantStudio™ 6 or 7 Instrument tray by doing either of the following:
   - From the QuantStudio™ 6 or 7 Instrument touchscreen, touch .
   - From the QuantStudio™ 6 and 7 Flex Software, select **Instrument** ▸ **Instrument Console**, select your instrument icon, then click **Open Door**.
2. Load the reaction plate or array card into the plate adapter. When you load the reaction plate or array card, ensure that:
   - Well A1 is positioned at the top-left of the tray for any of the plate formats.
   - The barcode (for any of the plate formats) is facing the front of the instrument.



   - If using reaction tubes or tube strips, make sure you use adaptors. The adaptors are attached to the plate transport arm. The tray containing the tubes or tube strips must be placed on the adaptor and not into the sample block directly.

     IMPORTANT! For optimal performance with partial loads, load at least 16 tubes in one of the following arrangements. You can load empty tubes if you do not have enough reaction volume to load the required number of tubes:

     Adjacent columns of 8 tubes, using rows A through H. For example, use wells in columns 6 and 7 (rows A through H).

     Or

     Adjacent rows of 8 tubes, using columns 3 through 10. For example, use wells in row D (columns 3 through 10) and row E (columns 3 through 10).

3. Close the QuantStudio™ 6 or 7 Instrument tray by doing either of the following:
   - From the instrument touchscreen, touch .
   - From the Instrument Console screen, click **Close Door**.

Start the experiment

IMPORTANT! Ensure that instrument calibration is up-to-date. If a calibration has expired, you will get a warning when you start a run. For information on calibrating the QuantStudio™ 6 or 7 Instrument, refer to instrument user guide.

IMPORTANT! Do not attempt to open the access door during the run. The door is locked while the QuantStudio™ 6 or 7 Instrument is in operation.

Note: Ensure each experiment name is unique. If you have named two different experiments with the same name, you cannot run them on the same instrument. You will receive the following error message when attempting to start the run:

If you do not want to delete the existing experiment, rename the duplicate experiment and then proceed to the run.

You can run the experiment in either of the following two ways:

- Start the experiment from the QuantStudio™ 6 and 7 Flex Software
- Start the experiment from the QuantStudio™ 6 or 7 Instrument touchscreen

Note: The example experiments in each of the getting started guide booklets start a run from the QuantStudio™ 6 and 7 Flex Software.

Start the experiment from the QuantStudio™ 6 and 7 Flex Software

1. In the QuantStudio™ 6 and 7 Flex Software, click **Run** in the navigation pane.

   IMPORTANT! Ensure that the *.eds file you created is open before you start a run.

2. Click **START RUN**. Select the instrument for the run from the My Instruments drop-down menu.

   IMPORTANT! If the preferred instrument for running the experiment is not present under My Instruments or the custom group, or if it is unavailable, clicking START RUN does not display instrument names in the drop-down menu. See "Add the instrument to the My Instruments group" on page 31 for instructions on adding an instrument to the My Instruments group.

Start the experiment from the QuantStudio™ 6 or 7 Instrument touchscreen

1. Touch the QuantStudio™ 6 or 7 Instrument touchscreen to awaken it.

   Note: If the touchscreen is not at the Main Menu screen, touch  .

2. In the Main Menu screen, touch **Browse Experiments**.

3. In the Browse screen, touch  **Folders**, to display the folders containing the experiment setup files.

4. Touch any of the folder names to display the experiments in that folder.

   Note: You can create and save new experiments from the QuantStudio™ 6 or 7 Instrument touchscreen, or transfer experiments created and saved in the QuantStudio™ 6 and 7 Flex Software to folders in the QuantStudio™ 6 or 7 Instrument touchscreen via a USB flash drive.

5. In the Experiments screen, select the desired experiment, then: to view or edit the experiment before starting the run.
   - Touch 

View/Edit, then go to step 6 to view or edit the experiment before starting the run.
   - Touch 

Save and Start Run, then go to step 7 to start the run immediately.

6. *(Optional)* Modify the experiment parameters as needed.
   a. In the Edit Experiment screen, you can use the:
      - 

Add and 

Delete buttons to add and delete a stage or step to the thermal profile.
      - 

Add Melt Curve button to add a melt curve to the thermal profile.
      - **Save** button to save the experiment you modify.

   b. In the Save Experiment screen, touch each field to edit the:
      - Experiment name
      - Folder to save the experiment
      - Reaction volume
      - Barcode Number
      - Notes

      When finished, touch 

Save & Start Run to start the experiment.

7. In the Start Run screen, touch each field as needed to modify the associated parameter, then touch 

Start Run Now to start the experiment.

Monitor the experiment

Note: If the connection between the QuantStudio™ 6 and 7 Flex Software and the QuantStudio™ 6 or 7 Instrument is disrupted while running an experiment, remove and then add the instrument to the My Instruments list in the Instrument Console. You may then resume monitoring the experiment.

You can monitor an experiment run in three ways:

- From the Run screen of the QuantStudio™ 6 and 7 Flex Software, while the experiment is in progress, as shown below.
- From the Instrument Console of the QuantStudio™ 6 and 7 Flex Software (to monitor an experiment started from another computer or from the QuantStudio™ 6 or 7 Instrument touchscreen) as described in "From the QuantStudio™ 6 and 7 Flex Software Instrument Console" on page 36.
- From the QuantStudio™ 6 or 7 Instrument touchscreen, as described in "From the QuantStudio™ 6 or 7 Instrument touchscreen" on page 40.

From the QuantStudio™ 6 and 7 Flex Software Run screen

1. Click **Amplification Plot** from the Run Experiment Menu to monitor the amplification plot of the experiment you are running.

   Note: For Melt Curve experiments, click **Melt Curve Plot** from the Run Experiment Menu.

2. Click **Temperature Plot** from the Run Experiment Menu to monitor the temperature plot of the experiment you are running.

From the QuantStudio™ 6 and 7 Flex Software Instrument Console

1. In the Instrument Console screen, select the icon of the instrument that you are using to run the experiment.
2. Click **Manage Instrument**.
3. On the Instrument Manager screen, click **Monitor Running Instrument**.

You can view the progress of the run in real time from the Run screen. During the run, periodically view the Amplification Plot, Temperature Plot and Run Method (see page 37, 38, and 38 respectively) available from the QuantStudio™ 6 and 7 Flex Software for potential problems.

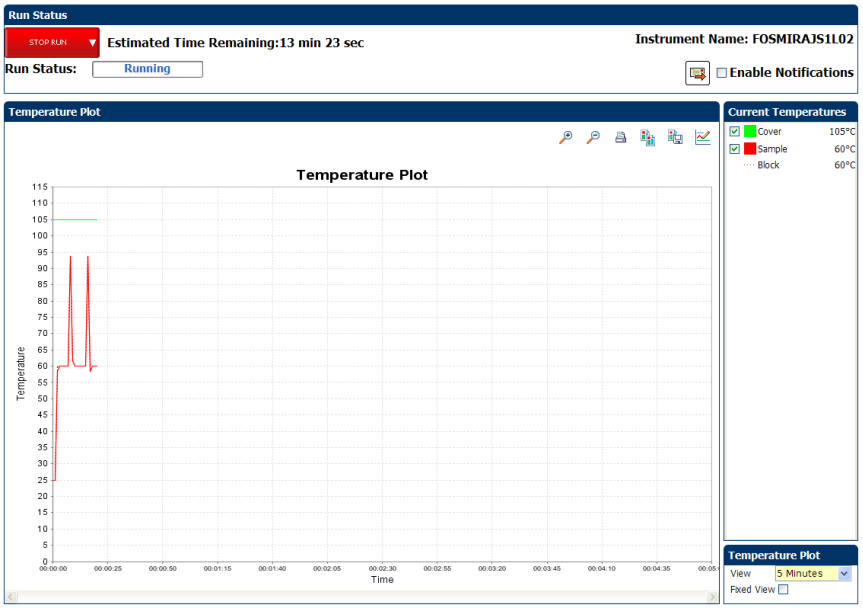
To...	Action
Stop the run	• In the QuantStudio™ 6 and 7 Flex Software, click **STOP RUN**. • In the Stop Run dialog, click one of the following: – **Stop Immediately** to stop the run immediately. – **Stop after Current Cycle/Hold** to stop the run after the current cycle or hold. – **Cancel** to continue the run.
View amplification data in real time	Select **Amplification Plot**. See "To monitor the Amplification Plot" on page 37.
View temperature data for the run in real time	Select **Temperature Plot**. See "To monitor the Temperature Plot" on page 38.
View progress of the run in the Run Method screen	Select **Run Method**. See "To monitor the Run Method" on page 38.

To...	Action
Enable/disable the Notification Settings	Select or deselect **Enable Notifications**. See "Enable or change the Notification Settings" on page 32.

Note: The individual experiment booklets provide illustrations of the different experiments in real time.

Note: For Melt Curve experiments, click **Melt Curve Plot** from the Run Experiment Menu.

The following is an image of the Run screen for a Standard Curve experiment:



To monitor the Amplification Plot

To view data in the Amplification Plot, click **Amplification Plot** from the Run Experiment Menu, select the Plate Layout tab, then select the wells to view.

The Amplification Plot screen allows you to view sample amplification as your instrument collects fluorescence data during a run. If a method is set up to collect real-time data, the Amplification Plot screen displays the data for the wells selected in the Plate Layout tab. The plot contrasts normalized dye fluorescence (ΔRn) and cycle number.

The Amplification Plot screen is useful for identifying and examining abnormal amplification, including:

- Increased fluorescence in negative control wells.
- Absence of detectable fluorescence at an expected cycle (determined from previous similar experiments run using the same reagents under the same conditions).

  Note: If you notice abnormal amplification or a complete absence of fluorescence, refer to the instrument user guide to troubleshoot the error.

To monitor the Temperature Plot

To view data in the Temperature Plot screen, click **Temperature Plot** from the Run Experiment Menu.

During a run, the Temperature Plot screen displays the temperatures of the sample block(s), the heated cover, and samples (calculated) in real-time.

To...	Action
Add or remove temperature plots	Select **Cover** or **Sample Block** to view the presence of the associated data in the plot.
Change the time to display in the plot	From the **View** drop-down menu, select the amount of time to display in the plot.
Display a fixed time window during the instrument run	Select **Fixed View**. If the entire plot does not fit in the screen, the screen is not updated as the run progresses. For example, if you select 10 minutes from the View drop-down menu, the plot will show data for 10 minutes. If the Fixed View is: • Deselected, the plot updates as the run progresses even after 10 minutes. • Selected, the plot does not update as the run progresses even after 10 minutes.

The Temperature Plot screen can be useful for identifying hardware failures. When monitoring the Temperature Plot screen, observe the Sample and Block plots for abnormal behavior.

- The Sample and Block plots should mirror each other approximately. A significant deviation of the plots may indicate a problem.
- The Cover plot should maintain the constant temperature specified in the method. A departure from the constant temperature may indicate a problem.

  Note: If you notice abnormal temperatures, refer to the instrument user guide to troubleshoot the error.

To monitor the Run Method

To view data in the Run Method screen, click **Run Method** from the Run Experiment Menu.

The Run Method screen displays the run method selected for the run in progress. The software updates the Run Status field throughout the run.

To...	Action
Change the number of cycles	In the **Adjust # of Cycles** field, enter the number of cycles to apply to the Cycling Stage.
Add a melt curve stage to the end of the run	Select **Add Melt Curve Stage to End**.
Add a Hold stage to the end of the run	Select **Add Holding Stage to End**.

To...	Action
Apply your changes	Click **Send to Instrument**.

If an alert appears, click the error for more information and troubleshoot the problem as explained in the QuantStudio™ 6 and 7 Flex Software Help (click or press **F1**).

Editing the run method during a run

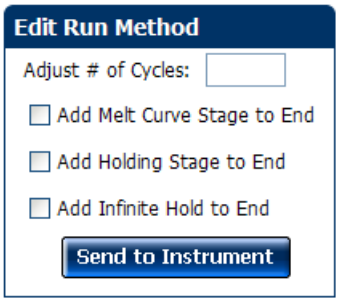
You can edit the run method while an experiment run is in progress on the Run Method screen from the Setup menu.

1. Increase or decrease the number of cycles by entering the cycle number in the Adjust # of Cycles box.

   Note: Ensure that you select the stage for which you want to increase or decrease the number of cycles in the graphical view of the run method. The Adjust # of Cycles box appears disabled if the corresponding stage is not selected.

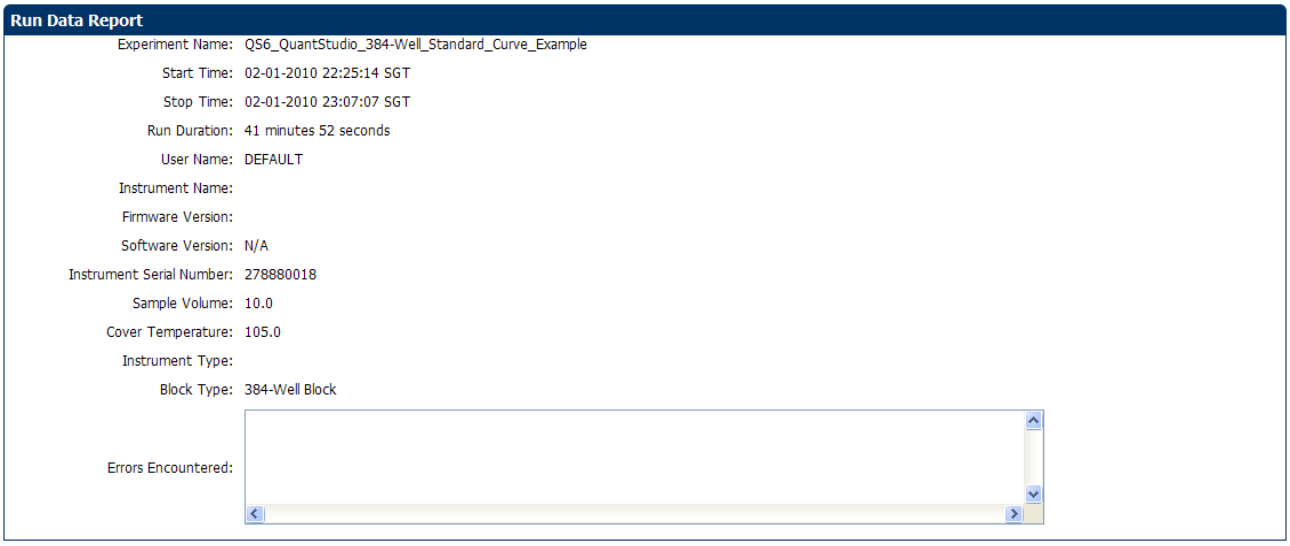
2. Select the appropriate check box to add a melt curve stage, holding stage, or infinite hold stage respectively, to the end of the run.

3. Click **Send to Instrument**.



To view the run data

After a run is complete, you can view a run report by clicking **View Run Data**. The View Run Data screen displays information about the completed run, as in the following image from a Standard Curve experiment:



The run report data helps in:

- Comparing two experiments of the same type run on two different instruments.
- Troubleshooting. For example, after a firmware upgrade, you can compare an experiment run before and after the upgrade to determine if the upgrade affected performance.

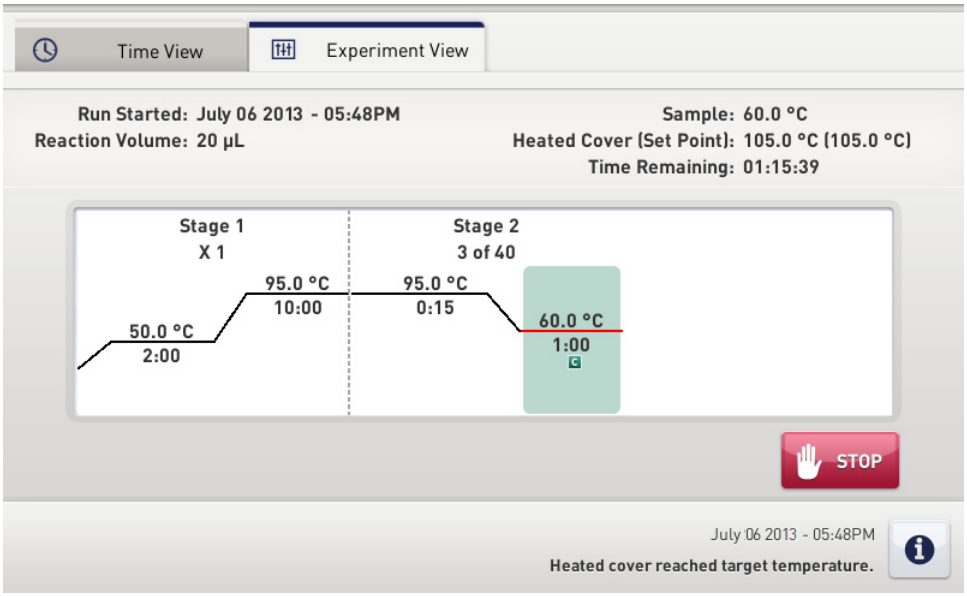
From the QuantStudio™ 6 or 7 Instrument touchscreen

The touchscreen displays the method for the experiment, the date and time at which the run started, the time remaining in the run, and other information.

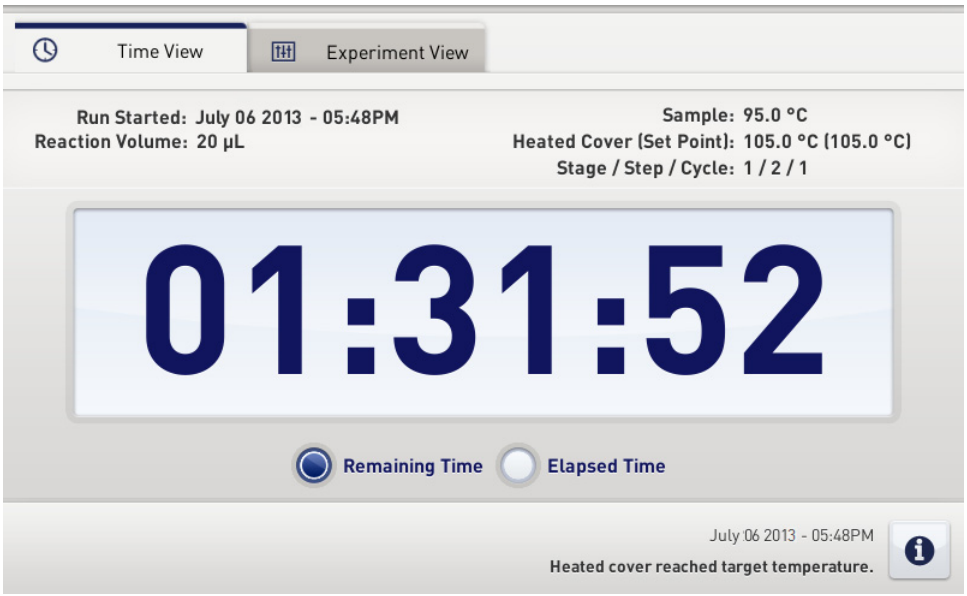
To...	Action
Display the time elapsed and the time remaining in the run	Touch the **Time View tab**, then touch **Experiment View** tab to return to the Run Method screen.
Stop the run	Touch STOP **STOP** to stop the protocol run immediately.
View the Events Log	Touch to view the list of run events that occurred during the run. Touch again to close the event list.

The run method on the QuantStudio™ 6 or 7 Instrument touchscreen looks like this:

Experiment View



Time View



Unload the instrument

When your QuantStudio™ 6 or 7 Instrument displays the Main Menu screen, unload the reaction plate from the instrument and transfer the experiment data to the computer for analysis.

Unload the reaction plate or array card

CAUTION! PHYSICAL INJURY HAZARD. During instrument operation, the temperature of the sample block(s) can exceed 100°C. Allow the consumable to cool to room temperature before removing.

When the QuantStudio™ 6 or 7 Instrument displays the Main Menu screen, you can unload the plate or array card as follows:

1. After the run, touch  on the QuantStudio™ 6 or 7 Instrument touchscreen or click **Open Door** in the Instrument Console screen of the QuantStudio™ 6 and 7 Flex Software to eject the plate or array card.
2. Remove the reaction plate or array card from the instrument tray and dispose of it according to your laboratory regulations.
3. Touch  on the QuantStudio™ 6 or 7 Instrument touchscreen or click **Close Door** to retract the plate adapter back into the instrument.

   If the QuantStudio™ 6 or 7 Instrument does not eject the plate, remove the plate as follows:

   a. Power off the QuantStudio™ 6 or 7 Instrument.
   b. Wait for 15 minutes, then power on the QuantStudio™ 6 or 7 Instrument and eject the plate.
   c. If the plate does not eject, power off and unplug the QuantStudio™ 6 or 7 Instrument, then open the access door.
   d. Wearing powder-free gloves, reach into the QuantStudio™ 6 or 7 Instrument and remove the plate from the heated cover, then close the access door.

Transfer experiment results

You can transfer the experiment results in either of the following two ways:

Download the experiment from the QuantStudio™ 6 or 7 Instrument over the network

When the QuantStudio™ 6 or 7 Instrument completes a experiment without a connection to the QuantStudio™ 6 and 7 Flex Software, the software allows you to download the results from the instrument through the network connection.

1. In the QuantStudio™ 6 and 7 Flex Software, select **Instrument** ▸ **Instrument Console**.
2. Select the instrument icon of the QuantStudio™ 6 or 7 Instrument from the My Instruments list, then click Manage Instrument to open the Instrument Manager.

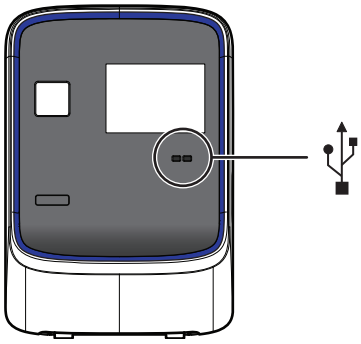
   Note: If the Manage Instrument button is inactive, add your QuantStudio™ 6 or 7 Instrument to the My Instruments group as explained in "Add the instrument to the My Instruments group" on page 26.
3. From the Instrument Manager, click **Manage Files**, then click **File Manager**.
4. From the File Manager screen, download the file(s):

   a. From the Folders field, select the folder that contains the files that you want to download.
   b. From the Experiments field, select the files to download. To select multiple files, **Ctrl-click** or **Shift-click** files in the list.
   c. From the Folders field, select the folder that contains the files that you want to download.
5. From the Save dialog box, select the folder to hold the experiment results and click **Save**. The experiments folder is located at:

C:\Applied Biosystems\QuantStudio 6 and 7 Flex Software\User Files\experiments

Transfer the experiment from the QuantStudio™ 6 or 7 Instrument to the computer via a USB drive:

1. Plug a USB drive into the USB port below the touchscreen.



IMPORTANT! Do not use the USB ports on the rear panel of the QuantStudio™ 6 or 7 Instrument. The rear USB ports are only for use by Life Technologies personnel to service the instrument

2. Touch the QuantStudio™ 6 or 7 Instrument touchscreen, to awaken it.
3. If the touchscreen is not at the Main Menu screen, touch  .
4. From the Main Menu of the QuantStudio™ 6 or 7 Instrument touchscreen, touch  **Collect Results** to save the data to the USB drive.
5. Select one or multiple experiments (by touching them). Then touch  **Save to USB** to copy selected experiments to the USB drive.

   Note: If your instrument cannot find the USB drive, remove the USB drive, then try again. If the instrument still does not recognize the USB drive, try another USB drive.
6. Touch  to return to the Main Menu.
7. Remove the USB drive from your instrument, then connect it to one of the USB ports on your computer.
8. In the computer desktop, use the Windows explorer to open the USB drive.
9. Copy the example experiment file to:

   C:\Applied Biosystems\QuantStudio 6 and 7 Flex Software\User Files\experiments

Review experiment results

About analysis results

Immediately after a run, the QuantStudio™ 6 and 7 Flex Software automatically analyzes the data using the default analysis settings, then displays the Amplification Plot screen.

Note: For auto-analysis of data, after a run, go to **Tools** ▸ **Preferences** ▸ **Experiment** and select the **Auto Analysis** check box.

To reanalyze the data, select all the wells in the plate layout, then click **Analyze**.

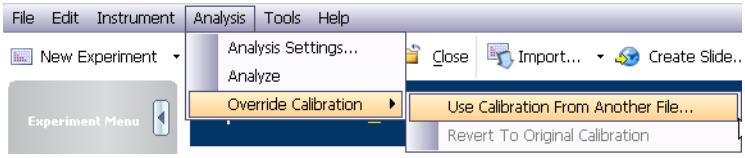
To override calibration

Each experiment file (.eds) stores the calibration data from the QuantStudio™ 6 or 7 Instrument it was run on. The calibration data can affect the analysis results of an experiment.

If you have run multiple experiments on different QuantStudio™ 6 or 7 Instruments and prefer the analysis results from a particular instrument, then you can choose to use the calibration data from another QuantStudio™ 6 or 7 Instrument.

To use the calibration data of another experiment

1. Open the experiment file (.eds), in which you want to import the calibration data from another QuantStudio™ 6 or 7 Instrument, in the QuantStudio™ 6 and 7 Flex Software.
2. Go to **Analysis** ▸ **Override Calibration** ▸ **Use Calibration From Another File....**



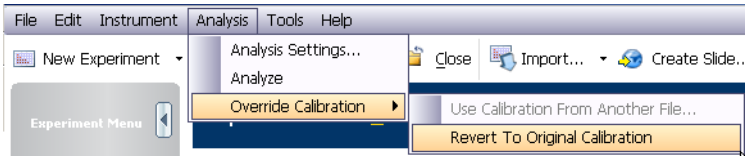
3. Browse to experiment file (.eds) from which you want to use the calibration data.

   Note: You can choose to override the calibration data in an experiment with the calibration data of any other experiment type; however the calibration data being used must be from the same instrument type, QuantStudio™ 6 Instrument or the QuantStudio™ 7 Instrument. Calibration data from an experiment in the QuantStudio™ 7 Instrument can be used to override calibration data of an experiment in the QuantStudio™ 6 Instrument, but not vice-versa.

4. Click **Open**.

To revert to the original calibration data

1. Open the experiment file (.eds), in which you want to import the original calibration data, in the QuantStudio™ 6 and 7 Flex Software.
2. Go to **Analysis** ▸ **Override Calibration** ▸ **Revert To Original Calibration.**



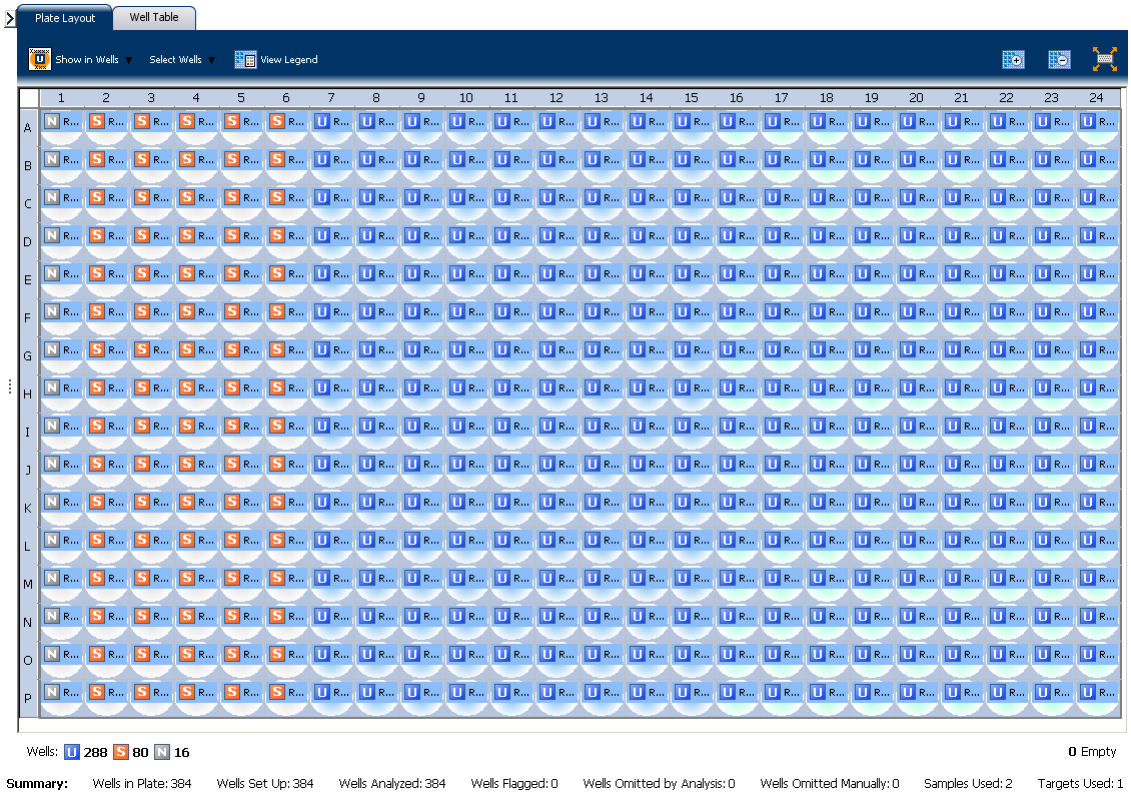
The experiment file will display analysis results as per the calibration data of the QuantStudio™ 6 or 7 Instrument that the experiment was run on.

To display wells

To display specific wells in the analysis plots, select the wells in the Plate Layout tab:

- To select wells of a specific type, use the Select Wells With drop-down menus: Select **Sample**, **Target**, or **Task**, then select the sample, target, or task name.
- To select a single well, click the well in the plate layout.
- To select multiple wells, click and drag over the desired wells, press **Ctrl-click**, or press **Shift-click** in the plate layout.
- To select all the wells, click the upper left corner of the plate layout.

The following is an image of the plate layout for a Standard Curve experiment:

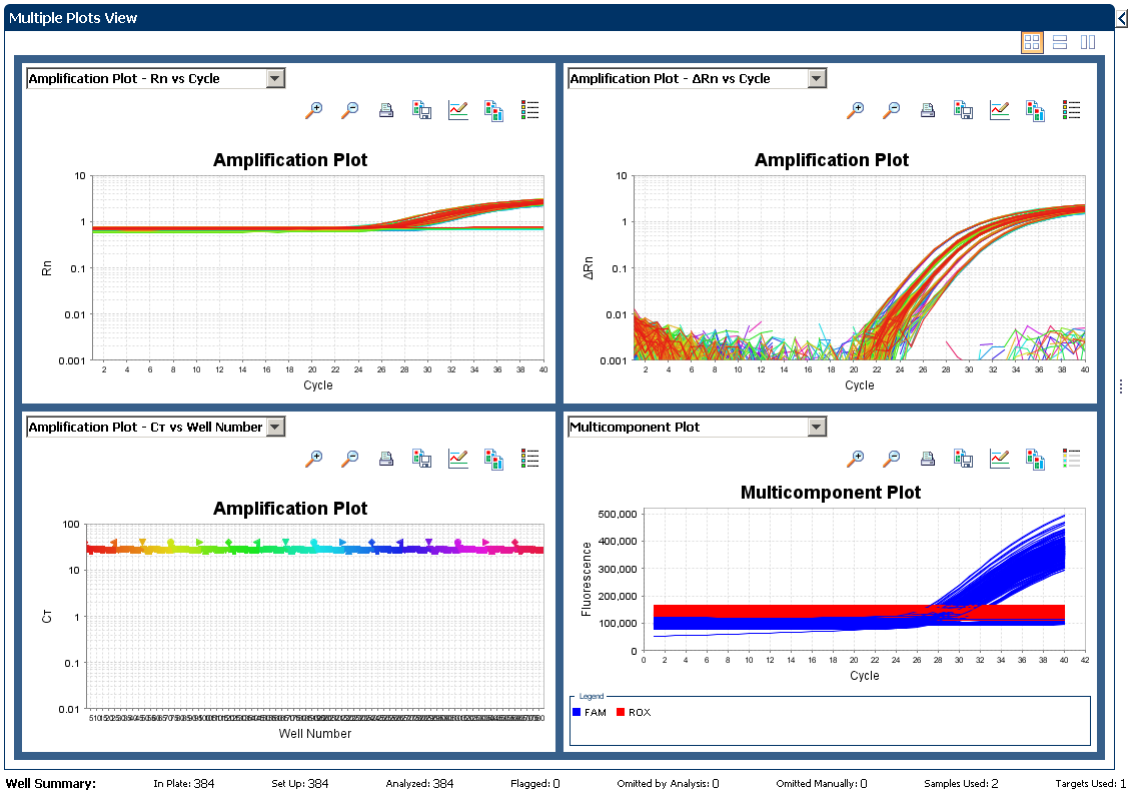


To display multiple plots

Use the Multiple Plots View screen to display up to four plots simultaneously. To navigate within the Multiple Plots View screen, from the Experiment Menu pane, select **Analysis ▸ Multiple Plots View**.

- To display four plots, click **Show plots in a 2 × 2 matrix**.
- Similarly, to display two plots in rows, click **Show plots in two rows**. and to display two plots vertically, click **Show plots in two columns**.
- To display a specific plot, select the plot from the drop-down menu above each plot display.

The following is an image of the Multiple Plots View screen for a Standard Curve example experiment:



To display an expanded view of a plot or wells

- Click  to expand the view of a plot, displayed on the left-hand side of the screen.
- Click  to expand the view of the Plate Layout or Well Table displayed on the right-hand side of the screen.

To edit plot properties

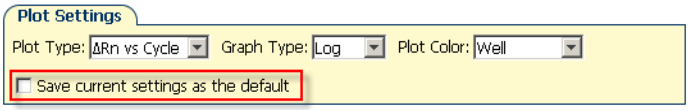
Use the Plot Properties dialog box on the Analysis screen to edit plot settings such as the font and color of the plot text, and the labels on the X axis and Y Axis.

1. Click  on the Analyze screen (the icon appears above the plot) to open the Plot Properties dialog box
2. Edit the settings under the General, X Axis, and Y Axis tab.
   - Click the General tab to edit the plot title text, font, or color. You can also select whether to show the plot title.
   - Click the X Axis tab to edit the x axis label text, font, or color; select the tick marks and tick mark labels to display; and select the range to display.
   - Click the Y Axis tab to edit the y axis label text, font, or color; select the tick marks and tick mark labels to display; and select the range to display.
3. Click **OK**.

To save current settings as default

You can change the Plot Settings for the different analysis plots, and save them as defaults.

Select the **Save current settings as the default** check box on the respective plot screens under the Analysis Experiment Menu.



To publish the analyzed data

To...	Click
Save a plot as an image file	
Print a plot	
Copy a plot to the clipboard	
Print a report	
Export data	

To...	Go to	Then
Print the plate layout	**File** ▸ **Print...**	Select the background color, and click **Print**
Create slides	**File** ▸ **Send to PowerPoint...**	Select the slides for your presentation, and click **Create Slides**
Print a report	**File** ▸ **Print Report...**	Select data for the report, and click **Print Report**

Export an experiment

About exporting an experiment

The Export feature of QuantStudio™ 6 and 7 Flex Software allows you to export:

Data type	Description
Plate setup files for future experiments	Plate setup files contain setup information such as the well number, sample name, sample color, target name, dyes, and other reaction plate contents.

Data type	Description
Analyzed data in different formats for further analysis	The data can be exported in the QuantStudio™ 6 and 7 format, QuantStudio™ Dx/ ViiA™ 7 format, the 7900 SDS format, and the RDML format. • The 7900 format is applicable only to Standard Curve, Relative Standard Curve, Genotyping, Presence/Absence, and Melt Curve experiments. • The RDML export format is applicable only to Standard Curve, Relative Standard Curve, Comparative C_T, and Melt Curve experiments. The RDML format is available only in a single file format. • For Standard Curve experiments, you can also export the analyzed data from the QuantStudio™ 6 and 7 Flex Software to the external applications, TaqMan® Protein Expression Data Analysis Software and CopyCaller® Software if they are installed on your computer before the QuantStudio™ 6 and 7 Flex Software is installed. The applications appear in the Tools menu.
Gene Expression studies	These are used to carry out a comparative analysis.

Export procedure

1. Open the experiment file that contains the data to export, and from the Experiment Menu, click  **Export.**

   Note: If you want the data to be exported automatically after analysis, select the **Auto Export** check box during experiment setup or before running an experiment.

2. Select the format for exported data:
   - **QuantStudio™ 6 and 7 Format** - Supports .txt, .xls, and .xlsx data.
   - **QuantStudio™ Dx/ ViiA™ 7 Format** - Supports .txt, .xls, and .xlsx data.
   - **7900 Format** - Supports only .txt data, where:
     - Single experiments are exported in the SDS 2.4 detector centric export format of the 7900 Sequence Detection System.
     - Studies are exported in the SDS 2.3 RQ manager detector centric export format of the 7900 Sequence Detection System
   - **RDML Format** - RDML (Real Time Data Markup Language) - Supports only .xml type of data.

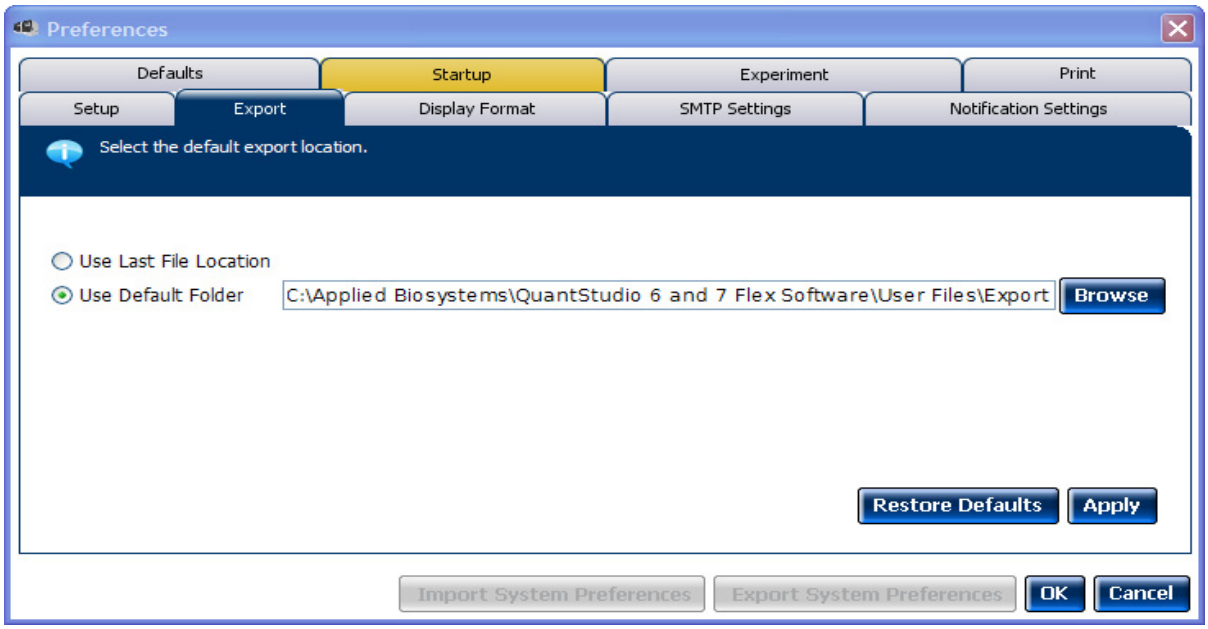
3. Select to export all data in one file or in separate files for each data type.
   - **One File** - All data types are exported in one file.
     - If you select the *.xls format, a worksheet is created for each data type.
     - If you select the *.txt format, the data are grouped by data type.
   - **Separate Files** - Each data type is exported in a separate file. For example, if you select three different data types Results, Amplification, and Multicomponent to export, three separate files (one each for Results, Amplification, and Multicomponent) are created. You can select the type of file (*.xls, *.xlsx or *.txt) to export from the **File Type** drop-down menu.

     Note: You cannot use an exported ***.xls** or an ***.xlsx** file when importing plate setup information.

4. *(Optional)* Select the **Open file(s) when export is complet**e check box to automatically open the file when export is complete.

5. Enter a file name and location.
   a. Enter a name for the export file in the **Export File Name** field.
   b. Enter the **Export File Location**. Click **Browse** if you do not want to save the export file in the default export folder.

      Note: To set up the Export File Location, go to **Tools** ▸ **Preferences**, and select the **Export** tab. You can select the **Use Last File Location** or **Use Default Folder** check box.



6. Select the data to export:

Select...	To export...
Sample setup	Well, sample name, sample color, and target name of samples in the plate
Raw data	Raw fluorescence data for each filter, for each cycle
Amplification data	Amplification results, such as C_T values, Rn, or ΔRn

Select...	To export...
Multicomponent data	Fluorescence data for each dye, for each cycle
Results	Results information, such as C_T values, Rn, or calls
Technical Replicate Results (Tech. Rep. Results)	Technical replicates information, such as Sample name, Target name, Task, or RQ
Biological Replicate Results (Bio. Rep. Results)	Biological replicates information, such as Biogroup name, Target name, Task, or RQ
Clipped Data	Information that is unique to the 7900 format. Data from the last three raw data points per step (clipped from the rest). The three data points are averaged to give you the final fluorescence data value for each step.
Reagent Information	Information about the reagent selected for the experiment

Note: Results data are not available for export until the run status is complete and the data are analyzed.

Note: The Technical Replicate Results, Biological Replicates Results, and Clipped Data are available only in Relative Standard Curve and Comparative C_T experiments.

7. (*Optional*) For Standard Curve experiments, select the external application, **TaqMan® Protein Expression Data Analysis Software** or **CopyCaller® Software** if either Software is installed on your computer.

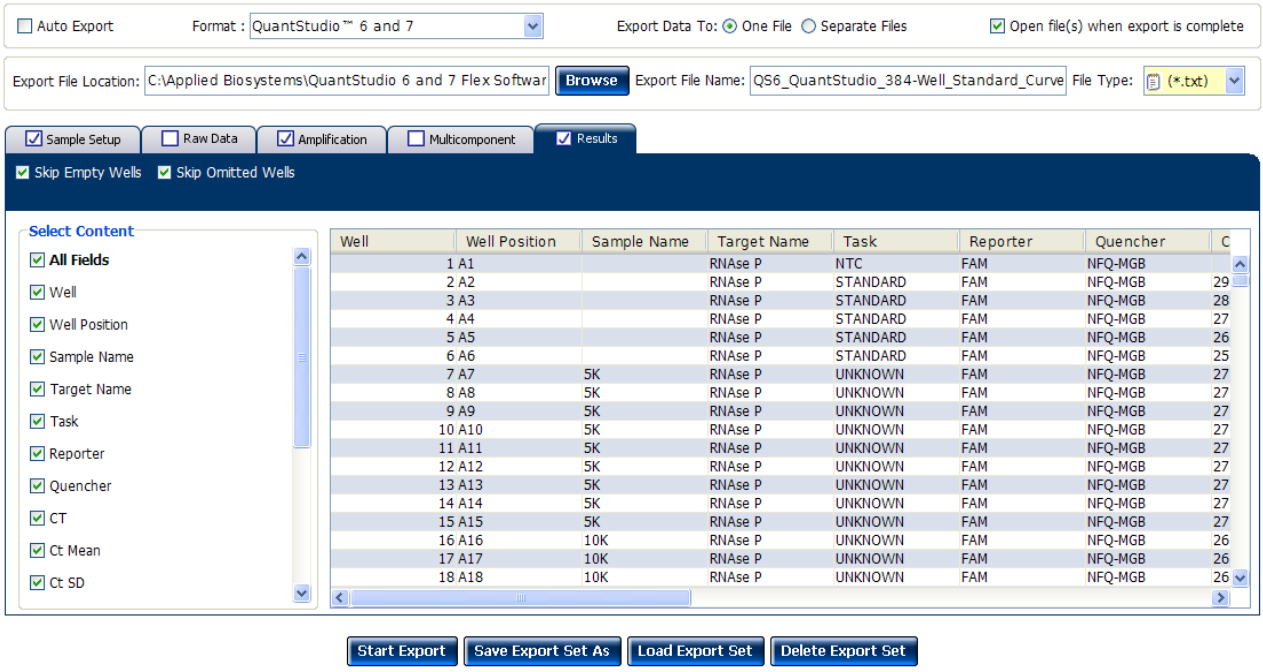
   Note: For more information on the TaqMan® Protein Expression Data Analysis Software or CopyCaller® Software, contact Life Technologies.

8. *(Optional)* After you have defined the export properties or after moving the table headings order, you can save those export settings as an export set by clicking **Save Export Set As**. Later you can import the heading order into another file by clicking **Load Export Set**.

   Note: It is advisable to keep the default order of the table headings if you are using the external Life Technologies applications, **TaqMan® Protein Expression Data Analysis Software** or **CopyCaller® Software** for further analysis.

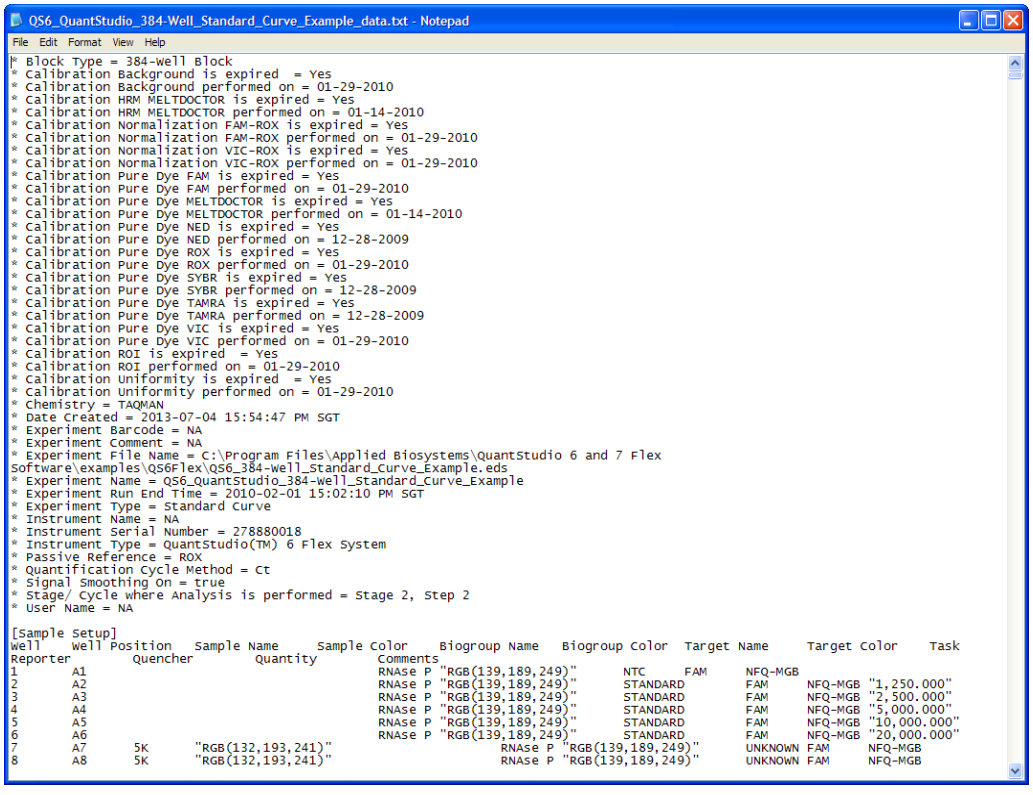
9. Click **Start Export**.

The following is an image of the Export screen for a Standard Curve experiment:



Well	Well Position	Sample Name	Target Name	Task	Reporter	Quencher	C
1	A1		RNAse P	NTC	FAM	NFQ-MGB	
2	A2		RNAse P	STANDARD	FAM	NFQ-MGB	29
3	A3		RNAse P	STANDARD	FAM	NFQ-MGB	28
4	A4		RNAse P	STANDARD	FAM	NFQ-MGB	27
5	A5		RNAse P	STANDARD	FAM	NFQ-MGB	26
6	A6		RNAse P	STANDARD	FAM	NFQ-MGB	25
7	A7	5K	RNAse P	UNKNOWN	FAM	NFQ-MGB	27
8	A8	5K	RNAse P	UNKNOWN	FAM	NFQ-MGB	27
9	A9	5K	RNAse P	UNKNOWN	FAM	NFQ-MGB	27
10	A10	5K	RNAse P	UNKNOWN	FAM	NFQ-MGB	27
11	A11	5K	RNAse P	UNKNOWN	FAM	NFQ-MGB	27
12	A12	5K	RNAse P	UNKNOWN	FAM	NFQ-MGB	27
13	A13	5K	RNAse P	UNKNOWN	FAM	NFQ-MGB	27
14	A14	5K	RNAse P	UNKNOWN	FAM	NFQ-MGB	27
15	A15	5K	RNAse P	UNKNOWN	FAM	NFQ-MGB	27
16	A16	10K	RNAse P	UNKNOWN	FAM	NFQ-MGB	26
17	A17	10K	RNAse P	UNKNOWN	FAM	NFQ-MGB	26
18	A18	10K	RNAse P	UNKNOWN	FAM	NFQ-MGB	26

The following is an image of the exported file when opened in Notepad:



QS6_QuantStudio_384-Well_Standard_Curve_Example_data.txt - Notepad

File Edit Format View Help

```
* Block Type = 384-well Block
* Calibration Background is expired  = Yes
* Calibration Background performed on = 01-29-2010
* Calibration HRM MELTDOCTOR is expired = Yes
* Calibration HRM MELTDOCTOR performed on = 01-14-2010
* Calibration Normalization FAM-ROX is expired = Yes
* Calibration Normalization FAM-ROX performed on = 01-29-2010
* Calibration Normalization VIC-ROX is expired = Yes
* Calibration Normalization VIC-ROX performed on = 01-29-2010
* Calibration Pure Dye FAM is expired = Yes
* Calibration Pure Dye FAM performed on = 01-29-2010
* Calibration Pure Dye MELTDOCTOR is expired = Yes
* Calibration Pure Dye MELTDOCTOR performed on = 01-14-2010
* Calibration Pure Dye NED is expired = Yes
* Calibration Pure Dye NED performed on = 12-28-2009
* Calibration Pure Dye ROX is expired = Yes
* Calibration Pure Dye ROX performed on = 01-29-2010
* Calibration Pure Dye SYBR is expired = Yes
* Calibration Pure Dye SYBR performed on = 12-28-2009
* Calibration Pure Dye TAMRA is expired = Yes
* Calibration Pure Dye TAMRA performed on = 12-28-2009
* Calibration Pure Dye VIC is expired = Yes
* Calibration Pure Dye VIC performed on = 01-29-2010
* Calibration ROI is expired  = Yes
* Calibration ROI performed on = 01-29-2010
* Calibration Uniformity is expired  = Yes
* Calibration Uniformity performed on = 01-29-2010
* Chemistry = TAQMAN
* Date Created = 2013-07-04 15:54:47 PM SGT
* Experiment Barcode = NA
* Experiment Comment = NA
* Experiment File Name = C:\Program Files\Applied Biosystems\QuantStudio 6 and 7 Flex
Software\examples\QS6Flex\QS6_384-Well_Standard_Curve_Example.eds
* Experiment Name = QS6_QuantStudio_384-Well_Standard_Curve_Example
* Experiment Run End Time = 2010-02-01 15:02:10 PM SGT
* Experiment Type = Standard Curve
* Instrument Name = NA
* Instrument Serial Number = 278880018
* Instrument Type = QuantStudio(TM) 6 Flex System
* Passive Reference = ROX
* Quantification Cycle Method = Ct
* Signal Smoothing On = true
* Stage/ Cycle where Analysis is performed = Stage 2, Step 2
* User Name = NA

[Sample Setup]
Well    Well Position   Sample Name     Sample Color    Biogroup Name   Biogroup Color  Target Name     Target Color    Task
Reporter        Quencher        Quantity        Comments
1       A1                                      RNAse P "RGB(139,189,249)"      NTC     FAM     NFQ-MGB
2       A2                                      RNAse P "RGB(139,189,249)"      STANDARD        FAM     NFQ-MGB "1,250.000"
3       A3                                      RNAse P "RGB(139,189,249)"      STANDARD        FAM     NFQ-MGB "2,500.000"
4       A4                                      RNAse P "RGB(139,189,249)"      STANDARD        FAM     NFQ-MGB "5,000.000"
5       A5                                      RNAse P "RGB(139,189,249)"      STANDARD        FAM     NFQ-MGB "10,000.000"
6       A6                                      RNAse P "RGB(139,189,249)"      STANDARD        FAM     NFQ-MGB "20,000.000"
7       A7      5K      "RGB(132,193,241)"                      RNAse P "RGB(139,189,249)"      UNKNOWN FAM     NFQ-MGB
8       A8      5K      "RGB(132,193,241)"                      RNAse P "RGB(139,189,249)"      UNKNOWN FAM     NFQ-MGB
```

2 Experiment Shortcuts

This chapter provides you with shortcuts to use in the QuantStudio™ 6 and 7 Flex Software after you have learned experiment basics.

You can reuse experiment settings and plate setup information by: directly importing and editing a template, using the QuickStart feature with a template, importing experiment setup information, or importing a sample definition file; you can also prepare several experiments at once or create a new experiment using the ReadiApp feature.

The chapter covers:

Using experiment templates

You can use a template (.edt) to create a new experiment. Templates are useful when you want to create many experiments with the same experiment parameters.

You can create an experiment from a template from the QuantStudio™ 6 and 7 Flex Software and from the QuantStudio™ 6 or 7 Instrument touchscreen.

Note: To access the QuantStudio™ 6 and 7 Flex Software example templates, navigate to the templates folder located at *<drive>*:\Program Files\Applied Biosystems\QuantStudio 6 and 7 Flex Software\examples\User Sample Files.

where, *<drive>* is the computer hard drive on which the QuantStudio™ 6 and 7 Flex Software is installed. The default installation drive for the software is the C: drive.

To create a template

1. Log in to the QuantStudio™ 6 and 7 Flex Software and, from the Home screen, open an existing experiment, or create a new experiment.

   Note: To create a new experiment using the Experiment Setup, see "Set up an experiment" on page 12.

2. Select **File ▸ Save As Template**.

   Note: The information saved in a template includes plate setup information (defined targets and samples, plate assignment of targets and samples), reagent information, thermal protocol, and analysis settings such as quantification cycling method.

3. Enter a file name, select a location for the template, then click **Save** and **Close**.

   You can use that experiment as a template for similar experiments.

To create a new experiment using a template

1. From the Home screen, click  **Template**.
2. Locate and select the template file, then click **Open**.
   A new experiment is created using the setup information from the template.
3. Confirm that the following are correct before you prepare the reactions and run the experiment:
   - Experiment properties (experiment name, experiment type, block type, reagent, run properties)
   - Plate definitions (targets, samples, and biological replicates)
   - Plate assignments (targets, samples, and biological replicates)
   - Run method (thermal protocol)
4. Proceed to preparing reactions, running the experiment, and analyzing the data.

To create an experiment using a template on the QuantStudio™ 6 or 7 Instrument touchscreen

You can run experiments using templates from the QuantStudio™ 6 or 7 Instrument touchscreen by importing the templates from the QuantStudio™ 6 and 7 Flex Software instrument console or a USB drive. You can also modify the experiment parameters in the templates as per your requirement.

To edit a template before running the experiment

1. Touch 

New on the View Templates screen to create a new experiment from the existing template.

   Note: Select a template before you touch **New**.
2. Edit the experiment parameters in the Create New Experiment screen.
3. Touch **Save & Exit** to save and exit the experiment or touch 

Save & Start Run to save and start an experiment run.

To run a pre-existing template

1. Touch 

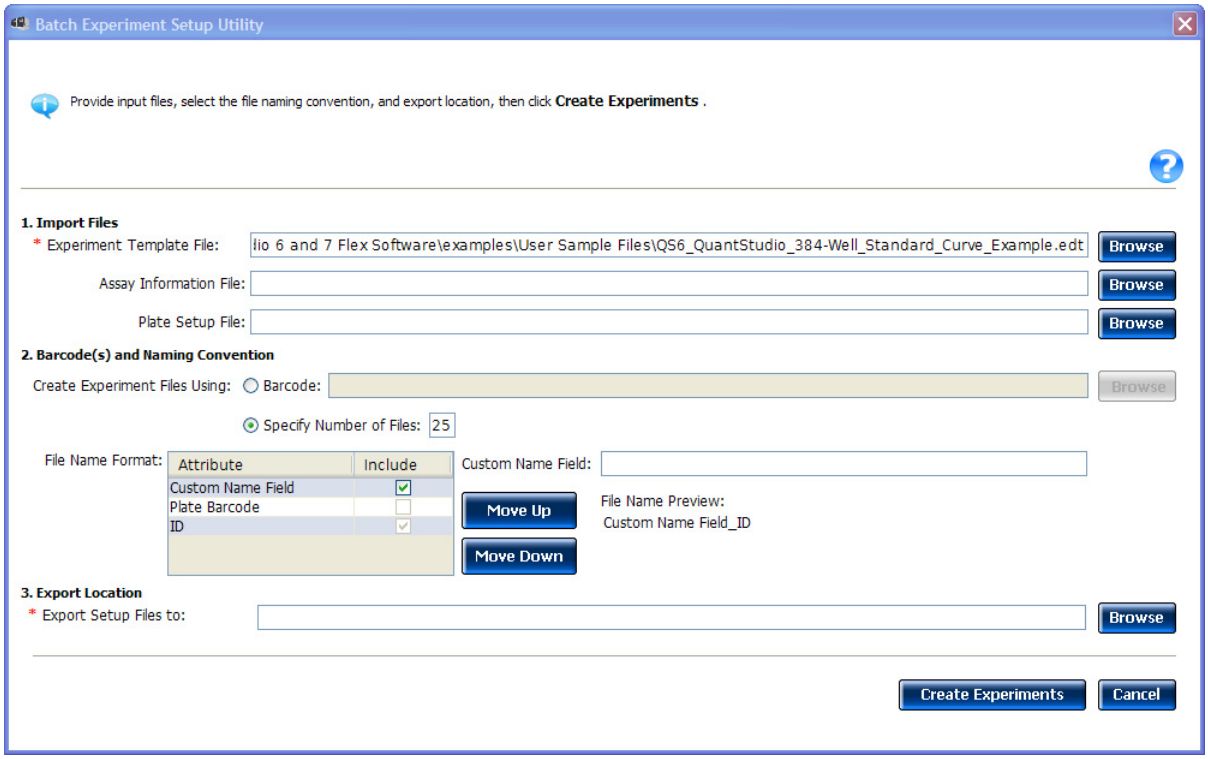
View Templates on the Home screen of the QuantStudio™ 6 or 7 Instrument touchscreen.
2. Select a pre-existing template from the templates list on the View Templates screen.
3. Touch 

View to see the run profile before you start a run.
4. After confirming the template setup is correct, touch  to go back to View Templates screen. Touch **Start Run**.

Use a template to create a batch of experiments

Use the batch experiment utility to create multiple experiment files from the same template without using Experiment Setup.

1. In the menu bar, select **Tools** ▸ **Batch Experiment Setup**. The following is an image of the Batch Experiment Setup Utility dialog box:



2. Select the file(s) to use to create the new experiments:
   a. Click **Browse** in the Experiment Template File field.

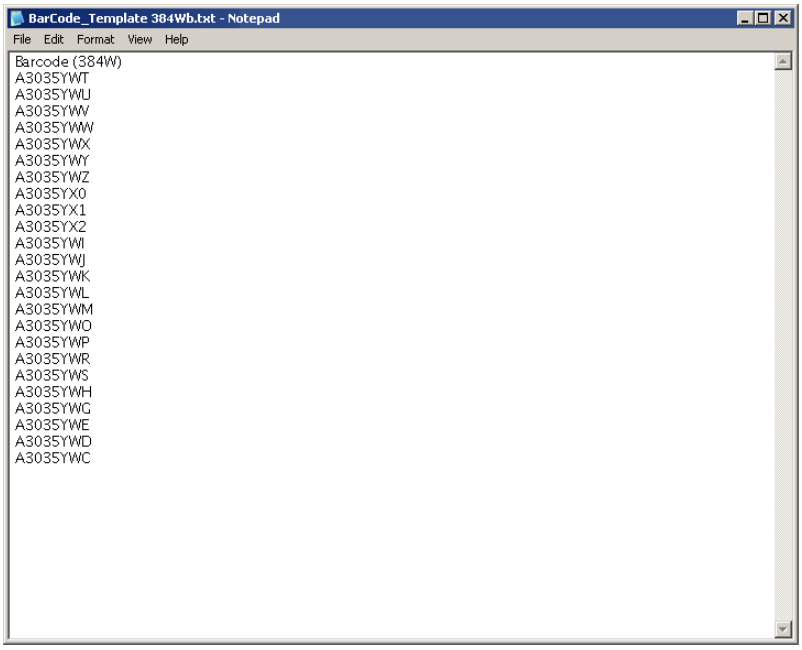
      Note: To use one of the example setup files, browse to <drive>:\Program Files\Applied Biosystems\QuantStudio 6 and 7 Flex Software\examples\User Sample Files

   b. Locate an *.edt file to import, then click **Select**.

   c. *(Optional)* Repeat **steps 2a** and **2b** for the remaining setup file types to import (Assay Information File (*.txt or *.xml), Plate Setup File (*.txt)).

3. Select the option to create experiment files. The selected option determines the number of experiment files created:
   - **Specify Number of Files** - Enter a number from 1 to 100.
   - **Barcode** - Click **Browse** and select a Barcode File (*.txt) to import. The software automatically adds the Plate Barcode attribute to the file name format. The number of experiments created equals the number of barcodes present in the barcode file.

     Note: A Barcode File contains one barcode per line. An example Barcode File looks like the following image:



4. *(Optional)* Edit the file name format. Use the File Name Preview to verify your settings.
   - Select the check box to include or exclude the **Custom Name Field** attribute from the file name. If included, click the Custom Name Field and enter up to 100 letters and/or numbers to identify the batch of experiments.

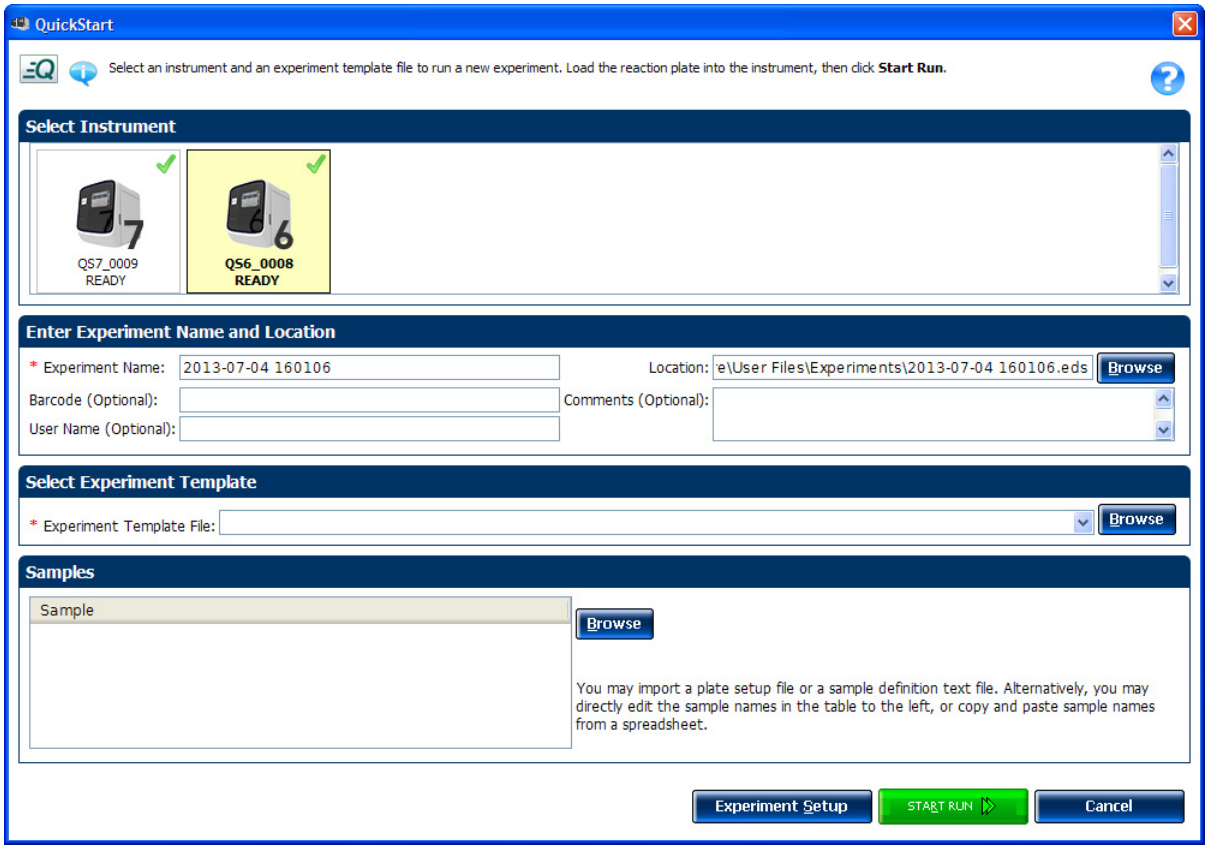
     Note: The file name can contain a total of 100 characters, including all file name attributes.
   - Click **Move Up** or **Move Down** to change the order of the selected file name attributes.
5. Select the location for the experiment files to be created:
   a. Click **Browse** in the Export Setup Files to: field.
   b. Review the location for the experiment files. Navigate to a new location if you do not want to export the experiment files to that folder, then click **Select**.
6. Click **Create Experiments**. A confirmation message appears when the batch of experiments has been created.

Run an experiment with QuickStart

You can use a template to run an experiment with the QuantStudio™ 6 and 7 Flex Software QuickStart feature:

QuickStart from the QuantStudio™ 6 and 7 Flex Software

1. Prepare the reactions.
2. Log in to the QuantStudio™ 6 and 7 Flex Software and, from the Home screen, click **QuickStart** to access the QuickStart dialog box.
3. In the QuickStart dialog box, enter or select the:
   a. Instrument icon of the instrument to perform the run on.
   b. Experiment name.
   c. Experiment template file.
   d. (*Optional*) Barcode and User Name for the experiment.
4. *(Optional)* To review the experiment or to make changes to any of the experiment parameters, click **Experiment Setup**.
   The following is an image of the QuickStart dialog box:



5. Proceed to running the experiment and analyzing the data.

QuickStart from the QuantStudio™ 6 or 7 Instrument touchscreen

You can QuickStart an experiment from the QuantStudio™ 6 or 7 Instrument touchscreen in the following ways:

- Start an experiment using a pre-defined template.
- Start an experiment with a pre-defined short-cut button.

Start an experiment using a pre-defined template

You can use a pre-existing template from the default experiments folder or use a custom template from another folder to start a run.

Start an experiment with a pre-defined short-cut button

The QuantStudio™ 6 or 7 Instrument touchscreen displays up to 18 shortcut buttons to templates or folders that contain experiments to be run. The shortcut buttons are present under MY SHORTCUTS on the Home screen. To start a run, touch any of the pre-defined experiment or folder buttons.

To create a shortcut button for a preferred experiment or a folder that contains experiments:

1. Touch 

Settings to open the Settings Menu.
2. Touch **Set Up Shortcuts** to list the Shortcut Targets.
3. On the Shortcut Targets list screen, select an existing template Shortcut Target button or an unused button.
4. Touch **Set Shortcut**. If you selected an unused button, then touching Set Shortcut will list out the templates and folders to set the shortcut for.
5. Under the  **From Templates** tab, select the templates for which you are creating the shortcut button.
6. *(Optional)* Create a shortcut button to show the templates or experiments in a particular folder for quick access, from those listed under the *From Folders* tab. You can touch *Edit* to create or edit shortcut buttons.

Import experiment setup

Import plate setup for an experiment

You can import the plate setup for a new experiment from an exported file with one of the following formats:

- *.txt - Text format
- *.xml - XML format
- *.csv - Comma separated values format
- *.eds - EDS file format
- *.edt - EDS template files format
- *.sdt - Sequence Detection System (SDS) template files format
- *.sds - 7900 v2.4 format

IMPORTANT! Make sure the file you select contains only plate setup data and that the experiment types match.

Note: For instructions on exporting an experiment, see "Export an experiment" on page 47.

To Import the plate setup data:

1. Create a new experiment or open an existing experiment.
2. In the Experiment Setup screen, select **File ▸ Import Plate Setup** or access the Import drop-down menu in the toolbar and select **Import Plate Setup**.
3. Click **Browse**, locate and select the file to import, then click **Select**.

   Note: To use one of the example setup files, browse to C:\Program Files\Applied Biosystems\QuantStudio 6 and 7 Flex Software\examples\User Sample Files

4. Click **Start Import**. The setup data from the exported text file is imported into the open experiment.

   Note: If your experiment already contains plate setup information, the software asks if you want to replace the plate setup with the data from the import file. Click **Yes** to replace the plate setup.
5. After importing plate setup information, use Experiment Setup to set up your experiment, and then run the experiment.

Note: You can import plate setup information from a 96-well plate into a 384-well plate, provided that the file you are importing the information from is a .txt file.

Import sample information

You can import sample information from a sample definition file to include in the plate setup for your experiment. A sample definition file is a comma-delimited file (*.csv) or a tab-delimited text file (*.txt) that contains the following setup information: well number, sample name, and custom sample properties.

Note: Make sure that the sample definition file you select contains only sample information.

Create a sample definition file

1. Open a text editing program such as Notepad.
2. Enter the following column headers in the first row (press the Tab key between each entry if you are saving the file as *.txt or enter a comma between each entry if you are saving the file as *.csv):
   - Well
   - Sample Name
   - *(Optional)* Column header names for up to 32 user-defined custom fields (for example, **Custom 1**, **Custom 2**, etc.)

3. For each subsequent row, enter the well number, press the **Tab** key or enter a comma, then enter the sample name. Optionally, press the **Tab** key, then enter the custom properties for the sample.

4. Save the file with the .txt or .csv file extension.

   An example sample definition, saved with the .csv extension, file looks like this:

	A	B	C	D	E	F	G	H
1	Well	Sample Name	ID	Age	Sex	Weight	HairColor	Smoker
2	1	Sample 1	1	22	Female	25	black	Yes
3	2	Sample 2	2	25	Male	26	brown	No
4	3	Sample 3	3	45	Female	50	blonde	Yes
5	4	Sample 4	4	31	Male	33	red	Yes
6	5	Sample 5	5	29	Female	46	grey	No
7	6	Sample 6	6	26	Male	35	black	No
8	7	Sample 7	7	31	Female	33	black	Yes
9	8	Sample 8	8	32	Male	67	black	No
10	9	Sample 9	9	32	Female	55	brown	Yes
11	10	Sample 10	10	33	Male	44	blonde	Yes
12	11	Sample 11	11	34	Female	25	red	No
13	12	Sample 12	12	34	Male	26	grey	No
14	13	Sample 13	13	35	Female	50	black	Yes
15	14	Sample 14	14	35	Male	33	black	No
16	15	Sample 15	15	36	Female	46	black	Yes
17	16	Sample 16	16	36	Male	35	brown	Yes
18	17	Sample 17	17	37	Female	33	blonde	No
19	18	Sample 18	18	37	Male	67	red	No
20	19	Sample 19	19	38	Female	55	grey	Yes
21	20	Sample 20	20	38	Male	44	black	No

Import sample information from a sample definition file

1. Create a new experiment or open the experiment to receive the setup data (select **File ▸ Open**, select the file to open, then click **Open**).

2. From the open experiment, select **File ▸ Import Plate Setup.**

3. Click **Browse** to browse your computer for a sample definition text file (*.csv). After you locate the file and select it, click **Select**.

   Note: To use one of the example setup files, browse to C:\Program Files\Applied Biosystems\QuantStudio 6 and 7 Flex Software\examples\User Sample Files

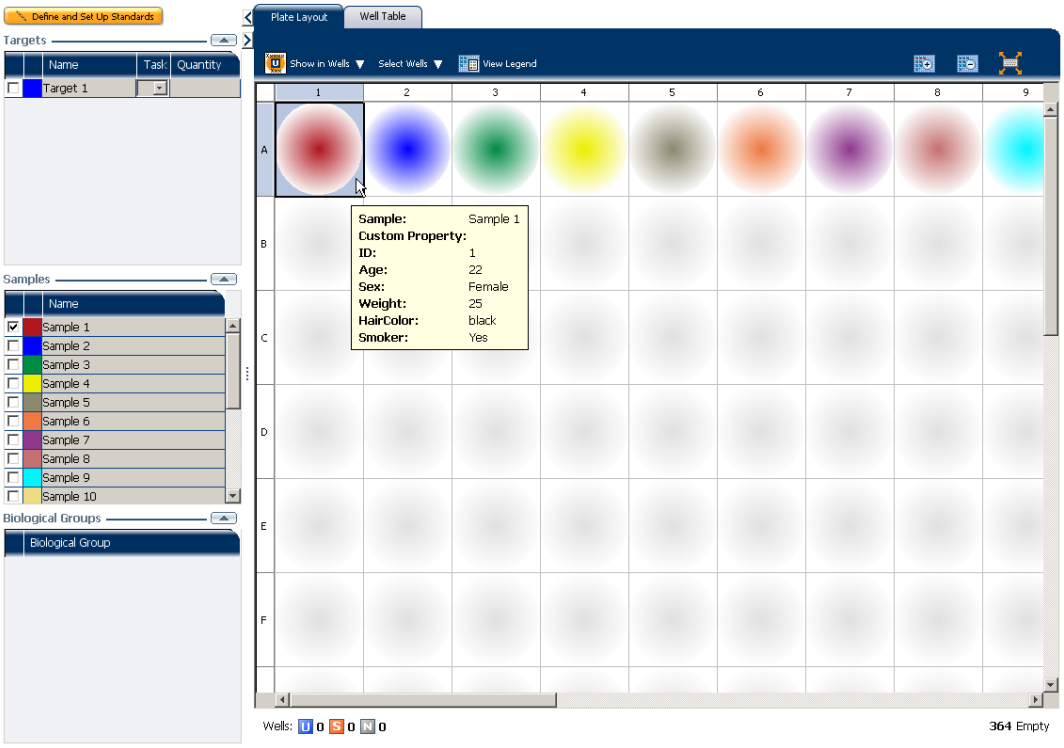
4. Click **Start Import**.

5. If your experiment already contains plate setup information, the software asks you if you want to replace the plate setup with the data from the file. Click **Yes** to replace the plate setup information.

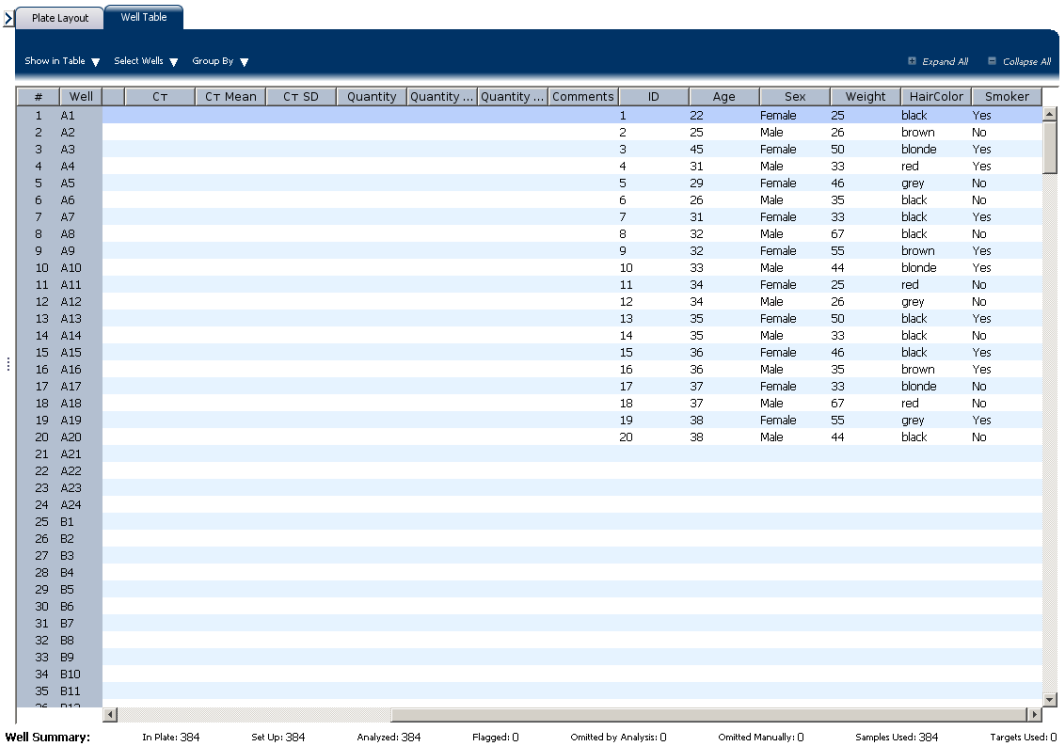
The samples appear in the Samples table for the experiment. All samples and well assignments in the experiment are replaced with those in the file. If defined, the custom sample properties also appear in the Well Table of the Analysis Section, and also in the Plate Layout tooltips in both the Setup and Analysis screens. The custom fields can be exported with the results data.

Note: You cannot edit the custom sample properties from within the Well Table. To modify this information, edit the custom fields in the sample definition file and import the file again. All of the sample information in the experiment is replaced with the information in the new file.

The following is an image of the Assign screen with information from the above sample definition file:



The following is an image of the Well Table in the Analysis section:



#	Well	CT	CT Mean	CT SD	Quantity	Quantity ...	Quantity ...	Comments	ID	Age	Sex	Weight	HairColor	Smoker
1	A1								1	22	Female	25	black	Yes
2	A2								2	25	Male	26	brown	No
3	A3								3	45	Female	50	blonde	Yes
4	A4								4	31	Male	33	red	Yes
5	A5								5	29	Female	46	grey	No
6	A6								6	26	Male	35	black	No
7	A7								7	31	Female	33	black	Yes
8	A8								8	32	Male	67	black	No
9	A9								9	32	Female	55	brown	Yes
10	A10								10	33	Male	44	blonde	Yes
11	A11								11	34	Female	25	red	No
12	A12								12	34	Male	26	grey	No
13	A13								13	35	Female	50	black	Yes
14	A14								14	35	Male	33	black	No
15	A15								15	36	Female	46	black	Yes
16	A16								16	36	Male	35	brown	Yes
17	A17								17	37	Female	33	blonde	No
18	A18								18	37	Male	67	red	No
19	A19								19	38	Female	55	grey	Yes
20	A20								20	38	Male	44	black	No
21	A21													
22	A22													
23	A23													
24	A24													
25	B1													
26	B2													
27	B3													
28	B4													
29	B5													
30	B6													
31	B7													
32	B8													
33	B9													
34	B10													
35	B11													

Create an experiment using ReadiApp

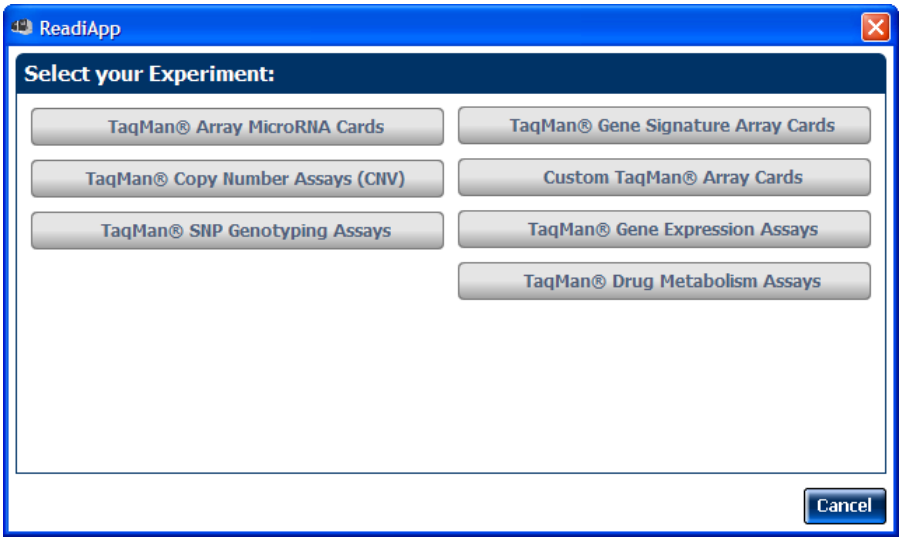
You can use the ReadiApp feature to set up an experiment in the QuantStudio™ 6 and 7 Flex Software. The ReadiApp feature provides a shortcut to create experiments for the assays purchased from Life Technologies.

The default ReadiApp templates available in the QuantStudio™ 6 and 7 Flex Software include:

- TaqMan® Gene Signature Array Cards
- Custom TaqMan® Array Cards
- TaqMan® Gene Expression Assays
- TaqMan® Drug Metabolism Assays
- TaqMan® Array MicroRNA Cards
- TaqMan® Copy Number Assays (CNV)
- TaqMan® SNP Genotyping Assays

1. Log in to the QuantStudio™ 6 and 7 Flex Software and, from the Set Up menu on the Home screen, click **ReadiApp**.
2. Click the assay to use to set up an experiment.

   Note: Click **Cancel** to exit the ReadiApp dialog box.



   A new experiment is created using the setup information from the template.
3. (*Optional*) Edit the experiment properties.
4. Proceed to preparing reactions, running the experiment, and analyzing the data.

Index

W

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lifetechnologies.com

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USER GUIDE

applied biosystems®
by life technologies™

QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Getting Started Guide for Standard Curve Experiments

Booklet 2

Publication Number 4489822
Revision A

life technologies™

Contents

1 About Standard Curve Experiments

IMPORTANT! First-time users of the QuantStudio™ 6 and 7 Flex Real-Time PCR System Software please read Booklet 1, *Getting Started with QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Experiments* and Booklet 7, *QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Experiments - Appendixes* of this binder thoroughly. The booklet provides information and general instructions that are applicable to all the experiments described in this binder

Note: For more information about any of the topics discussed in this guide, access the Help from within QuantStudio™ 6 and 7 Flex Real-Time PCR System Software by pressing F1, clicking in the toolbar, or selecting **Help ▸ QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Help**.

Before you begin

The Standard Curve method is used for determining absolute target quantity in samples. With the standard curve method, the software measures amplification of the target in samples and in a standard dilution series. Data from the standard dilution series are used to generate the standard curve. Using the standard curve, the software interpolates the absolute quantity of target in the samples.

Assemble required components

- **Sample** – The tissue group that you are testing for a target gene.
- **Standard** – A sample that contains known quantities of the target; used in quantification experiments to generate standard curves.
- **Standard dilution series** – A set of standards containing a range of known quantities. The standard dilution series is prepared by serially diluting standards.
- **Replicates** – The total number of identical reactions containing identical samples, components, and volumes.
- **Negative controls** – Wells that contain water or buffer instead of sample template. No amplification of the target should occur in negative control wells.

PCR Options

When performing real-time PCR, choose between:

- Singleplex and multiplex PCR (below)

  and

- 1-step and 2-step RT-PCR (page 6)

Singleplex and Multiplex PCR

You can perform a PCR reaction using either:

- **Singleplex PCR** – In singleplex PCR a single primer set is present in the reaction tube or well. Only one target or endogenous control can be amplified per reaction.

  or

- **Multiplex PCR** – In multiplex PCR, two or more primer sets are present in the reaction tube or well. Each set amplifies a specific target or endogenous control. Typically, a probe labeled with FAM™ dye detects the target and a probe labeled with VIC® dye detects the endogenous control.

IMPORTANT! SYBR® Green reagents cannot be used for multiplex PCR.

1- and 2-Step RT-PCR

You can perform reverse transcription (RT) and PCR in a single reaction (1-step) or in separate reactions (2-step). The reagent configuration you use depends on whether you are performing 1- or 2-step RT-PCR:

- **1-step RT-PCR**– In 1-step RT-PCR, RT and PCR take place in one buffer system. Using one buffer system provides the convenience of a single-tube preparation for RT and PCR amplification. However, you cannot use Fast PCR master mix or the carryover prevention enzyme, AmpErase® UNG (uracil-N-glycosylase), to perform 1-step RT-PCR.
- **2-step RT-PCR** – 2-step RT-PCR is performed in two separate reactions: First, total RNA is reverse-transcribed into cDNA, then the cDNA is amplified by PCR. This method is useful for detecting multiple transcripts from a single cDNA template or for storing cDNA aliquots for later use. The AmpErase® UNG enzyme can be used to prevent carryover contamination.

  Note: The Standard Curve example experiment is designed for singleplex PCR, where every well contains a primer/probe set for a single target; the reactions are set up for a 2-step RT-PCR.

About the example experiment

To illustrate how to perform Standard Curve experiments, this guide leads you through the process of designing and performing an example experiment. The example experiment represents a typical setup that you can use to quickly familiarize yourself with the QuantStudio™ 6 and 7 Flex Real-Time PCR System Software.

The objective of the Standard Curve example experiment is to determine the quantity of the RNase P gene in two populations.

In the standard curve example experiment:

- The samples are genomic DNA isolated from two populations.
- The target is the RNase P gene.
- One standard curve is set up for the RNase P gene (target). The standard used for the standard dilution series contains known quantities of the RNase P gene. Because a single target is being studied, only one standard curve is required.

  Note: In experiments where multiple targets are being studied, a standard curve is required for each target.
- Three replicates of each sample and each dilution point in the standard curve are performed to ensure statistical significance.
- The experiment is designed for singleplex PCR, where every well contains a primer/probe set for a single target.
- Reactions are set up for 2-step RT-PCR.
- Primer/probe sets are from Life Technologies RNase P assay.

  Note: The human RNase P FAM™ dye-labeled MGB probe is not available as a TaqMan® Gene Expression Assay. It can be ordered as a Custom TaqMan® Gene Expression Assay (Part no. 4331348).

2 Design the Experiment

This chapter explains how to design the example experiment from the Experiment Setup menu.

This chapter covers:

Note: To automatically export the analyzed data to a specified location, select the Auto Export task from the Export screen, before running the experiment. For more information on Auto Export, refer to Chapter 2 in Booklet 1, *Getting Started with QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Experiments*.

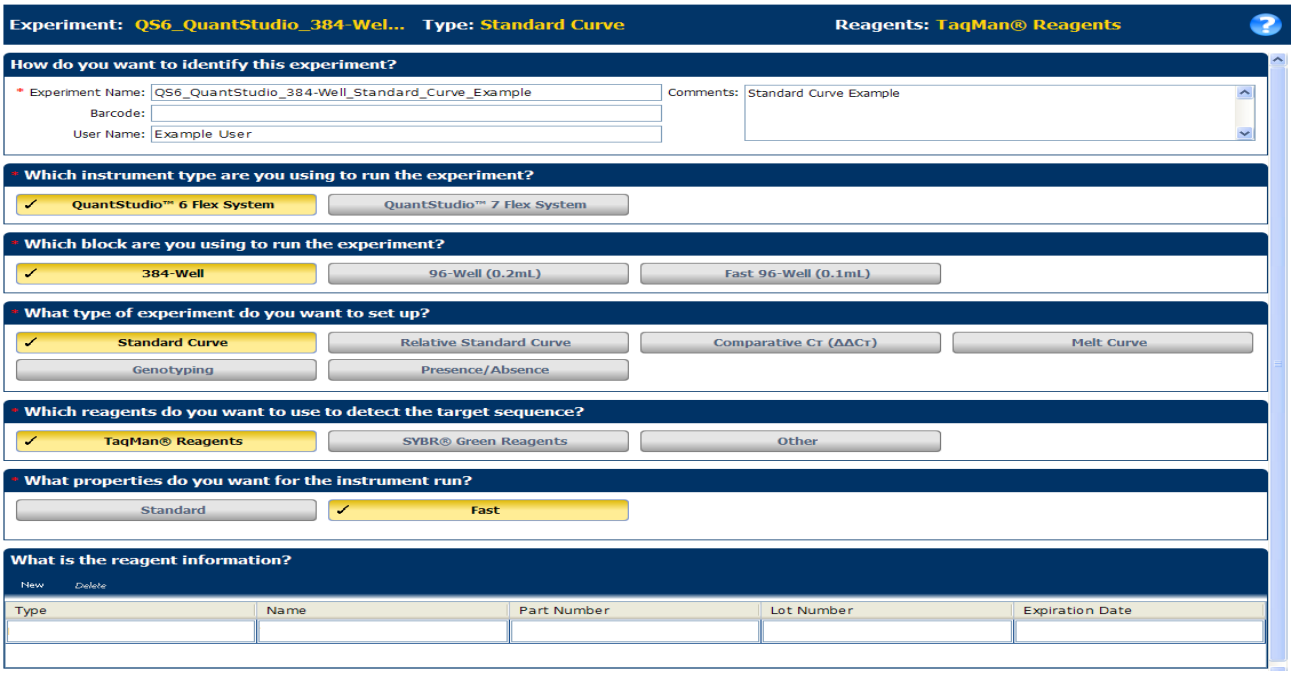
Define the experiment properties

Click **Experiment Setup** ▸ **Experiment Properties** to create a new experiment in the QuantStudio™ 6 and 7 Flex Software. Enter:

Field or selection	Entry
Experiment Name	QS6_QuantStudio_384-Well_Standard_Curve_Example
Barcode	Leave field empty
User Name	Example User
Comments	Standard Curve example
Instrument type	QuantStudio™ 6 Flex System
Block	384-Well Block
Experiment Type	Standard Curve
Reagents	TaqMan® Reagents
Ramp speed	Fast
Reagent Information	NA

Save the experiment.

Your Experiment Properties screen should look like this:



Define targets, samples, and biological replicates

Click **Define** to access the Define screen. Enter:

1. Targets

Target name	Reporter	Quencher	Color
RNaseP	FAM	NFQ-MGB	

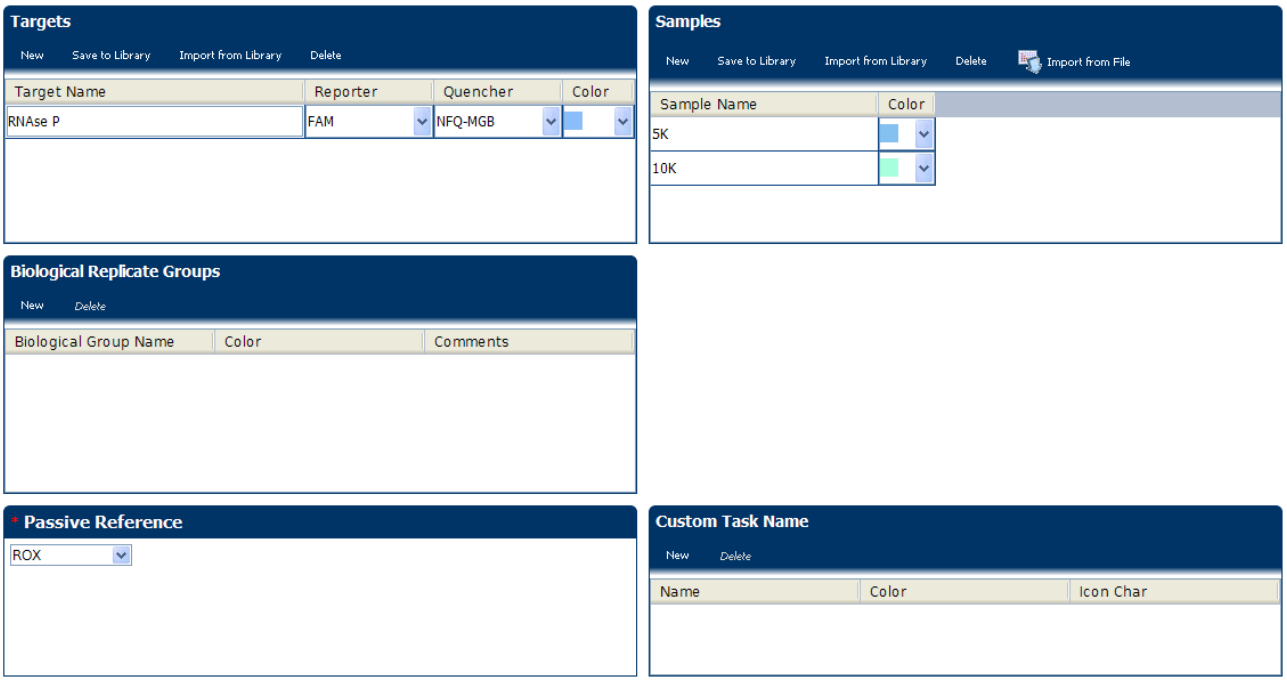
2. Samples

Sample name	Color
5K	
10K	

3. Dye to be used as a Passive Reference
   ROX

4. Custom Task Name
   Not applicable

Your Define screen should look like this:



Note: This example experiment does not define biological replicate groups. Leave Biological Replicate Groups blank.

Assign targets, samples, and biological groups

Click **Assign** to access the Assign screen.

1. Define and set up standards.
    a. Click **Define and Set Up Standards** on the Assign screen.
    b. Select a target.

Field	Select
Select a target for this standard curve	RNaseP

    c. Define the standard curve.

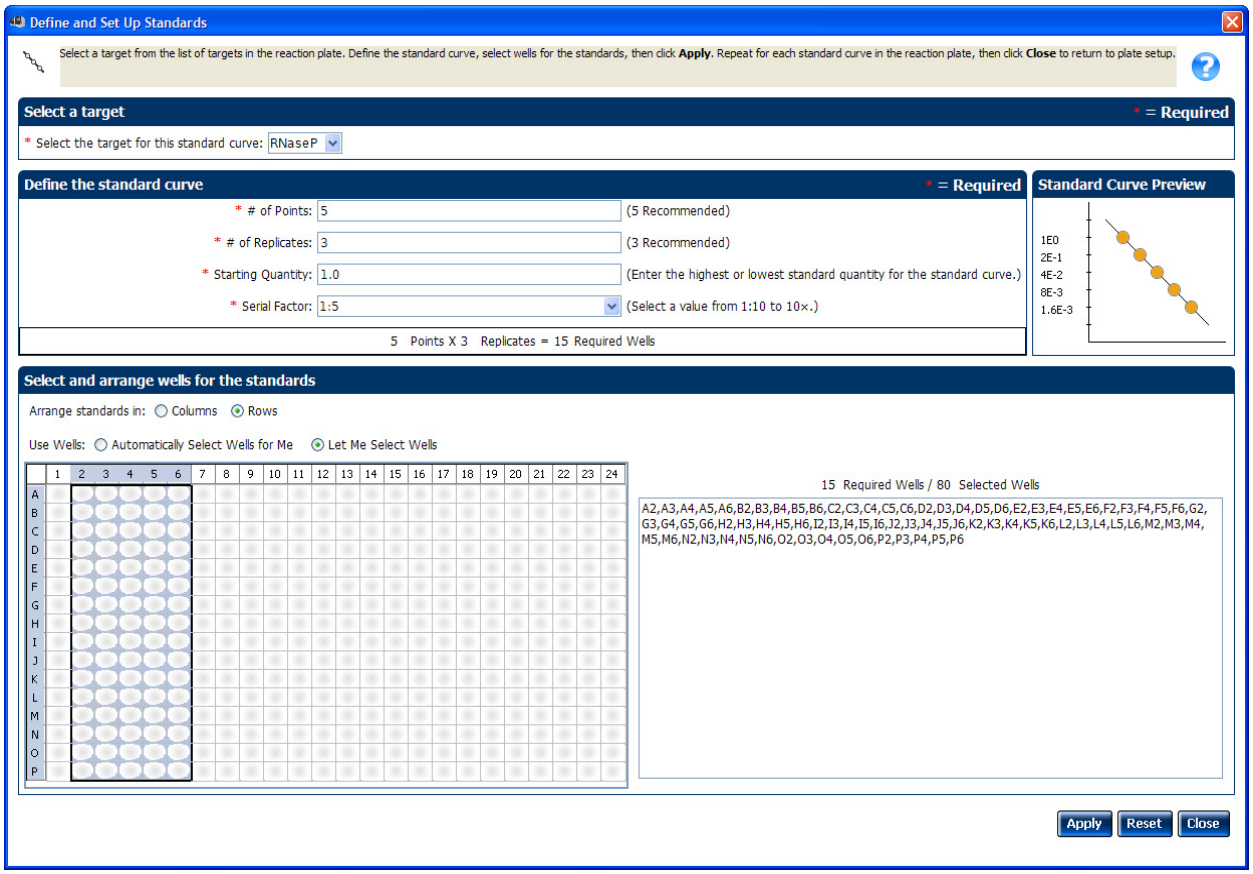
Field	Enter
# of Points	5
# of Replicates	16
Starting Quantity	1250.0
Serial Factor	2x

    d. Select and arrange wells for the standards.

Field	Select
Use Wells	Let Me Select Wells

e. Click **Apply**, and then **Close**.

Your Define and Set Up Standards dialog box should look like this:

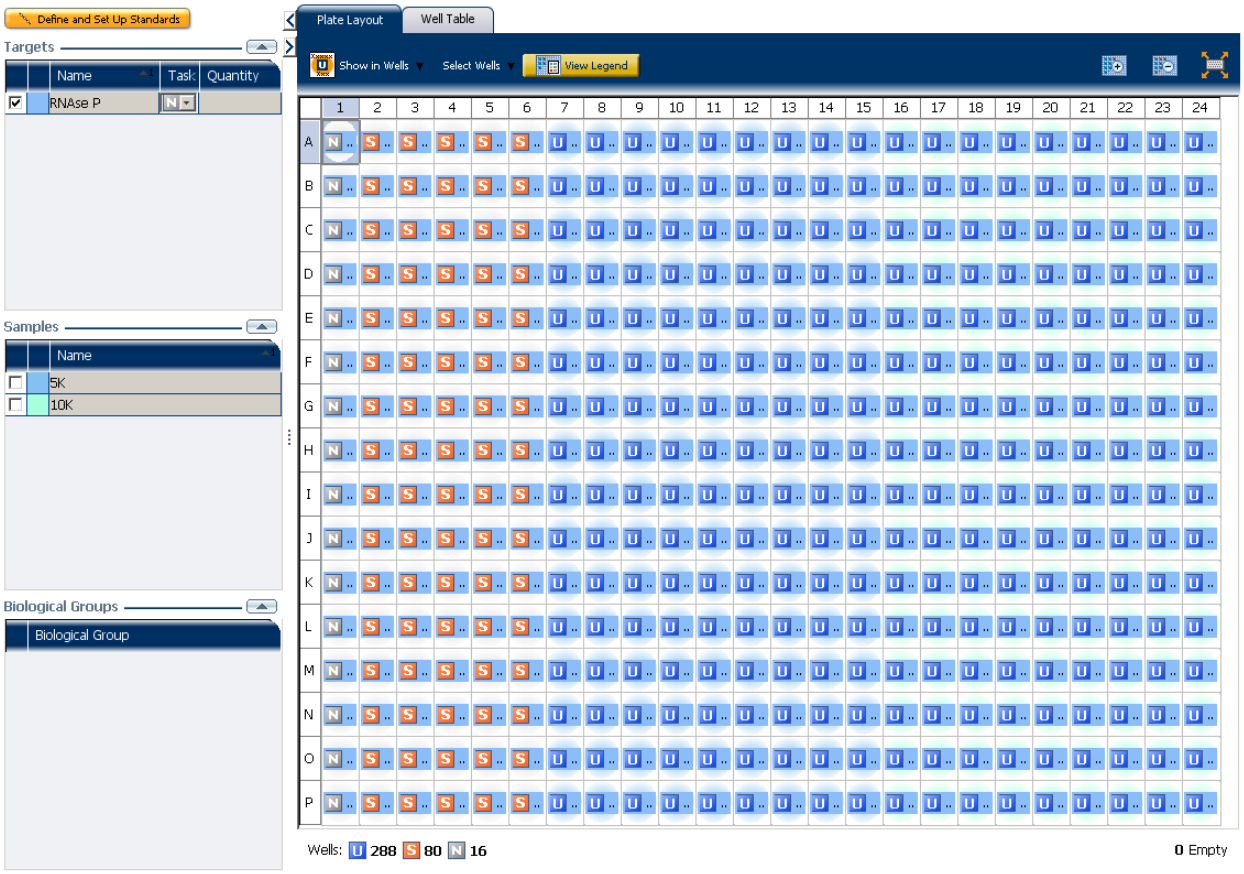


2. Assign targets and samples.

Target name	Well number	Task	Quantity	Sample name
RNaseP	A1 - P1 (column 1)	Negative	None	None
RNaseP	A2 - P2 (column 2)	Standard	1250	None
RNaseP	A3 - P3 (column 3)	Standard	2500	None
RNaseP	A4 - P4 (column 4)	Standard	5000	None
RNaseP	A5 - P5 (column 5)	Standard	10000	None

Target name	Well number	Task	Quantity	Sample name
RNaseP	A6 - P6 (column 6)	Standard	20000	None
RNaseP	A7 - P15 (columns 7 -15)	Unknown	Determined by run	5K
RNaseP	A16 - P24 (columns1 6 - 24)	Unknown	Determined by run	10K

Your Assign screen should look like this:



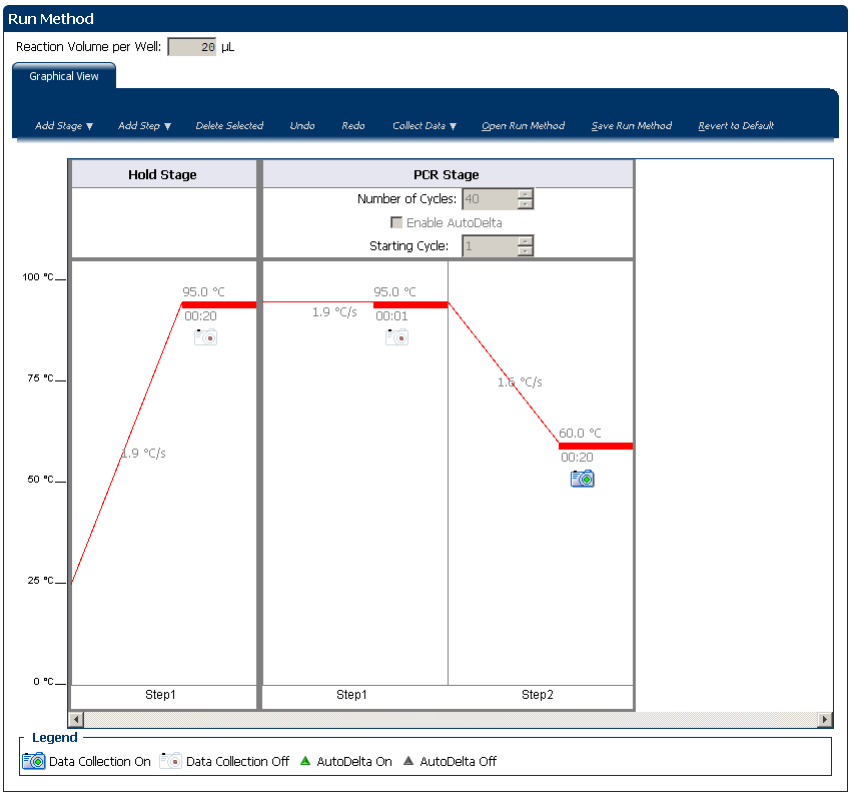
Set up the run method

Click **Run Method** to access the Run Method screen. Set the thermal profile under the Graphical View tab. Enter:

- Reaction Volume Per Well: 20 µL
- Thermal Profile

Stage	Step	Ramp rate	Temperature	Time
Hold Stage	Step 1	1.9°C/s	95°C	20 seconds
PCR Stage	Step 1	1.9°C/s	95°C	1 second
Number of Cycles: 40 (default) Enable AutoDelta: Unchecked (default) Starting Cycle: Disabled when Enable AutoDelta is unchecked	Step 2	1.6°C/s	60°C	20 seconds

Your Run Method screen should look like this:



For more information

For more information on...	Refer to...	Publication number
Consumables	Chapter 1 in Booklet 1, *Getting Started with QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Experiments* Appendix A in Booklet 7, *QuantStudio™ 6 and 7 Flex Real-Time PCR System Software - Appendixes*	4489822

For more information on...	Refer to...	Publication number
Using other quantification methods	Booklet 3, *Running Relative Standard Curve and Comparative C_T Experiments*.	Chapter 1 in Booklet 1, *Getting Started with QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Experiments* Appendix A in Booklet 7, *QuantStudio™ 6 and 7 Flex Real-Time PCR System Software - Appendixes*
Amplification efficiency	*Amplification Efficiency of TaqMan® Gene Expression Assays Application Note*	127AP05-03
Using Alternative Setup	Chapter 3 in Booklet 1, *Getting Started with QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Experiments*	4489822

3 Prepare the Reactions

Assemble required materials

- Items listed in Booklet 1, *Getting Started with QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Experiments*
- Samples - Human Raji cell line-derived cDNA samples (100 ng/µL)
- Example experiment reaction mix components:
  - **TaqMan® Fast Universal PCR Master Mix**
  - RNase P Assay Mix (20×) (Part no. 4316831)

Note: Fast Universal Master Mix is not recommended to be used with the 96-well (0.2 mL) reaction plates or reaction tubes and tube strips sealed with caps.

Prepare the sample dilutions

To determine the quantity of the RNase P gene in the example experiment, dilute the samples (as directed below) before adding the samples to the final reaction mix.

The stock concentration of each sample is 100 ng/µL. After dilution, the sample Pop1 has a concentration of 6.6 ng/µL and Pop 2 has a concentration of 3.3 ng/µL. Add 2µL to each reaction.

Use this table for sample dilution volumes for the example experiment.

Sample name	Stock concentration (ng/µL)	Sample volume (µL)	Diluent volume (µL)	Total volume of diluted sample (µL)
Pop1	100.0	25	355	380
Pop2	100.0	12.5	367.5	380

Note: For your own experiment, adjust the input amounts of the template depending on the template type and target abundance.

1. Label a separate microcentrifuge tube for each diluted sample:
   - **Pop 1**
   - **Pop 2**
2. Add the required volume of water (diluent) to each empty tube:

Tube	Sample name	Diluent volume (µL)
1	Pop 1	355
2	Pop 2	367.5

3. Add the required volume of sample stock to each tube:

Tube	Sample name	Sample volume (µL)
1	Pop 1	25
2	Pop 2	12.5

4. Vortex each diluted sample for 3 to 5 seconds, then centrifuge the tubes briefly.
5. Place the diluted samples on ice until you prepare the reaction plate.

Prepare the standard dilution series

The standard concentration in stock is 20,000 copies/µL:

Standard name (labeled tube)	Dilution point	Source	Source volume (µL)	Diluent volume (µL)	Total volume (µL)	Standard concentration (copies/µL)
RNase P Std. 1	1 (20,000)	Stock	18	18	36	10,000
RNase P Std. 1	2 (10,000)	Dilution 1	18	18	36	5,000
RNase P Std. 1	3 (5,000)	Dilution 2	18	18	36	2,500
RNase P Std. 1	4 (2500)	Dilution 3	18	18	36	1250
RNase P Std. 1	5 (1250)	Dilution 4	18	18	36	625

1. Prepare five standard dilutions:

   Note: For dilution 1, first vortex the stock for 3 to 5 seconds, then centrifuge the RNase P Std. 1 tube briefly before pipetting stock into the tube.

   For each dilution:

   a. Use a new pipette tip to add 18 µL of source to the tube containing the standard.

   b. Vortex the tube for 3 to 5 seconds, then centrifuge the tube briefly.

2. Place the standards on ice until you prepare the reaction plate.

Prepare the reaction mix ("cocktail mix")

For the RNase P assay (Standard Curve example experiment), the following table lists the universal assay conditions (volume and final concentration) for using the TaqMan® Fast Universal PCR Master Mix.

Reaction component	Volume for 1 reaction (µL)	Volume for 384 reactions + 10% excess (µL)
TaqMan® Fast Universal PCR Master Mix Kit	5	2115
RNase P Assay (20X)	0.5	211.5
Water	3.5	1480.5
Total reaction mix volume	9	3807

1. Label an appropriately sized tube for the reaction mix: **RNase P Reaction Mix**.
2. Add the required volumes of each cocktail mix component to the tube.

   Note: Do not add the sample or standard at this time.
3. Mix the cocktail thoroughly by gently pipetting up and down several times, then cap the tube.
4. Centrifuge the tube briefly to remove air bubbles.
5. Place the cocktail mix on ice until you prepare the reaction plate.

Note: You can separately add the sample to the reaction plate, as opposed to preparing individual reaction mixes for each sample.

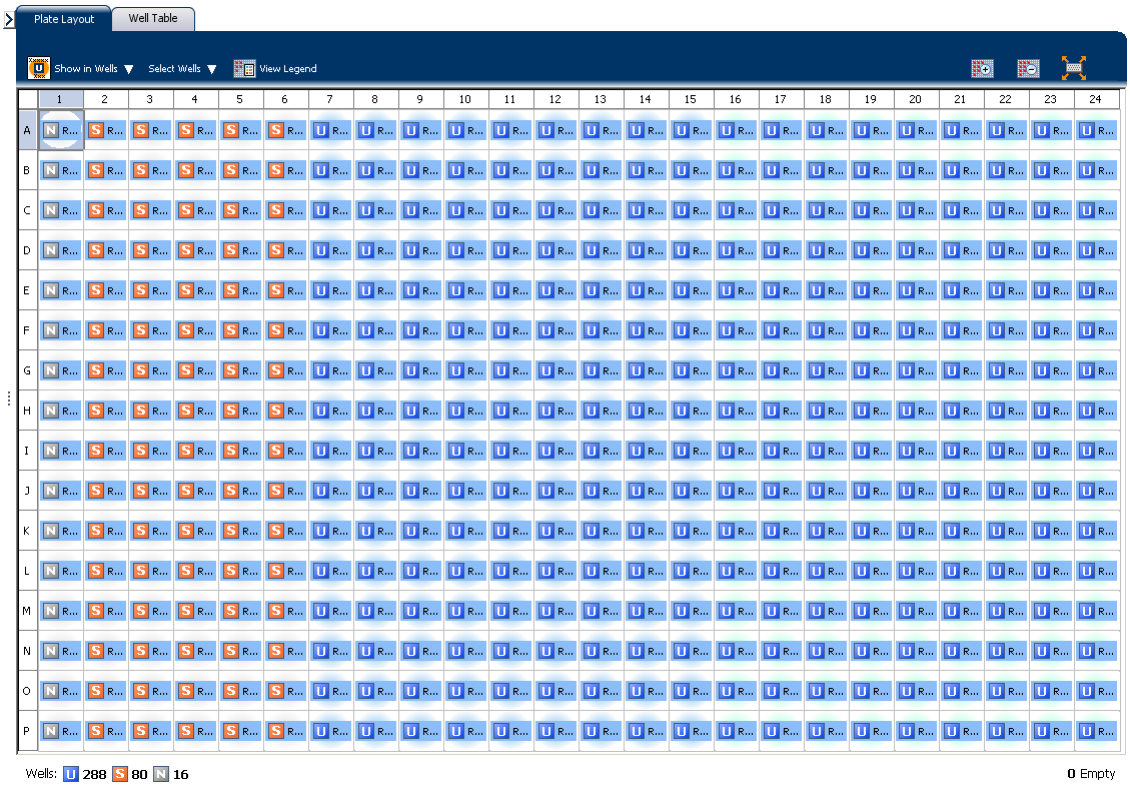
Prepare the reaction plate

Example experiment reaction plate components

The reaction plate for the Standard Curve example experiment contains:

- A MicroAmp® Optical 384-Well Reaction Plate
- Reaction volume: 20 µL/well
- 288 Unknown wells 
- 80 Standard wells 
- 16 Negative control wells 

The following is an image of the plate layout:



To prepare the reaction plate components

1. Prepare the negative control reactions for the target:
   a. To an appropriately sized tube, add the volumes of reaction mix and water listed below.

Tube	Reaction mix	Reaction mix volume (µL)	Water volume (µL)
1	RNase P reaction mix	157.5	17.5

   b. Mix the reaction by gently pipetting up and down, then cap the tube.
   c. Centrifuge the tube briefly to remove air bubbles.
   d. Add 10 µL of the negative control reaction to the appropriate wells in the reaction plate.
2. For each replicate group, prepare the standard reactions:
   a. To appropriately sized tubes, add the volumes of reaction mix and standard listed below.

Tube	Standard reaction	Reaction mix	Reaction mix volume (µL)	Standard	Standard volume (µL)
1	RNase P Std 1	RNase P reaction mix	157.5	RNase P Std 1	17.5
2	RNase P Std 2	RNase P reaction mix	157.5	RNase P Std 2	17.5
3	RNase P Std 3	RNase P reaction mix	157.5	RNase P Std 3	17.5
4	RNase P Std 4	RNase P reaction mix	157.5	RNase P Std 4	17.5
5	RNase P Std 5	RNase P reaction mix	157.5	RNase P Std 5	17.5

b. Mix the reactions by gently pipetting up and down, then cap the tubes.

c. Centrifuge the tubes briefly to remove air bubbles.

d. Add 20 µL of the standard reaction to the appropriate wells in the reaction plate.

3. For each replicate group, prepare the reactions for the unknowns:

a. To appropriately sized tubes, add the volumes of reaction mix and sample listed below.

Tube	Unknown reaction	Reaction mix	Reaction mix volume (µL)	Sample	Sample volume (µL)
1	RNase P pop1	RNase P reaction mix	1422	pop1	158
2	RNase P pop2	RNase P reaction mix	1422	pop2	158

b. Mix the reactions by gently pipetting up and down, then cap the tubes.

c. Centrifuge the tubes briefly to remove air bubbles.

d. Add 10 µL of the unknown (sample) reaction to the appropriate wells in the reaction plate.

4. Seal the reaction plate with optical adhesive film.

5. Centrifuge the reaction plate briefly to remove air bubbles.

6. Confirm that the liquid is at the bottom of each well of the reaction plate. If not, centrifuge the reaction plate again at a higher speed and for a longer period of time.
7. Until you are ready to perform the run, place the reaction plate at 4°C, in the dark.

For more information

For more information on...	Refer to...	Publication number
Assigning the reaction plate components	Chapter 2 in Booklet 1, *Getting Started with QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Experiments*	Chapter 1 in Booklet 1, *Getting Started with QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Experiments* Appendix A in Booklet 7, *QuantStudio™ 6 and 7 Flex Real-Time PCR System Software - Appendixes*
Sealing the reaction plate	Chapter 2 in Booklet 1, *Getting Started with QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Experiments*	Chapter 1 in Booklet 1, *Getting Started with QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Experiments* Appendix A in Booklet 7, *QuantStudio™ 6 and 7 Flex Real-Time PCR System Software - Appendixes*

4 Run the Experiment

This chapter explains how to run the example experiment on the QuantStudio™ 6 or 7 Instrument.

This chapter covers:

IMPORTANT! Run the experiment at the same ambient temperature at which you calibrated the instrument. Extreme variations in ambient temperature can affect the heating and cooling of the instrument and influence experimental results.

IMPORTANT! Do not attempt to open the access door during the run. The door is locked while the QuantStudio™ 6 or 7 Instrument is in operation.

Start the run

1. Open the Standard Curve example file that you created using instructions in Chapter 2.
2. Load the reaction plate into the instrument.
3. Start the run.

Monitor the run

Monitor the example experiment run:

- From the QuantStudio™ 6 and 7 Flex Real-Time PCR System Software using the Run screen, while the experiment is in progress.
- From the Instrument Console of the QuantStudio™ 6 and 7 Flex Real-Time PCR System Software (to monitor an experiment started from another computer or from the QuantStudio™ 6 or 7 Instrument touchscreen).
- From the QuantStudio™ 6 or 7 Instrument touchscreen.

From the Instrument Console of the QuantStudio™ 6 and 7 Flex Real-Time PCR System Software

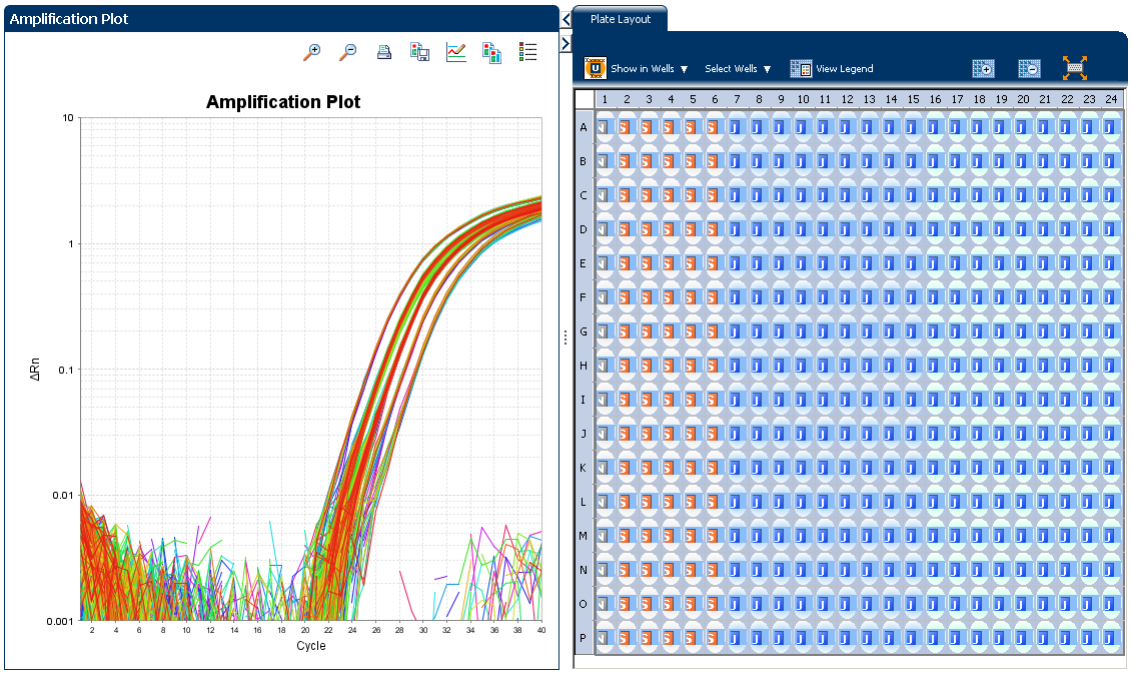
1. In the Instrument Console screen, select the instrument icon.
2. Click **Manage Instrument** or double-click on the instrument icon.
3. On the Manage Instrument screen, click **Monitor Running Experiment** to access the Run screen.

View the Amplification Plot

You can view the progress of the run in real time. During the run, periodically view all the three plots available from the QuantStudio™ 6 and 7 Flex Real-Time PCR System Software for potential problems.

Click **Amplification Plot** from the Run Experiment Menu, select the Plate Layout tab, then select the wells to view.

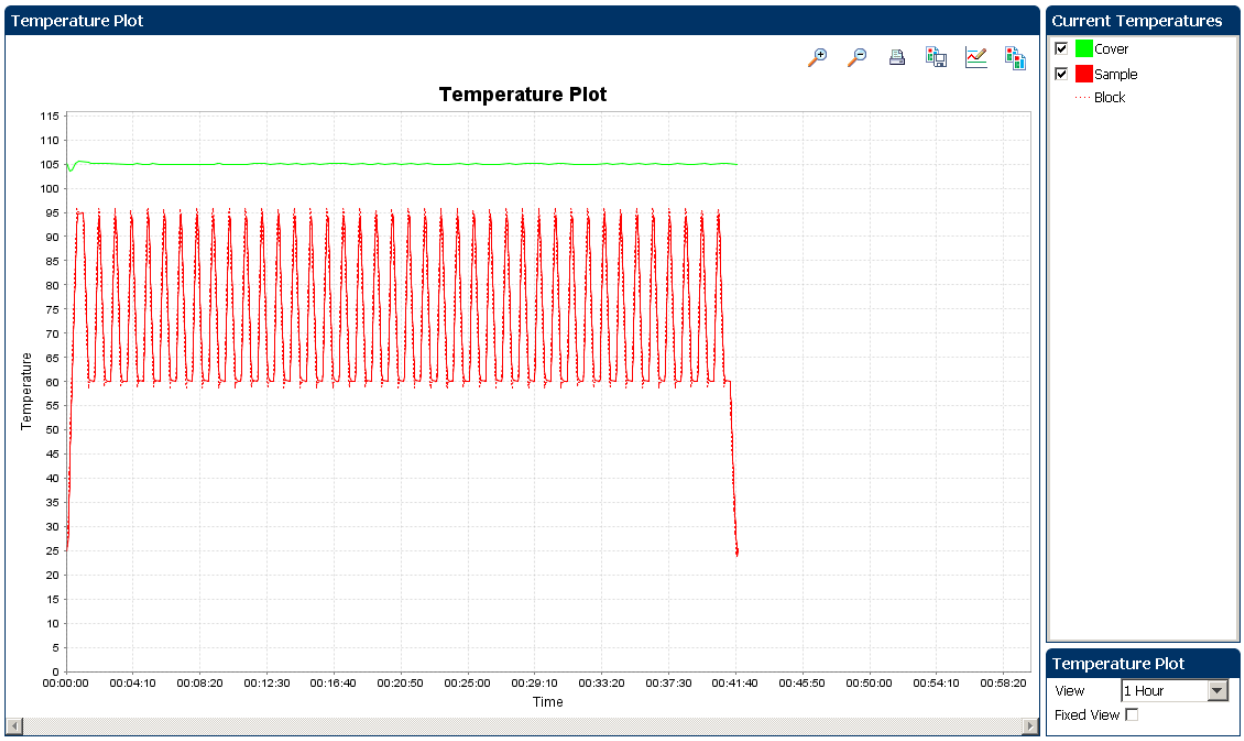
The following is an image of the Amplification Plot screen as it appears at the end of the example experiment.



View the Temperature Plot

Click **Temperature Plot** from the Run Experiment Menu.

The following is an image of the Temperature Plot screen as it appears during the example experiment.

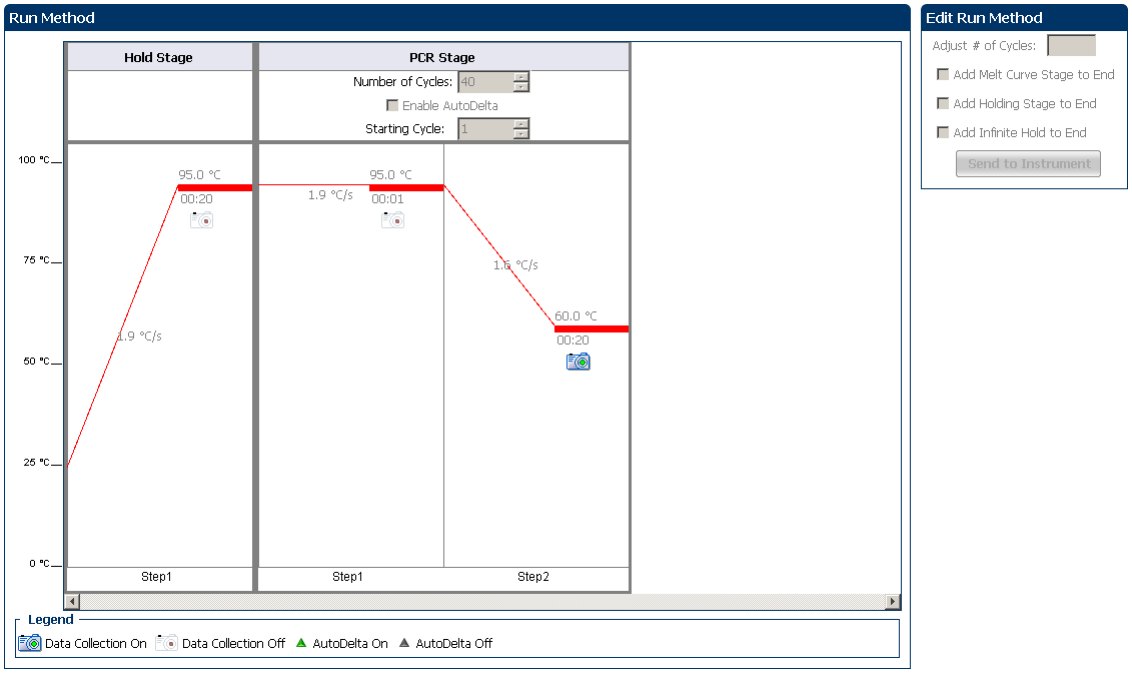


Note: The Sample temperature displayed in the Current Temperatures group is an estimated value.

View the Run Method

Click **Run Method** from the Run Experiment Menu.

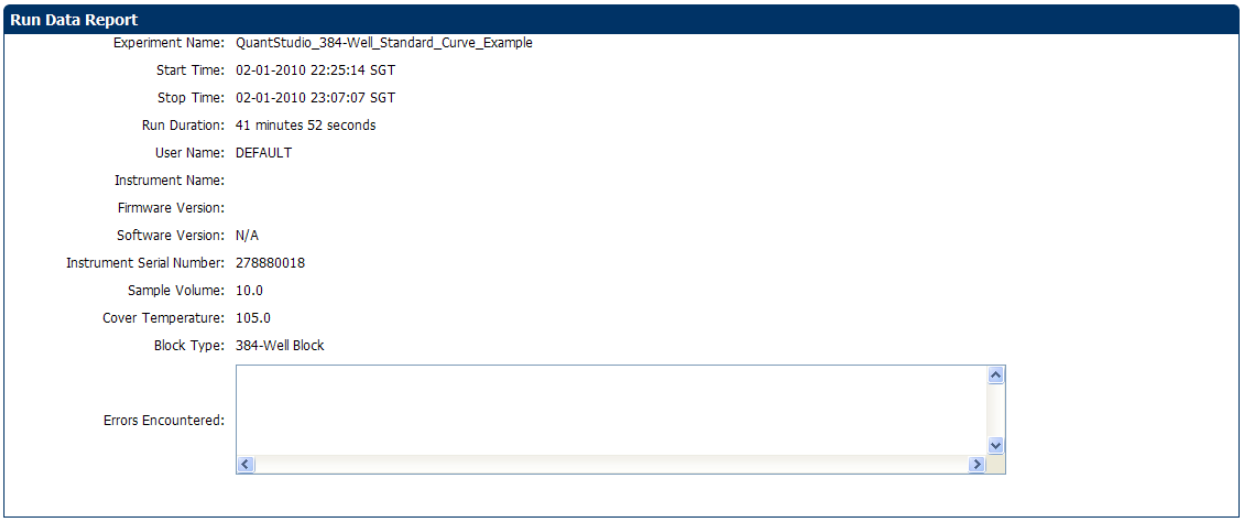
The following is an image of the Run Method screen as it appears in the example experiment.



View the run data

Click **View Run Data** from the Run Experiment Menu.

The following is an image of the View Run Data screen as it appears in the example experiment.

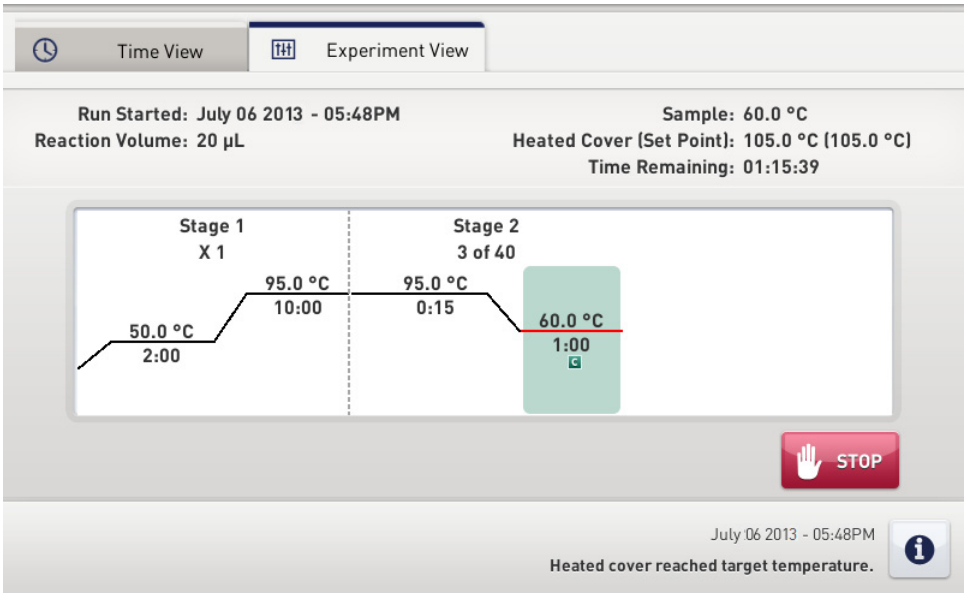


From the QuantStudio™ 6 or 7 Instrument touchscreen

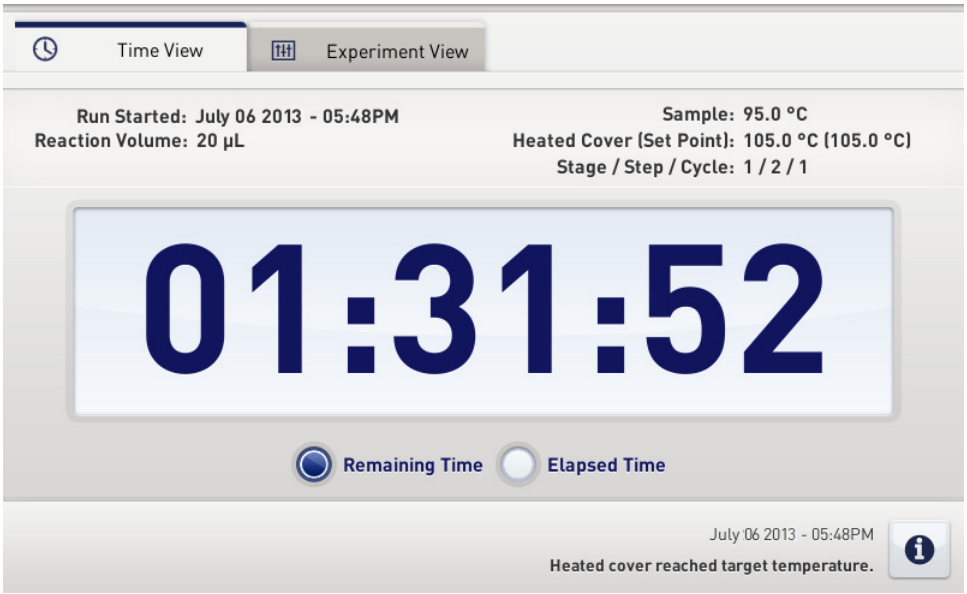
You can also view the progress of the run from the QuantStudio™ 6 or 7 Instrument touchscreen.

The following are images of the Run Method screen on the QuantStudio™ 6 or 7 Instrument touchscreen:

Experiment View



Time View



5 Review Results and Adjust Experiment Parameters

In Section 5.1 of this chapter you review the analyzed data using several of the analysis screens and publish the data. Section 5.2 of this chapter walks you through the process of modifying experiment parameters to troubleshoot problems with experiment results before rerunning an experiment.

This chapter covers:

Section 5.1 Review Results

Analyze the example experiment

1. Open the example experiment file that you ran in Chapter 4.
2. Click **Analyze**. The software analyzes the data using the default analysis settings.

   Note: You can also access the experiment to analyze from the Home screen.

View the Standard Curve Plot

The Standard Curve Plot screen displays the standard curve for samples designated as standards. The QuantStudio™ 6 and 7 Flex Real-Time PCR System Software calculates the quantity of an unknown target from the standard curve.

Purpose

The purpose of viewing the standard curve for the example experiment is to identify:

- Slope and amplification efficiency
- R^2 value (correlation coefficient)
- C_T values

To view and assess the Standard Curve Plot

1. From the Experiment Menu pane, select **Analysis ▸ Standard Curve.**

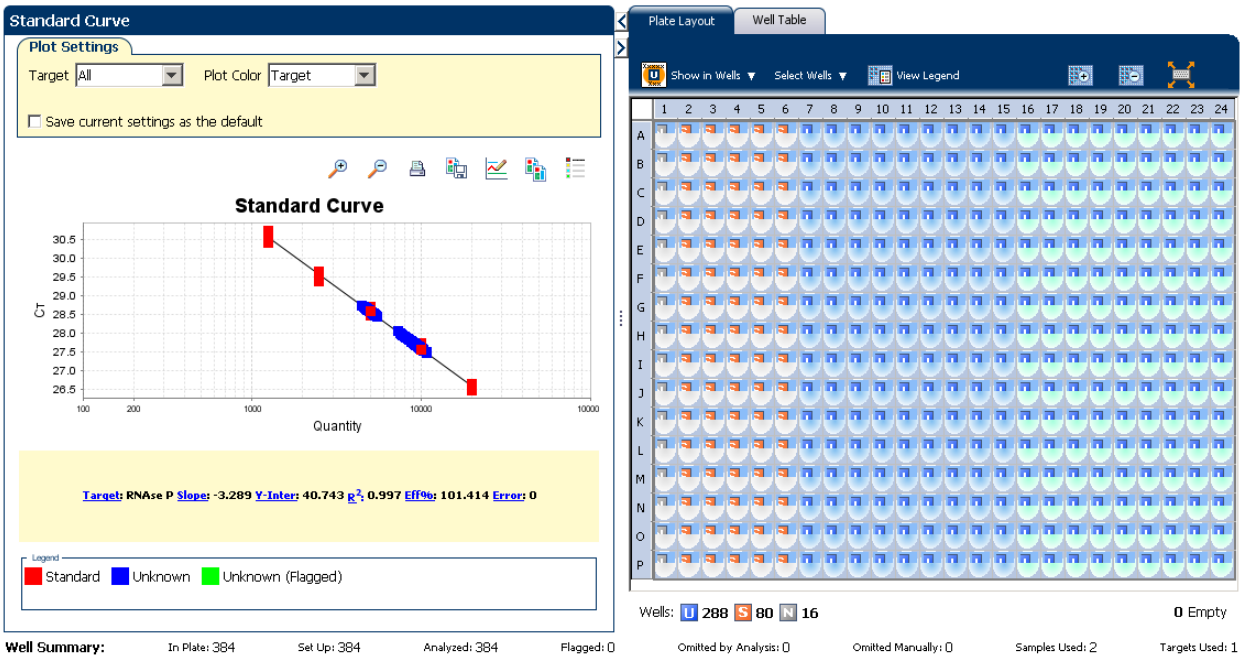
   Note: If no data are displayed, click **Analyze.**
2. Display all 384 wells in the Standard Curve Plot screen by clicking the upper left corner of the plate layout in the **Plate Layout** tab.
3. Enter the Plot Settings:

Menu	Selection
Target	All
Plot Color	Target
 (This is a toggle button. When the legend is displayed, the button changes to **Hide the plot legend**.)	Check (default)

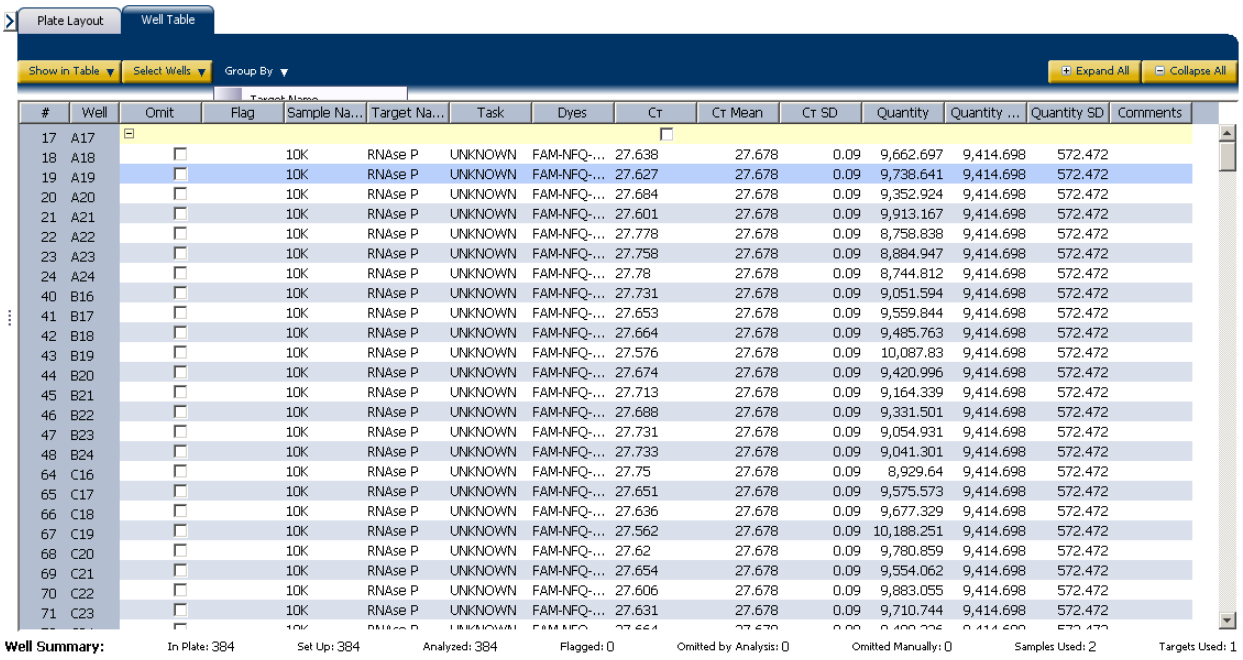
4. View the values displayed below the standard curve.

Menu	Selection
Slope	-3.289
R^2	0.997
Amplification efficiency	101.414%
Error	0

5. Check that all samples are within the standard curve. In the example experiment, as shown in the following image, all samples (blue dots) are within the standard curve (red dots).



6. Check the C_T values:
   a. Click the **Well Table** tab.
   b. From the Group By menu, select **Replicate**.
   c. Look at the values in the C_T column. In the example experiment, the C_T values fall within the expected range (>8 and < 35).



#	Well	Omit	Flag	Sample Na...	Target Na...	Task	Dyes	CT	CT Mean	CT SD	Quantity	Quantity ...	Quantity SD	Comments
17	A17													
18	A18			10K	RNAse P	UNKNOWN	FAM-NFQ-...	27.638	27.678	0.09	9,662.697	9,414.698	572.472	
19	A19			10K	RNAse P	UNKNOWN	FAM-NFQ-...	27.627	27.678	0.09	9,738.641	9,414.698	572.472	
20	A20			10K	RNAse P	UNKNOWN	FAM-NFQ-...	27.684	27.678	0.09	9,352.924	9,414.698	572.472	
21	A21			10K	RNAse P	UNKNOWN	FAM-NFQ-...	27.601	27.678	0.09	9,913.167	9,414.698	572.472	
22	A22			10K	RNAse P	UNKNOWN	FAM-NFQ-...	27.778	27.678	0.09	8,758.838	9,414.698	572.472	
23	A23			10K	RNAse P	UNKNOWN	FAM-NFQ-...	27.758	27.678	0.09	8,884.947	9,414.698	572.472	
24	A24			10K	RNAse P	UNKNOWN	FAM-NFQ-...	27.78	27.678	0.09	8,744.812	9,414.698	572.472	
40	B16			10K	RNAse P	UNKNOWN	FAM-NFQ-...	27.731	27.678	0.09	9,051.594	9,414.698	572.472	
41	B17			10K	RNAse P	UNKNOWN	FAM-NFQ-...	27.653	27.678	0.09	9,559.844	9,414.698	572.472	
42	B18			10K	RNAse P	UNKNOWN	FAM-NFQ-...	27.664	27.678	0.09	9,485.763	9,414.698	572.472	
43	B19			10K	RNAse P	UNKNOWN	FAM-NFQ-...	27.576	27.678	0.09	10,087.83	9,414.698	572.472	
44	B20			10K	RNAse P	UNKNOWN	FAM-NFQ-...	27.674	27.678	0.09	9,420.996	9,414.698	572.472	
45	B21			10K	RNAse P	UNKNOWN	FAM-NFQ-...	27.713	27.678	0.09	9,164.339	9,414.698	572.472	
46	B22			10K	RNAse P	UNKNOWN	FAM-NFQ-...	27.688	27.678	0.09	9,331.501	9,414.698	572.472	
47	B23			10K	RNAse P	UNKNOWN	FAM-NFQ-...	27.731	27.678	0.09	9,054.931	9,414.698	572.472	
48	B24			10K	RNAse P	UNKNOWN	FAM-NFQ-...	27.733	27.678	0.09	9,041.301	9,414.698	572.472	
64	C16			10K	RNAse P	UNKNOWN	FAM-NFQ-...	27.75	27.678	0.09	8,929.64	9,414.698	572.472	
65	C17			10K	RNAse P	UNKNOWN	FAM-NFQ-...	27.651	27.678	0.09	9,575.573	9,414.698	572.472	
66	C18			10K	RNAse P	UNKNOWN	FAM-NFQ-...	27.636	27.678	0.09	9,677.329	9,414.698	572.472	
67	C19			10K	RNAse P	UNKNOWN	FAM-NFQ-...	27.562	27.678	0.09	10,188.251	9,414.698	572.472	
68	C20			10K	RNAse P	UNKNOWN	FAM-NFQ-...	27.62	27.678	0.09	9,780.859	9,414.698	572.472	
69	C21			10K	RNAse P	UNKNOWN	FAM-NFQ-...	27.654	27.678	0.09	9,554.062	9,414.698	572.472	
70	C22			10K	RNAse P	UNKNOWN	FAM-NFQ-...	27.606	27.678	0.09	9,883.055	9,414.698	572.472	
71	C23			10K	RNAse P	UNKNOWN	FAM-NFQ-...	27.631	27.678	0.09	9,710.744	9,414.698	572.472	

Tips for analyzing your own experiments

When you analyze your own standard curve experiment, look for:

- **Slope and amplification efficiency values** – The amplification efficiency is calculated using the slope of the regression line in the standard curve. A slope close to – 3.3 indicates optimal, 100% PCR amplification efficiency. Factors that affect amplification efficiency:
  - Range of standard quantities – For accurate and precise efficiency measurements, use a broad range of standard quantities, 5 to 6 logs (10^5 to 10^6 fold).
  - Number of standard replicates – For accurate efficiency measurements, include replicates to decrease the effects of pipetting inaccuracies.
  - PCR inhibitors – PCR inhibitors in the reaction can reduce amplification efficiency.
- **R^2 values (correlation coefficient)** – The R^2 value is a measure of the closeness of fit between the regression line and the individual C_T data points of the standard reactions. A value of 1.00 indicates a perfect fit between the regression line and the data points. An R^2 value >0.99 is desirable.
- **C_T values** – The threshold cycle (C_T) is the PCR cycle number at which the fluorescence level meets the threshold.
  - A C_T value >8 and <35 is desirable.
  - A C_T value <8 indicates that there is too much template in the reaction.
  - A C_T value >35 indicates a low amount of target in the reaction; for C_T values >35, expect a higher standard deviation.

If your experiment does not meet the guidelines above, troubleshoot as follows:

- Omit wells (see "Improve C_T precision by omitting wells" on page 52).

  Or
- Rerun the experiment.

Assess amplification results using the Amplification Plot

Amplification plots available for viewing

The Amplification Plot screen displays amplification of all samples in the selected wells. There are three plots available:

- **ΔRn vs Cycle** – ΔRn is the magnitude of normalized fluorescence signal generated by the reporter at each cycle during the PCR amplification. This plot displays ΔRn as a function of cycle number. Use this plot to identify and examine irregular amplification and to view threshold and baseline values for the run.
- **Rn vs Cycle** – Rn is the fluorescence signal from the reporter dye normalized to the fluorescence signal from the passive reference. This plot displays Rn as a function of cycle number. Use this plot to identify and examine irregular amplification.
- **C_T vs Well** – C_T is the PCR cycle number at which the fluorescence meets the threshold in the amplification plot. This plot displays C_T as a function of well position. Use this plot to locate outlying amplification (outliers).

Each plot can be viewed as a linear or log10 graph type.

Purpose

The purpose of viewing the amplification plot for the example experiment is to identify:

- Correct baseline and threshold values
- Outliers

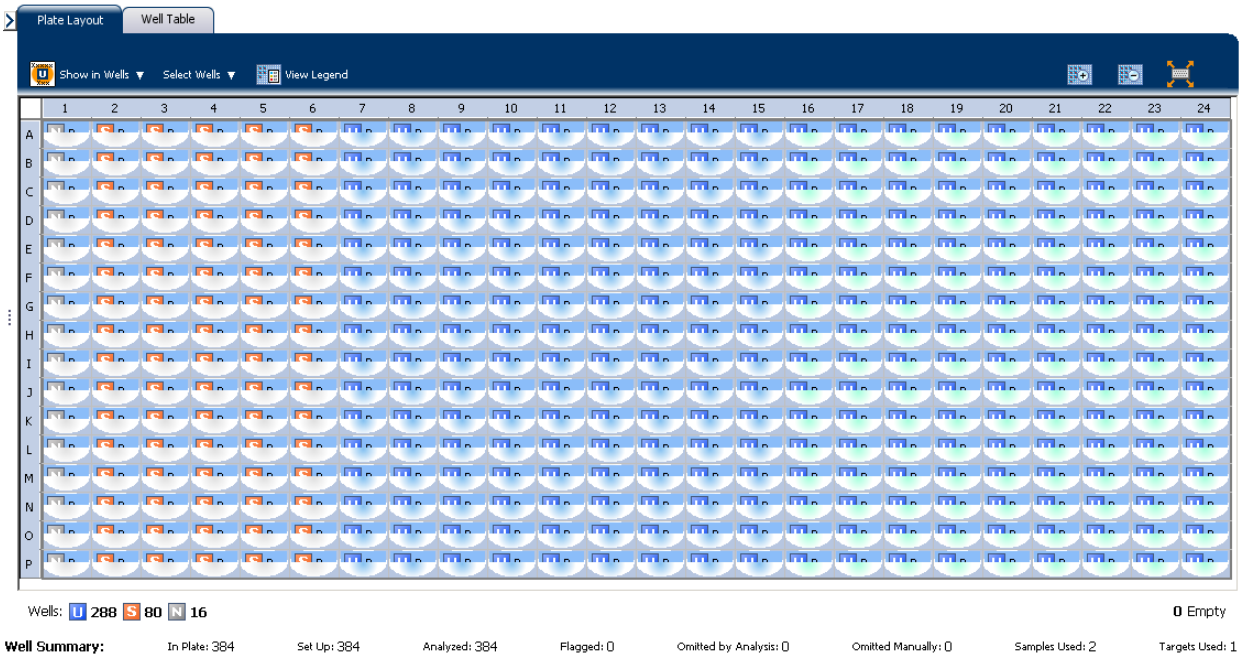
View the Amplification Plot

1. From the Experiment Menu pane, select **Analysis ▸ Amplification Plot.**

   Note: If no data are displayed, click **Analyze.**

2. Display the RNase P wells in the Amplification Plot screen. Click the **Plate Layout** tab. Enter the Plot Settings:

Menu	Selection
Select Wells With	**Target ▸ RNaseP**



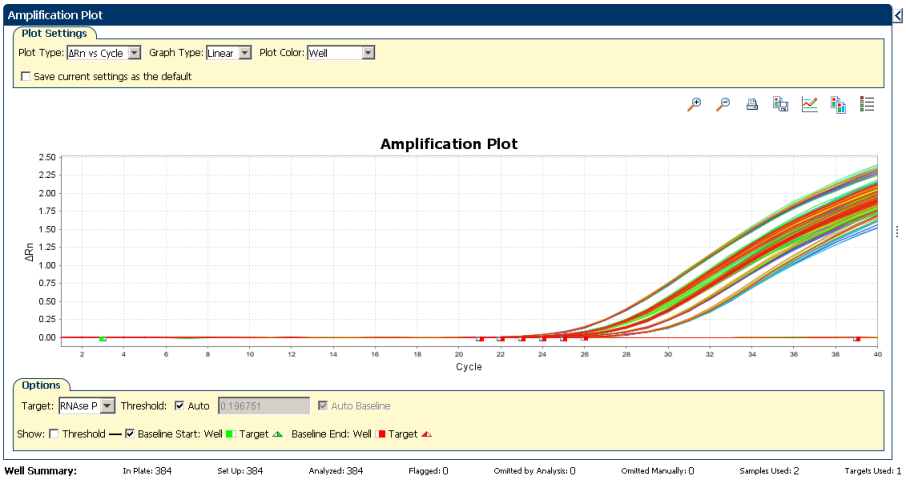
3. In the Amplification Plot screen, enter:

Menu	Select
Plot Type	**ΔRn vs Cycle**
Plot Color	**Well** (default)
 (This is a toggle button. When the legend is displayed, the button changes to **Hide the plot legend**.)	Check (default)

4. View the baseline values.
   a. From the Graph Type drop-down menu, select **Linear**.

b. Select the **Baseline** check box to show the start cycle and end cycle.

c. Verify that the baseline is set correctly: The end cycle should be set a few cycles before the cycle number where significant fluorescent signal is detected. In the example experiment, the baseline is set correctly.

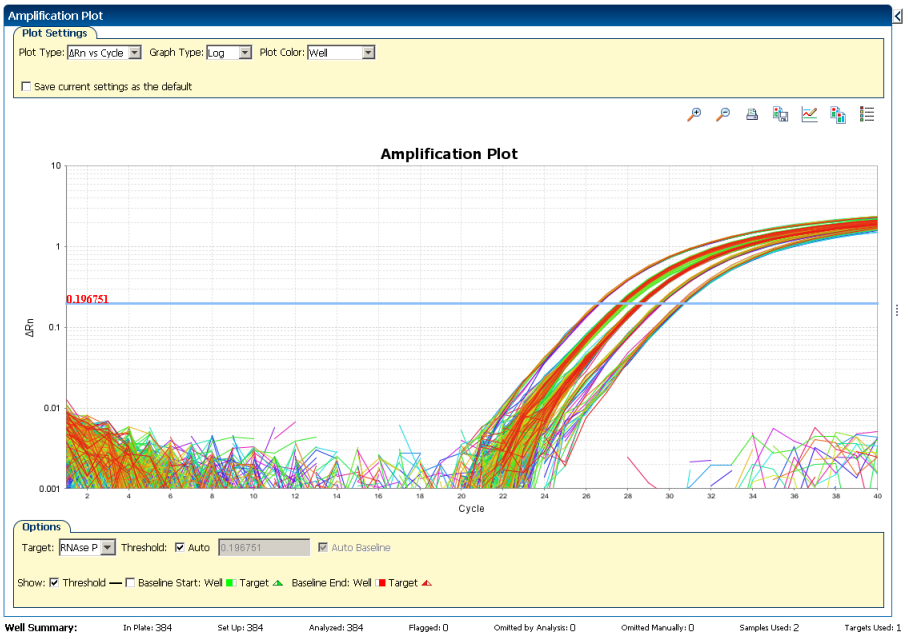


5. View the threshold values.

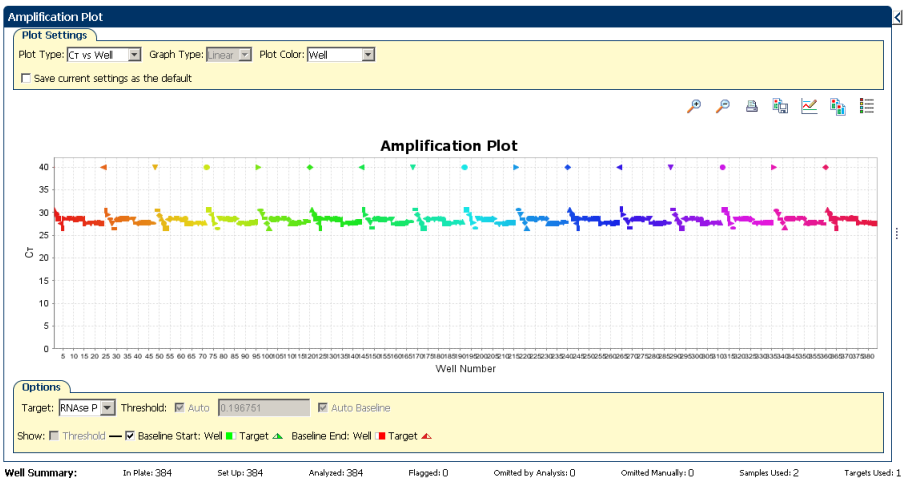
Menu	Select
Graph Type	Log
Target	RNaseP

a. Select the **Threshold** check box to show the threshold.

b. Verify that the threshold is set correctly. In the example experiment, the threshold is in the exponential phase.



6. Locate outliers:
   a. From the Plot Type drop-down menu, select **C_T vs Well**.
   b. Look for outliers from the amplification plot. In the example experiment, there are no outliers for RNase P.



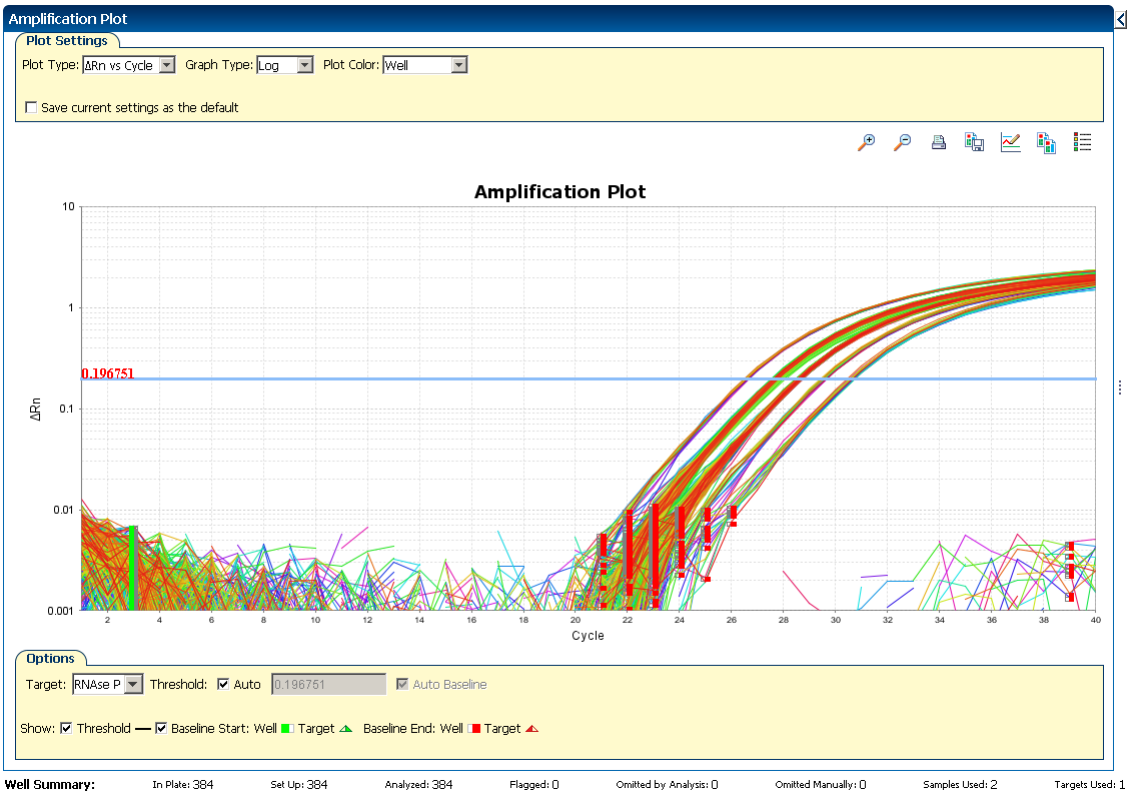
Tips for analyzing your own experiments

When you analyze your own standard curve experiment, look for:

Outliers

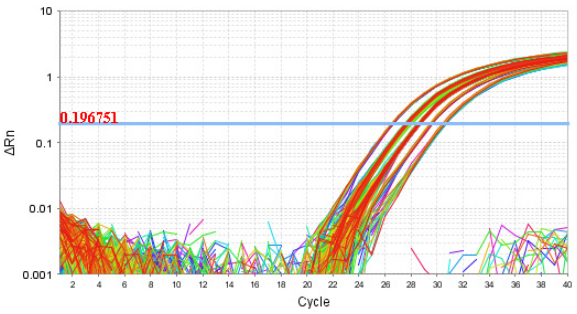
- **A typical amplification plot** – The QuantStudio™ 6 and 7 Flex Real-Time PCR System Software automatically calculates baseline and threshold values based on the assumption that the data exhibit a *typical* amplification plot. A typical amplification plot has four distinct sections:
  - Plateau phase
  - Linear phase
  - Exponential (geometric phase)
  - Baseline

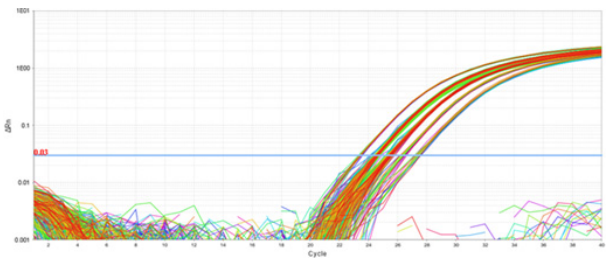
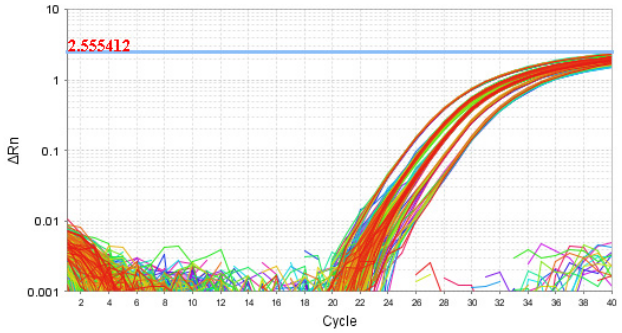
The following is an image of a typical amplification plot:



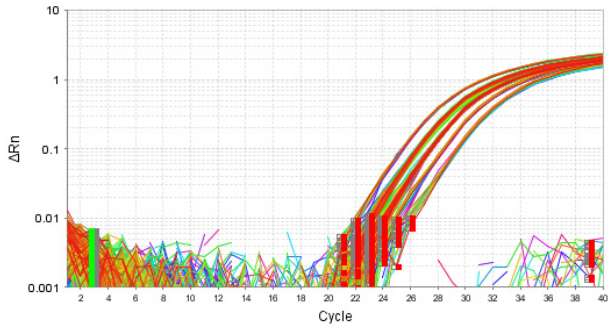
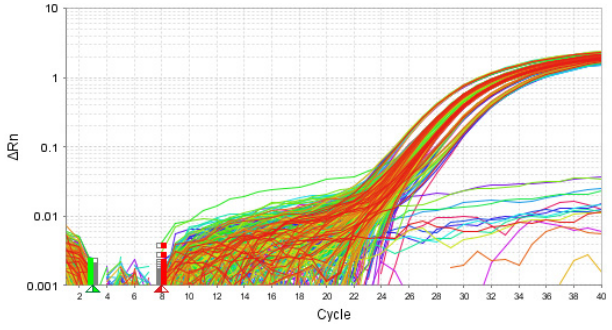
IMPORTANT! Experimental error (such as contamination or pipetting errors) can produce atypical amplification curves that can result in incorrect baseline and threshold value calculations by the QuantStudio™ 6 and 7 Flex Real-Time PCR System Software. Life Technologies recommends that you examine the Amplification Plot screen and review the assigned baseline and threshold values for each well after analysis.

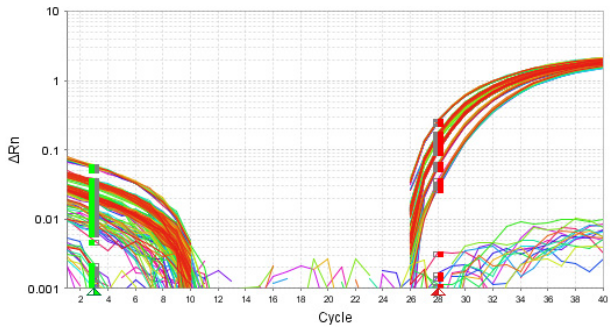
- **Correct threshold values:**

Threshold Set Correctly	
The threshold is set in the exponential phase of the amplification curve. Threshold settings above or below the optimum increase the standard deviation of the replicate groups.	

Threshold Set Too Low The threshold is set below the exponential phase of the amplification curve. The standard deviation is significantly higher than that for a plot where the threshold is set correctly. Drag the threshold bar up into the exponential phase of the curve.	
Threshold Set Too High The threshold is set above the exponential phase of the amplification curve. The standard deviation is significantly higher than that for a plot where the threshold is set correctly. Drag the threshold bar down into the exponential phase of the curve.	

- **Correct baseline values**:

Baseline Set Correctly The amplification curve begins after the maximum baseline.	
Baseline Set Too Low The amplification curve begins too far to the right of the maximum baseline. Increase the End Cycle value.	

Baseline Set Too High The amplification curve begins before the maximum baseline. Decrease the End Cycle value.	

If your experiment does not meet the guidelines above, troubleshoot as follows:

- Omit wells (see "Improve C_T precision by omitting wells" on page 52).

  Or
- Manually adjust the baseline and/or threshold (see "Adjust analysis settings" on page 49).

Identify well problems using the Well Table

The Well Table displays data for each well in the reaction plate, including:

- The sample name, target name, task, and dyes
- The calculated threshold cycle (C_T), normalized fluorescence (Rn), and quantity values
- Comments
- Flags

Purpose

The purpose of viewing the well table is to identify:

- Quantity values
- Flags
- C_T values (including C_T standard deviation)

View the well table

1. From the Experiment Menu pane, select **Analysis**, then select the **Well Table** tab.

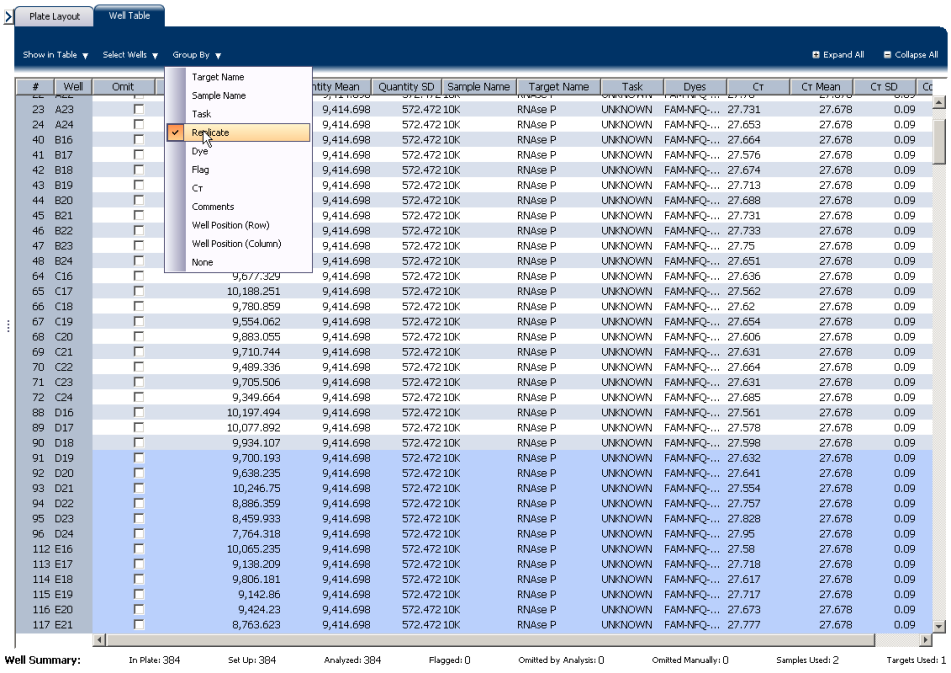
   Note: If no data are displayed, click **Analyze**.
2. Use the Group By drop-down menu to group wells by a specific category. For the example experiment, group the wells by replicate, flag, or C_T value.

   Note: You can select only one category at a time.

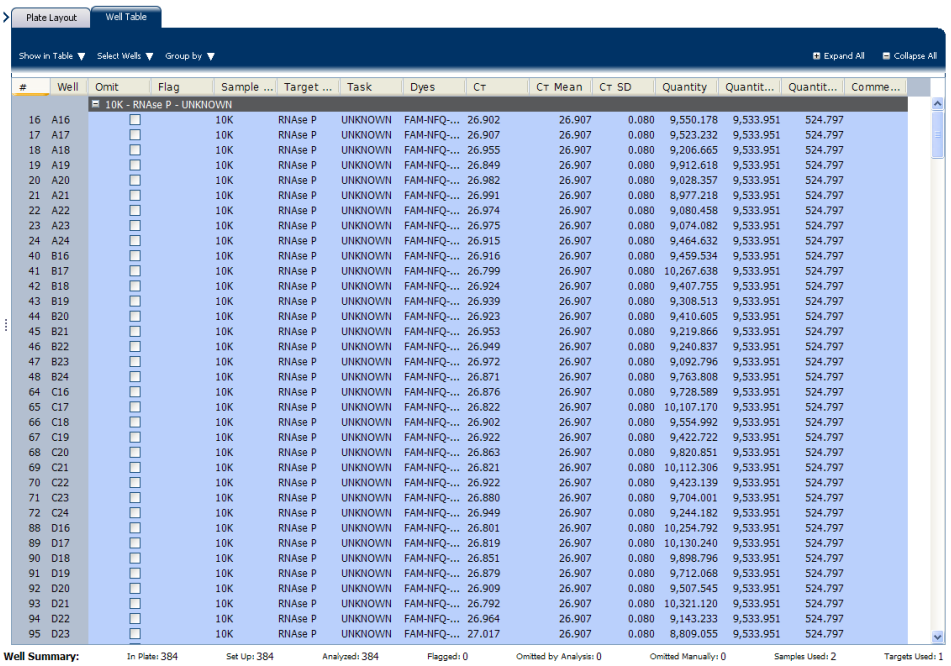
To group by replicate

From the Group By drop-down menu, select **Replicate**. The software groups the replicate wells: negative controls, standards, and samples. In the example experiment, note that the quantity values within each replicate group are similar.

Note: In the example experiment, the Quantity, Quantity Mean, and Quantity SD columns have been moved from their default locations to the beginning of the Well Table. To move a column, click and drag on the column heading.

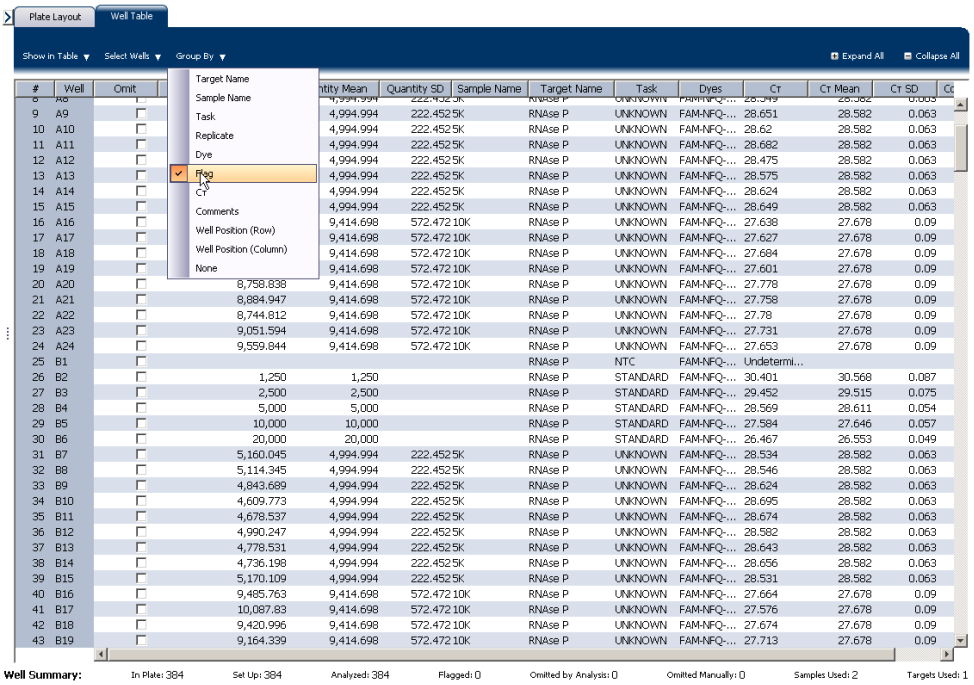


The well table looks like this:



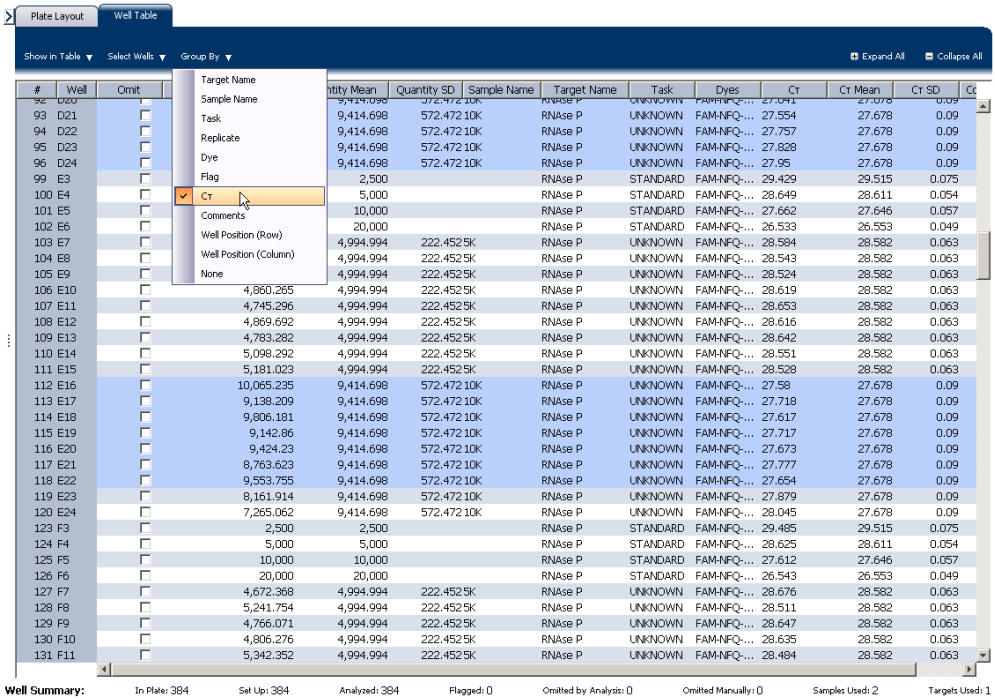
To group by flag

From the Group By drop-down menu, select **Flag**. The software groups the flagged and unflagged wells. In the example experiment, there are no flagged wells.



To group by C_T value

From the Group By drop-down menu, select **C_T**. The software groups the wells by C_T value: low, medium, high, and undetermined. In the example experiment, the C_T values are within the expected range (>8 and <35).



Tips for analyzing your own experiments

- **Replicate** – The software groups the wells by replicate: negative controls, standards, and samples. Look in the Quantity columns to make sure the quantity values for each replicate group are similar indicating tight C_T precision.
- **Flag** – The software groups the flagged and unflagged wells. A flag indicates that the software has found a potential error in the flagged well. For a description of the QuantStudio™ 6 and 7 Flex Real-Time PCR System Software flags, see "Flag Settings" on page 50.
- **C_T** – The threshold cycle (C_T) is the PCR cycle number at which the fluorescence level meets the threshold. A C_T value >8 and <35 is desirable. A C_T value <8 indicates that there is too much template in the reaction. A C_T value >35 indicates a low amount of target in the reaction; for C_T values >35, expect a higher standard deviation.

Confirm accurate dye signal using the Multicomponent Plot

The Multicomponent Plot screen displays the complete spectral contribution of each dye in a selected well over the duration of the PCR run.

Purpose

In the standard curve example experiment, you review the Multicomponent Plot screen for:

- ROX™ dye (passive reference)
- FAM™ dye (reporter)
- Spikes, dips, and/or sudden changes
- Amplification in the negative control wells

View the Multicomponent Plot

1. From the Experiment Menu pane, select **Analysis** ▸ **Multicomponent Plot**.

   Note: If no data are displayed, click **Analyze**.

2. Display the unknown and standard wells one at a time in the Multicomponent Plot screen:
   a. Click the **Plate Layout** tab.
   b. Select one well in the plate layout; the well is shown in the Multicomponent Plot screen.

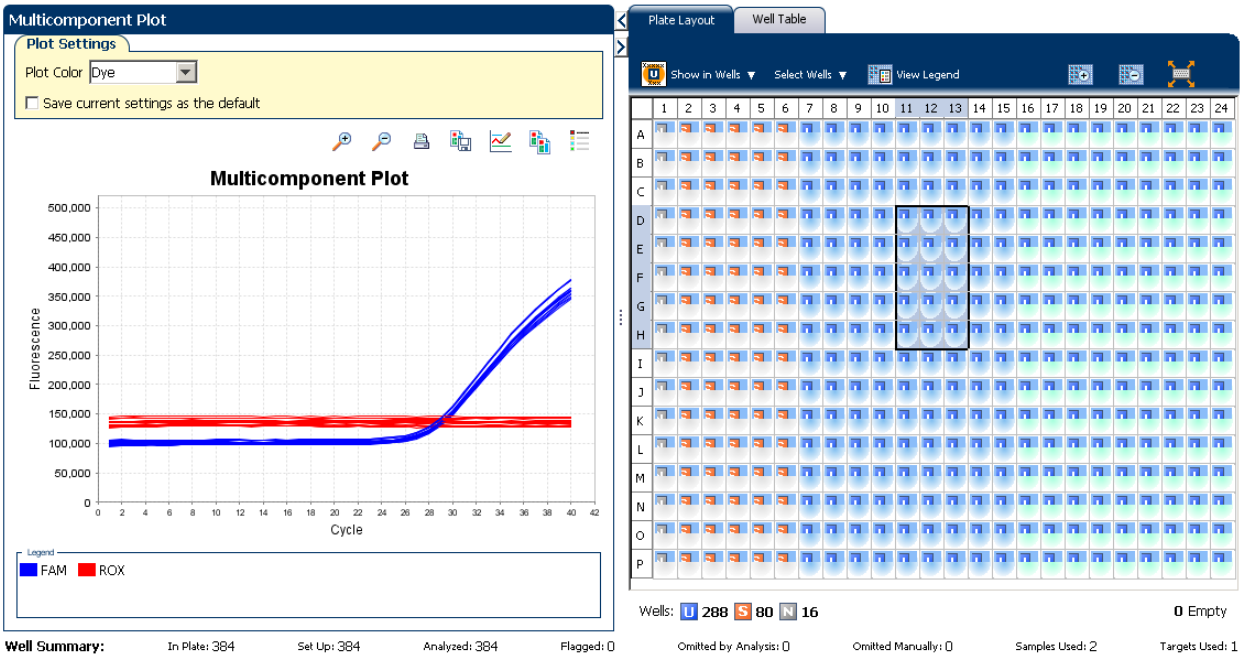
      Note: If you select multiple wells, the Multicomponent Plot screen displays the data for all selected wells simultaneously.

3. From the Plot Color drop-down menu, select **Dye**.
4. Click  **Show a legend for the plot** (default).

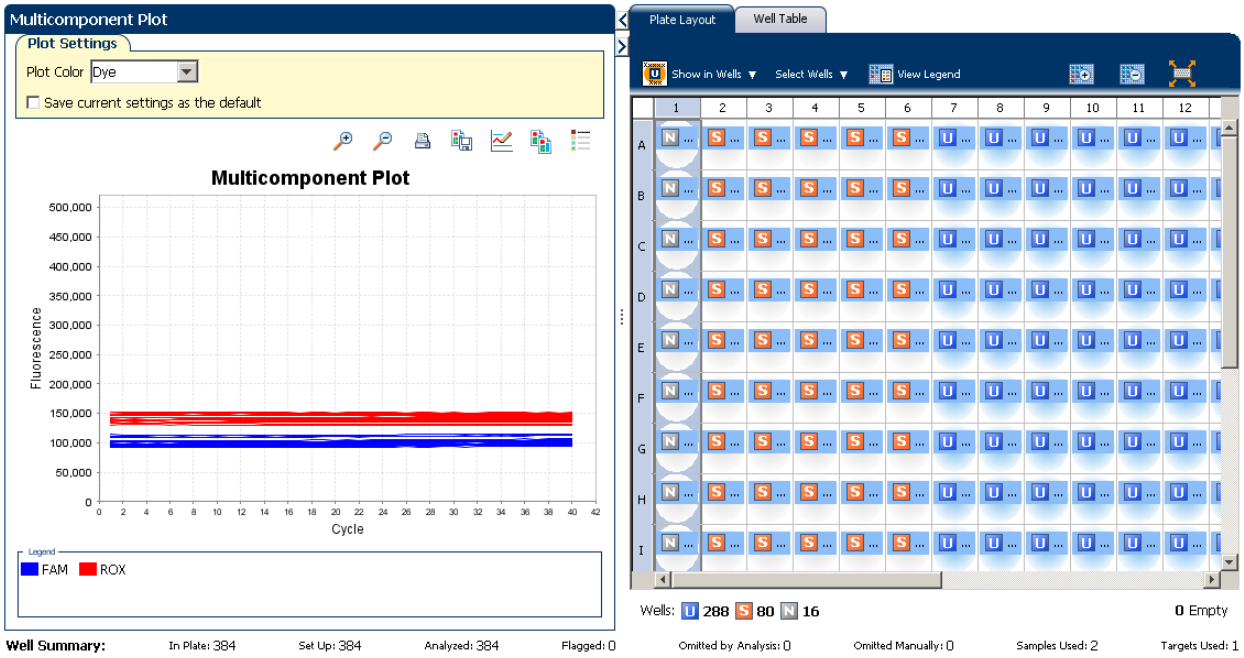
   Note: This is a toggle button. When the legend is displayed, the button changes to **Hide the plot legend**.

5. Check the ROX dye signal. In the example experiment, the ROX dye signal remains constant throughout the PCR process; a constant ROX dye signal indicates typical data.

6. Check the FAM dye signal. In the example experiment, the FAM dye signal increases throughout the PCR process; increase in FAM dye signal indicates normal amplification.



7. Select the negative control wells one at time and check for amplification. In the example experiment, there is no amplification in the negative control wells.



Tips for confirming dye accuracy in your own experiment

When you analyze your own standard curve experiment, look for:

- **Passive reference** – The passive reference dye fluorescence level should remain relatively constant throughout the PCR process.
- **Reporter dye** – The reporter dye fluorescence level should display a flat region corresponding to the baseline, followed by a rapid rise in fluorescence as the amplification proceeds.
- **Irregularities in the signal** – There should not be any spikes, dips, and/or sudden changes in the fluorescent signal.
- **Negative control wells** – There should not be any amplification in the negative control wells.

Determine signal accuracy using the Raw Data Plot

The Raw Data Plot screen displays the raw fluorescence signal (not normalized) for each optical filter for the selected wells during each cycle of the real-time PCR.

Purpose

In the standard curve example experiment, you review the Raw Data Plot screen for a stable increase in signal (no abrupt changes or dips) from the appropriate filter.

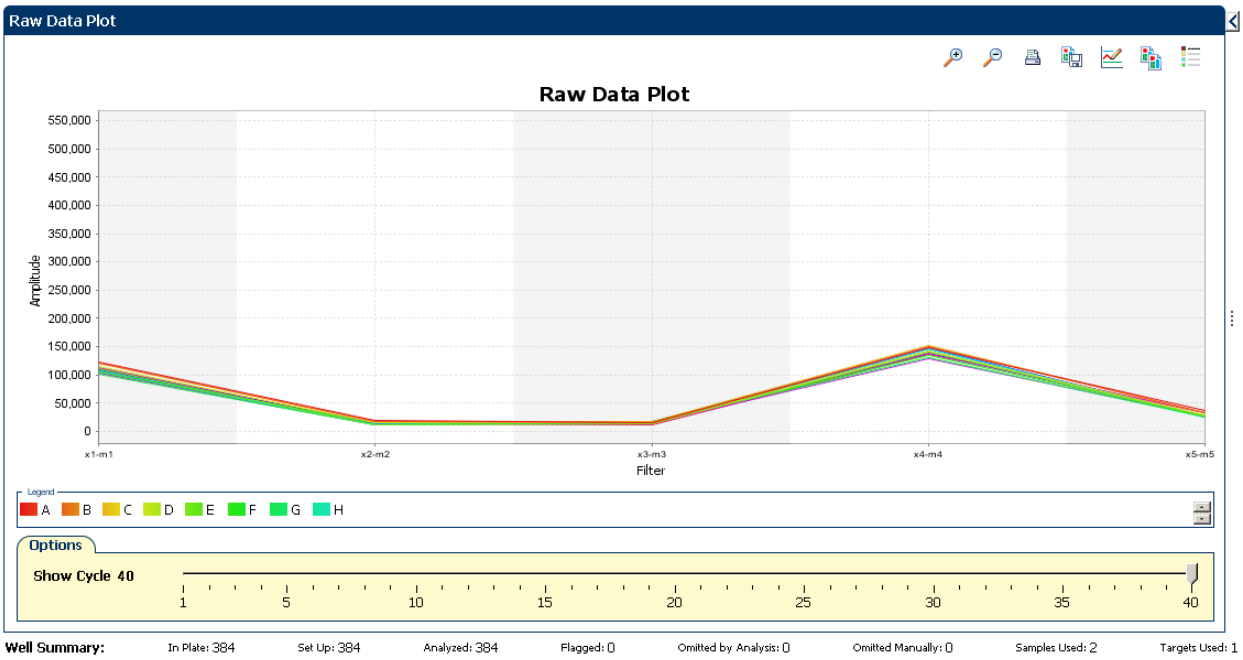
View the Raw Data Plot

1. From the Experiment Menu pane, select **Analysis ▸ Raw Data Plot**.

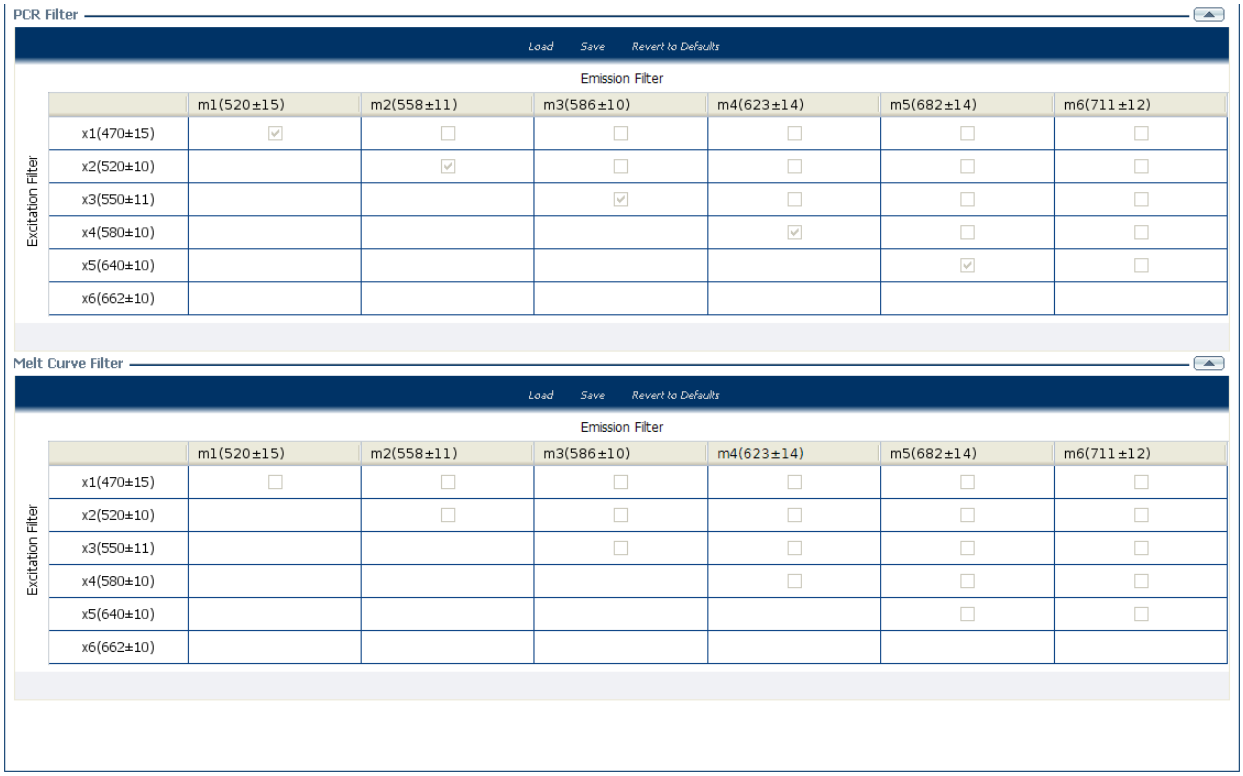
   Note: If no data are displayed, click **Analyze**.

2. Display all 384 wells in the Raw Data Plot screen by clicking the upper left corner of the plate layout in the Plate Layout tab.
3. Click  **Show a legend for the plot** (default). The legend displays the color code for each row of the reaction plate (see the legend in the Raw Data Plot shown below).

4. Click and drag the Show Cycle pointer from cycle 1 to cycle 40. In the example experiment, there is a stable increase in signal from filter 1, which corresponds to the FAM™ dye filter.



The filters used for the example experiment are:



Tips for determining signal accuracy in your own experiments

When you analyze your own standard curve experiment, look for the following in each filter:

- Characteristic signal growth
- No abrupt changes or dips

Review the flags in the QC Summary

The QC Summary screen displays a list of the QuantStudio™ 6 and 7 Flex Real-Time PCR System Software flags, including the flag frequency and location for the open experiment.

View the QC Summary

1. From the Experiment Menu pane, select **Analysis** ▸ **QC Summary**.

   Note: If no data are displayed, click **Analyze**.

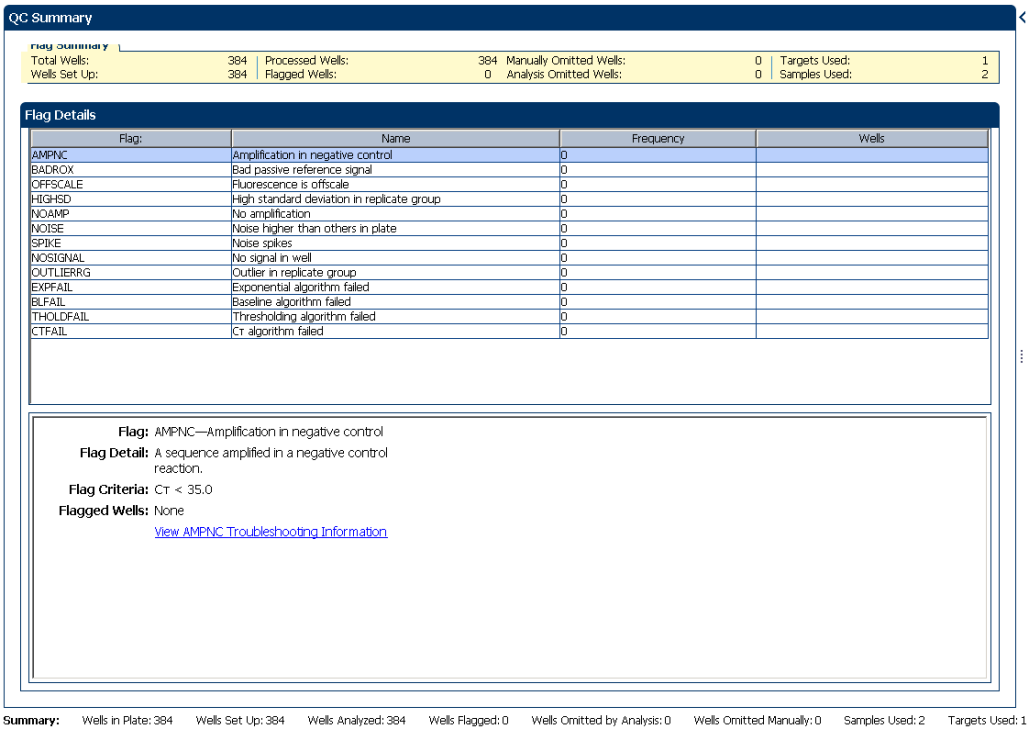
2. Review the Flags Summary.

   Note: A 0 displayed in the Frequency column indicates that the flag does not appear in the experiment. If the frequency is >0, the flag appears somewhere in the experiment; the well position is listed in the Wells column.

   In the example experiment, there are no flagged wells.

3. In the Flag Details table, click each flag with a frequency >0 to display detailed information about the flag. In the example experiment, the Frequency column displays 0 for all flags.

4. *(Optional)* For those flags with frequency >0, click the troubleshooting link to view information on correcting the flag.



Possible flags

The flags listed below may be triggered by the experiment data.

Note: To change the flag settings, refer to "Flag Settings" on page 50.

Flag	Description
Pre-processing flag	
OFFSCALE	Fluorescence is offscale
Primary analysis flags	
BADROX	Bad passive reference signal
NOAMP	No amplification
NOISE	Noise higher than others in plate
SPIKE	Noise spikes
NOSIGNAL	No signal in well
EXPFAIL	Exponential algorithm failed
BLFAIL	Baseline algorithm failed
THOLDFAIL	Thresholding algorithm failed
CTFAIL	C_T algorithm failed
Secondary analysis flags	
OUTLIERRG	Outlier in replicate group
AMPNC	Amplification in negative control
HIGHSD	High standard deviation in replicate group

For more information

For more information on...	Refer to...	Publication number
Publishing data	Chapter 2 in Booklet 1, *Getting Started with QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Experiments*	4489822

Section 5.2 Adjust parameters for re-analysis of your own experiments

Adjust analysis settings

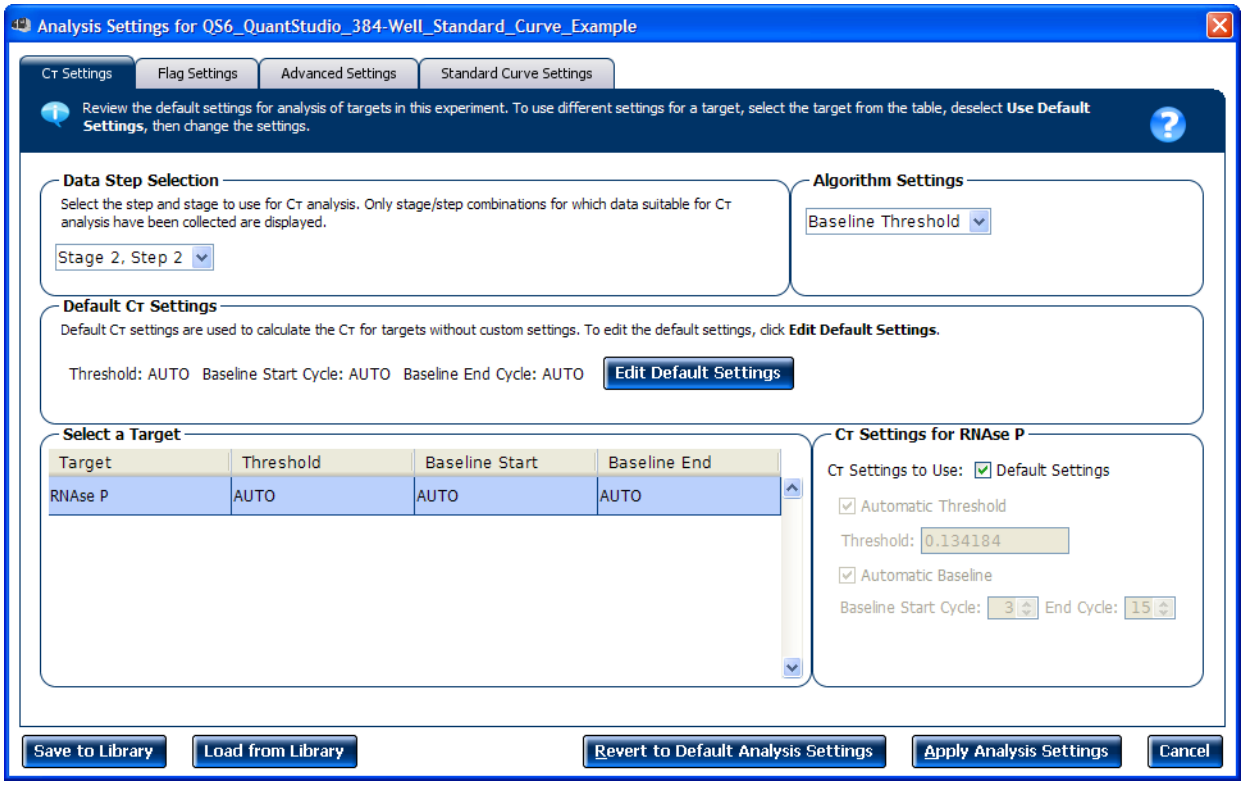
The Analysis Settings dialog box displays the analysis settings for the threshold cycle (C_T), flags, and advanced options.

If the default analysis settings in the QuantStudio™ 6 and 7 Flex Real-Time PCR System Software are not suitable for your own experiment, you can change the settings in the Analysis Settings dialog box, then reanalyze your experiment.

View the analysis settings

1. From the Experiment Menu pane, select **Analysis**.
2. Click **Analysis** ▸ **Analysis Settings** to open the Analysis Settings dialog box.
   In the example experiment, the default analysis settings are used for each tab:
   - C_T Settings
   - Flag Settings
   - Advanced Settings
   - Standard Curve Settings

   The following is an image of the Analysis Settings dialog box for a Standard Curve experiment:



3. View and, if necessary, change the analysis settings (see "Adjust analysis settings" below).

   Note: You can save the changes to the analysis settings to the Analysis Settings Library for later use. For more information, see About the Analysis Settings Library in Booklet 1, *Getting Started with QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Experiments*.

4. Click **Apply Analysis Settings** to apply the current analysis settings.

   Note: You can go back to the default analysis settings, by clicking **Revert to Default Analysis Settings**.

Adjust analysis settings

C_T Settings

- **Data Step Selection**

  Use this feature to select one stage/step combination for C_T analysis when there is more than one data collection point in the run method.

- **Algorithm Settings**

  Use the Baseline Threshold algorithm to determine the C_T values.

  The Baseline Threshold algorithm is an expression estimation algorithm that subtracts a baseline component and sets a fluorescent threshold in the exponential region for gene quantification.

- **Default C_T Settings**

  Use the default C_T settings feature to calculate C_T for the targets that do not have custom settings. To edit the default settings, click **Edit Default Settings**.

- **C_T Settings for Target**

  When you manually set the threshold and baseline, Life Technologies recommends:

Setting	Recommendation
Threshold	Enter a value for the threshold so that the threshold is: • Above the background. • Below the plateau and linear regions of the amplification curve. • Within the exponential phase of the amplification curve.
Baseline	Select the Start Cycle and End Cycle values so that the baseline ends before significant fluorescent signal is detected.

  Note: This setting is applicable only to the Baseline Threshold algorithm.

  Note: Selecting Automatic Threshold implies selection of automatic setting of the baseline. However, if Automatic Threshold is deselected, then you can choose between setting the baseline either automatically or manually.

Flag Settings

Use the Flag Settings tab to:

- Adjust the sensitivity so that more wells or fewer wells are flagged.
- Change the flags that are applied by the QuantStudio™ 6 and 7 Flex Real-Time PCR System Software.

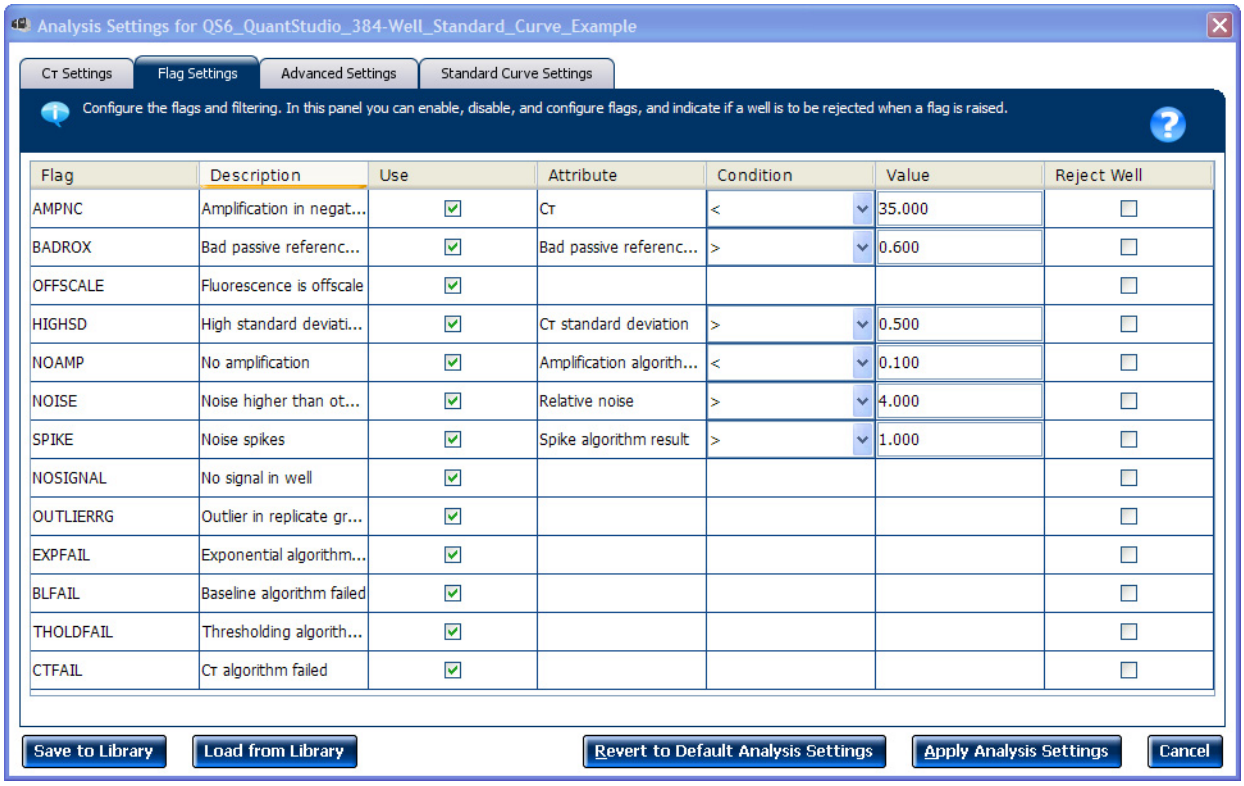
To adjust the flag settings

1. In the Use column, select the check boxes for flags to apply during analysis.
2. *(Optional)* If an attribute, condition, and value are listed for a flag, specify the setting for applying the flag.

   Note: If you choose to adjust the setting for applying a flag, make minor adjustments as you evaluate the appropriate setting.
3. In the Reject Well column, select the check boxes if you want the software to reject wells with the flag.

   Note: After you have rejected the flagged wells, analysis results depend on factors such as the experiment type and flag type. For example, rejecting wells flagged by HIGHSD in experiments using the Standard Deviation calculations may change the result of C_T SD. For some flags, analysis results calculated before the well is rejected are maintained.
4. Click **Apply Analysis Settings** in the Analysis Settings dialog box. If the run status is complete, the data are reanalyzed.

   The following is an image of the Flag Settings tab:



Advanced Settings

Use the Advanced Settings tab to change baseline settings well-by-well.

To use custom baseline settings for a well-target combination:

1. Select one or more well-target combinations in the table.
2. Deselect the **Use C_T Settings Defined for Target** check box.

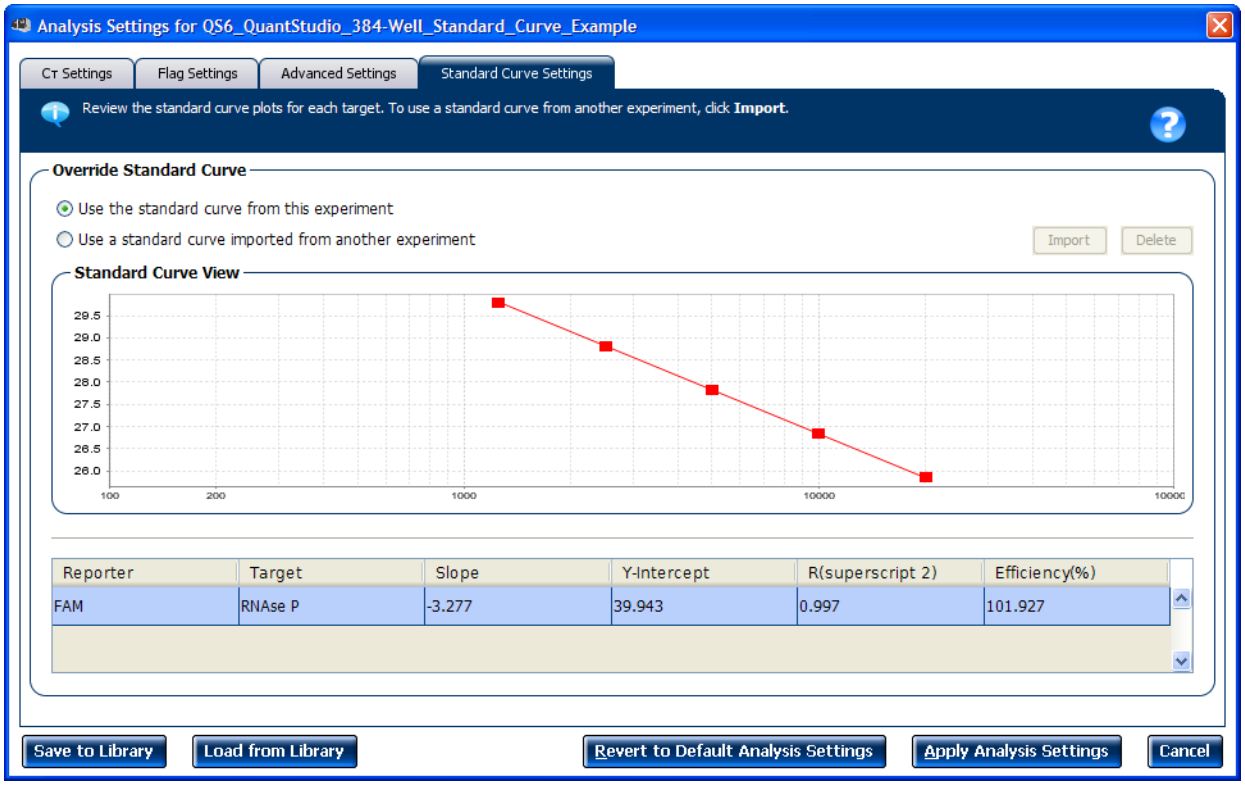
3. Define the custom baseline settings:
   - For automatic baseline calculations, select the **Automatic Baseline** check box.
   - To define the baseline, deselect the **Automatic Baseline** check box, then enter the baseline start cycle and baseline end cycle.

Standard Curve Settings

Use this tab to review the settings of the current standard curve experiment or to import the standard curve from an external experiment (with the same samples and targets) and apply it to the current experiment.

Note: The run method must be the same. Life Technologies recommends using the standard curve from the current experiment.

For the example experiment, the settings from the current experiment have been used.



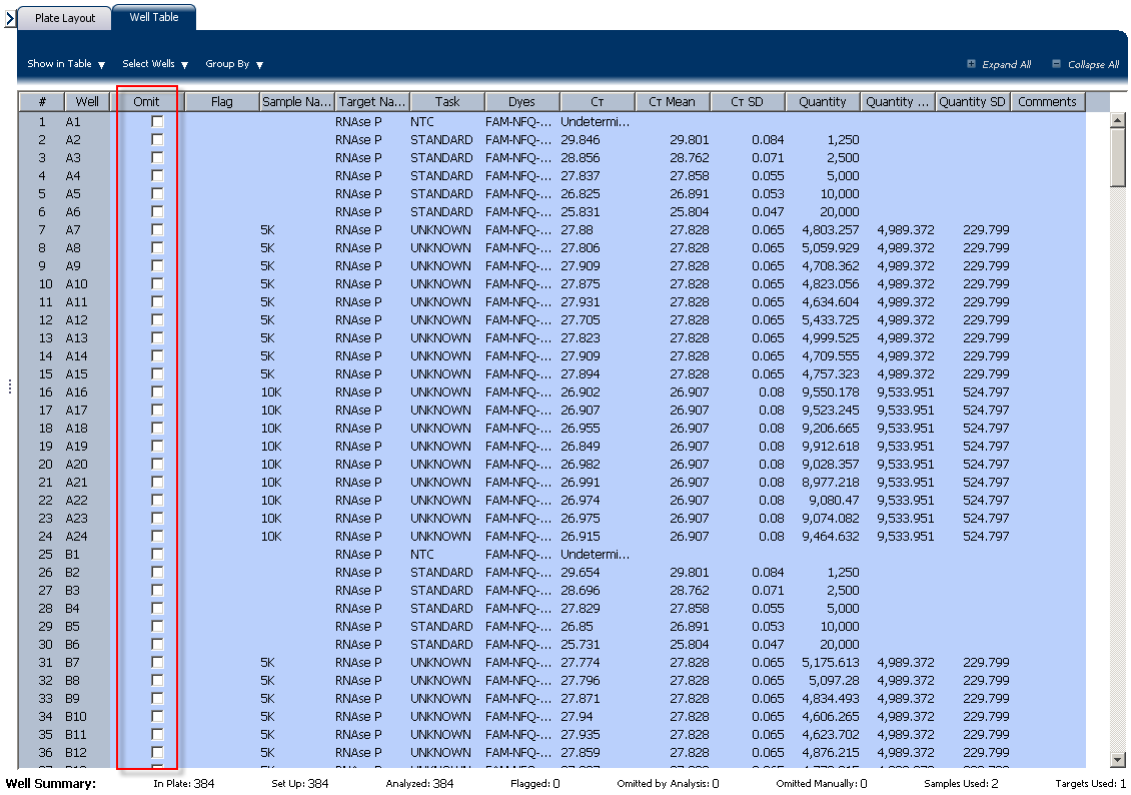
Improve C_T precision by omitting wells

Experimental error may cause some wells to be amplified insufficiently or not at all. These wells typically produce C_T values that differ significantly from the average for the associated replicate wells. If included in the calculations, these outliers can result in erroneous measurements; to ensure C_T precision, omit the outliers from the analysis.

1. From the Experiment Menu pane, select **Analysis** ▸ **Amplification Plot**.

   Note: If no data are displayed, click **Analyze**.

2. In the Amplification Plot screen, select **C_T vs Well** from the Plot Type drop-down menu.
3. Select the **Well Table** tab.
4. In the Well Table, view outliers:
   a. From the Group By drop-down menu, select **Replicate**.
   b. Look for any outliers in the replicate group (make sure they are flagged).
   c. Select the **Omit** check box next to outlying well(s), as shown in the following image.



#	Well	Omit	Flag	Sample Na...	Target Na...	Task	Dyes	CT	CT Mean	CT SD	Quantity	Quantity ...	Quantity SD	Comments
1	A1	☐			RNAse P	NTC	FAM-NFQ-...	Undetermi...						
2	A2	☐			RNAse P	STANDARD	FAM-NFQ-...	29.846	29.801	0.084	1,250			
3	A3	☐			RNAse P	STANDARD	FAM-NFQ-...	28.856	28.762	0.071	2,500			
4	A4	☐			RNAse P	STANDARD	FAM-NFQ-...	27.837	27.858	0.055	5,000			
5	A5	☐			RNAse P	STANDARD	FAM-NFQ-...	26.825	26.891	0.053	10,000			
6	A6	☐			RNAse P	STANDARD	FAM-NFQ-...	25.831	25.804	0.047	20,000			
7	A7	☐		5K	RNAse P	UNKNOWN	FAM-NFQ-...	27.88	27.828	0.065	4,803.257	4,989.372	229.799	
8	A8	☐		5K	RNAse P	UNKNOWN	FAM-NFQ-...	27.806	27.828	0.065	5,059.929	4,989.372	229.799	
9	A9	☐		5K	RNAse P	UNKNOWN	FAM-NFQ-...	27.909	27.828	0.065	4,708.362	4,989.372	229.799	
10	A10	☐		5K	RNAse P	UNKNOWN	FAM-NFQ-...	27.875	27.828	0.065	4,823.056	4,989.372	229.799	
11	A11	☐		5K	RNAse P	UNKNOWN	FAM-NFQ-...	27.931	27.828	0.065	4,634.604	4,989.372	229.799	
12	A12	☐		5K	RNAse P	UNKNOWN	FAM-NFQ-...	27.705	27.828	0.065	5,433.725	4,989.372	229.799	
13	A13	☐		5K	RNAse P	UNKNOWN	FAM-NFQ-...	27.823	27.828	0.065	4,999.525	4,989.372	229.799	
14	A14	☐		5K	RNAse P	UNKNOWN	FAM-NFQ-...	27.909	27.828	0.065	4,709.555	4,989.372	229.799	
15	A15	☐		5K	RNAse P	UNKNOWN	FAM-NFQ-...	27.894	27.828	0.065	4,757.323	4,989.372	229.799	
16	A16	☐		10K	RNAse P	UNKNOWN	FAM-NFQ-...	26.902	26.907	0.08	9,550.178	9,533.951	524.797	
17	A17	☐		10K	RNAse P	UNKNOWN	FAM-NFQ-...	26.907	26.907	0.08	9,523.245	9,533.951	524.797	
18	A18	☐		10K	RNAse P	UNKNOWN	FAM-NFQ-...	26.955	26.907	0.08	9,206.665	9,533.951	524.797	
19	A19	☐		10K	RNAse P	UNKNOWN	FAM-NFQ-...	26.849	26.907	0.08	9,912.618	9,533.951	524.797	
20	A20	☐		10K	RNAse P	UNKNOWN	FAM-NFQ-...	26.982	26.907	0.08	9,028.357	9,533.951	524.797	
21	A21	☐		10K	RNAse P	UNKNOWN	FAM-NFQ-...	26.991	26.907	0.08	8,977.218	9,533.951	524.797	
22	A22	☐		10K	RNAse P	UNKNOWN	FAM-NFQ-...	26.974	26.907	0.08	9,080.47	9,533.951	524.797	
23	A23	☐		10K	RNAse P	UNKNOWN	FAM-NFQ-...	26.975	26.907	0.08	9,074.082	9,533.951	524.797	
24	A24	☐		10K	RNAse P	UNKNOWN	FAM-NFQ-...	26.915	26.907	0.08	9,464.632	9,533.951	524.797	
25	B1	☐			RNAse P	NTC	FAM-NFQ-...	Undetermi...						
26	B2	☐			RNAse P	STANDARD	FAM-NFQ-...	29.654	29.801	0.084	1,250			
27	B3	☐			RNAse P	STANDARD	FAM-NFQ-...	28.696	28.762	0.071	2,500			
28	B4	☐			RNAse P	STANDARD	FAM-NFQ-...	27.829	27.858	0.055	5,000			
29	B5	☐			RNAse P	STANDARD	FAM-NFQ-...	26.85	26.891	0.053	10,000			
30	B6	☐			RNAse P	STANDARD	FAM-NFQ-...	25.731	25.804	0.047	20,000			
31	B7	☐		5K	RNAse P	UNKNOWN	FAM-NFQ-...	27.774	27.828	0.065	5,175.613	4,989.372	229.799	
32	B8	☐		5K	RNAse P	UNKNOWN	FAM-NFQ-...	27.796	27.828	0.065	5,097.28	4,989.372	229.799	
33	B9	☐		5K	RNAse P	UNKNOWN	FAM-NFQ-...	27.871	27.828	0.065	4,834.493	4,989.372	229.799	
34	B10	☐		5K	RNAse P	UNKNOWN	FAM-NFQ-...	27.94	27.828	0.065	4,606.265	4,989.372	229.799	
35	B11	☐		5K	RNAse P	UNKNOWN	FAM-NFQ-...	27.935	27.828	0.065	4,623.702	4,989.372	229.799	
36	B12	☐		5K	RNAse P	UNKNOWN	FAM-NFQ-...	27.859	27.828	0.065	4,876.215	4,989.372	229.799	

5. Click **Analyze** to reanalyze the experiment data with the outlying well(s) removed from the analysis.

   Note: You can also omit undesirable wells in an experiment from the Plate Layout screen. To omit a well from the Plate Layout screen, right-click the well and select **Omit**.

For more information

For more information on	Refer to	Publication number
Amplification efficiency	*Amplification Efficiency of TaqMan® Gene Expression Assays Application Note*.	127AP05-03

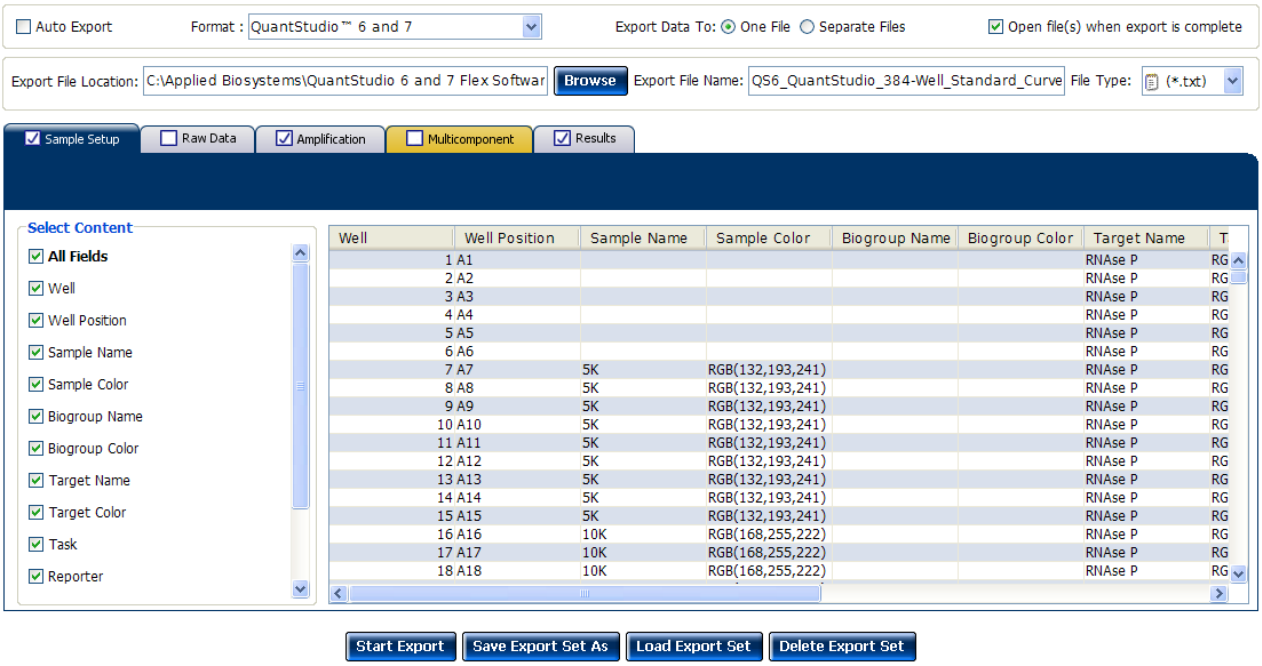
6 Export Experiment Results

1. Open the Standard Curve example experiment file that you analyzed in Chapter 5.
2. In the Experiment Menu, click  **Export.**

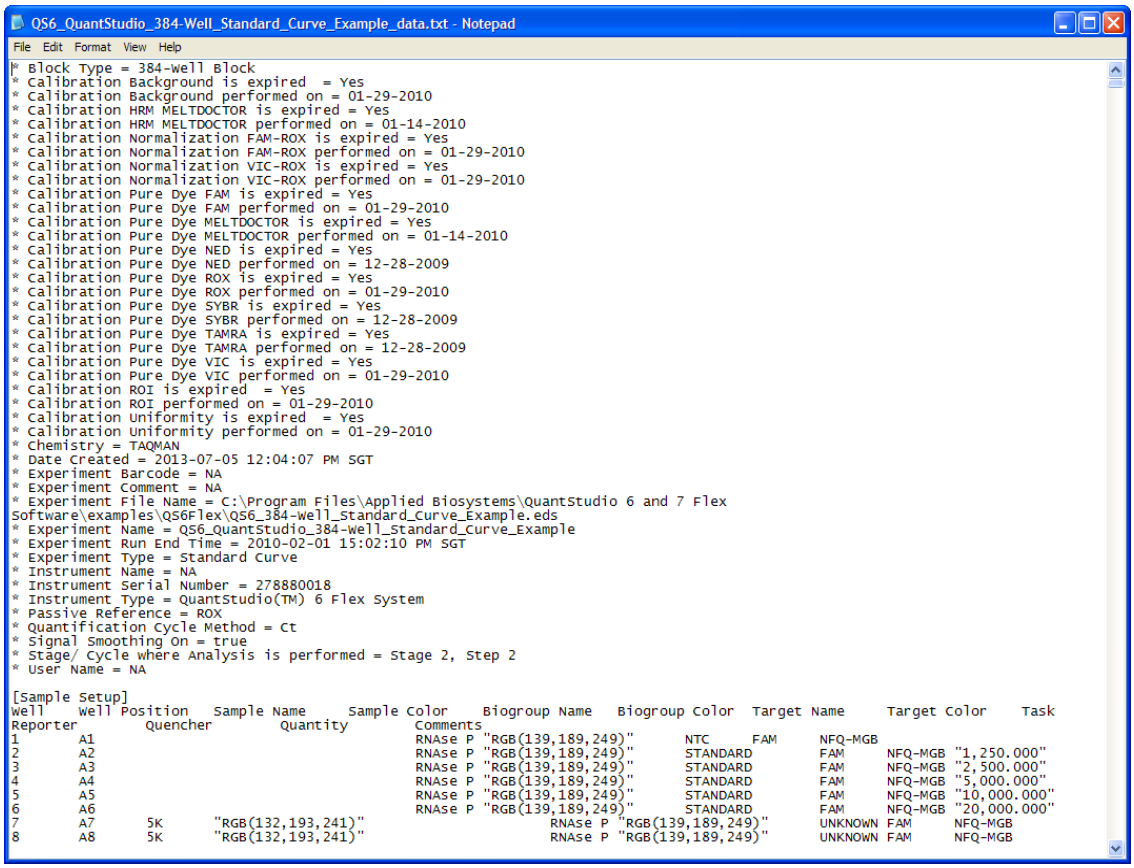
   Note: To export data automatically after analysis, select the **Auto Export** check box during experiment setup or before running the experiment. Auto export is unchecked for the example experiment.
3. Select **QuantStudio™ 6 and 7** format.
4. Complete the Export dialog box as shown below:

Field or Selection	Entry
Select Data to export/ Select Content	Results
Export Data To	One File
Export File Name	QS6_QuantStudio_384-Well_Standard_Curve_Example_data
File Type	*.txt
Export File Location	C:\Applied Biosystems\QuantStudio 6 and 7 Flex Software\User Files\Export

Your Export screen should look like this:



Your exported file when opened in Notepad should look like this:



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USER GUIDE

applied biosystems®
by life technologies™

QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Getting Started Guide for Relative Standard Curve and Comparative C_T Experiments

Booklet 3

Publication Number 4489822
Revision A

life technologies™

Contents

PART 1
Running Relative Standard Curve Experiments

1 About Relative Standard Curve Experiments

This chapter covers:

IMPORTANT! First-time users of the QuantStudio™ 6 and 7 Flex Real-Time PCR System Software, please read Booklet 1, *Getting Started with QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Experiments* and Booklet 7, *QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Experiments - Appendixes* of this binder thoroughly. The booklets provide information and general instructions that are applicable to all the experiments described in this binder.

Note: For more information about any of the topics discussed in this guide, access the Help from within QuantStudio™ 6 and 7 Flex Real-Time PCR System Software by pressing **F1**, clicking in the toolbar, or selecting **Help ▸ QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Help**.

About Relative Standard Curve experiments

The Relative Standard Curve method is used to determine relative target quantity in samples. The QuantStudio™ 6 and 7 Flex Real-Time PCR System Software measures amplification of the target and of the endogenous control in samples, in a reference sample, and in a standard dilution series. Measurements are normalized using the endogenous control. Data from the standard dilution series are used to generate the standard curve. Using the standard curve, the software interpolates target quantity and endogenous control quantity in the samples and the reference sample. For each sample and reference sample, the target quantity is normalized by endogenous control quantity (quantity of target/quantity of endogenous control). The normalized quotient from samples is divided by the quotient from the reference sample to get relative quantification (fold change). The software determines the relative quantity of target in each sample by comparing target quantity in each sample to target quantity in the reference sample.

Relative Standard Curve experiments are commonly used to:

- Compare expression levels of a gene in different tissues.
- Compare expression levels of a gene in a treated sample and an untreated sample.
- Compare expression levels of wild-type alleles and mutated alleles.
- Analyze the gene expression changes over time under specific treatment conditions.

The Comparative CT ($\Delta\Delta C_T$) method is used to determine the relative target quantity in samples. With the comparative C_T method, the QuantStudio™ 6 and 7 Flex Real-Time PCR System Software measures amplification of the target and of the endogenous control in samples and in a reference sample. For more information on Comparative CT ($\Delta\Delta C_T$) experiments, refer to Part II, Running Comparative CT ($\Delta\Delta C_T$) Experiments of this booklet.

Assemble required components

- **Sample** – The tissue group that you are testing for a target gene.
- **Reference sample (also called a calibrator)** – The sample used as the basis for relative quantification results. For example, in a study of drug effects on gene expression, an untreated control is an appropriate reference sample.
- **Standard** – A sample that contains known quantities of the target; used in quantification experiments to generate standard curves.
- **Standard dilution series** – A set of standards containing a range of known quantities. The standard dilution series is prepared by serially diluting standards.
- **Endogenous control** – A gene that is used to normalize template input differences, and sample-to-sample or run-to-run variation.
- **Replicates** – The total number of identical reactions containing identical components and identical volumes.
- **Negative controls** – Wells that contain water or buffer instead of sample template. No amplification of the target should occur in negative control wells.

PCR options

When performing real-time PCR, choose between:

- Singleplex and multiplex PCR (below)

  and

- 1-step and 2-step RT-PCR (page 13)

Singleplex and Multiplex PCR

You can perform a PCR reaction using either:

- **Singleplex PCR** – In singleplex PCR a single primer and probe set is present in the reaction tube or well. Only one target or endogenous control can be amplified per reaction.

  Or

- **Multiplex PCR** – In multiplex PCR, two or more primer and probe sets are present in the reaction tube or well. Each set amplifies a specific target or endogenous control. Typically, a probe labeled with FAM™ dye detects the target and a probe labeled with VIC® dye detects the endogenous control.

IMPORTANT! SYBR® Green reagents cannot be used for multiplex PCR.

1- and 2-Step RT-PCR

You can perform reverse transcription (RT) and PCR in a single reaction (1-step) or in separate reactions (2-step). The reagent configuration you use depends on whether you are performing 1- or 2-step RT-PCR:

- **1-step PCR**– In 1-step RT-PCR, RT and PCR take place in one buffer system. Using one buffer system provides the convenience of a single-tube preparation for RT and PCR amplification. However, you cannot use Fast PCR Master Mix or the carryover prevention enzyme, AmpErase® UNG (uracil-N-glycosylase), to perform 1-step RT-PCR.
- **2-step PCR**– 2-step RT-PCR is performed in two separate reactions: First, total RNA is reverse-transcribed into cDNA, then the cDNA is amplified by PCR. This method is useful for detecting multiple transcripts from a single cDNA template or for storing cDNA aliquots for later use. The AmpErase® UNG enzyme can be used to prevent carryover contamination.

About the example experiment

To illustrate how to perform a Relative Standard Curve, this guide leads you through the process of designing and performing an example experiment. The example experiment represents a typical setup that you can use to quickly familiarize yourself with the QuantStudio™ 6 and 7 Flex Real-Time PCR System Software.

The objective of the Relative Standard Curve example experiment is to compare the expression of the FAS transcriptional factor (an oncoprotein that activates the transcription of growth-associated genes) in Human cDNA tissues.

In the Relative Standard Curve example experiment:

- The samples are kidney, liver, brain, and heart.
- The target is FAS.
- The endogenous control is HPRT.
- The reference sample is brain.
- One standard curve is set up for FAS. The standard used for the standard dilution series is a Human cDNA sample of known total concentration.
- One standard curve is set up for HPRT (endogenous control). The standard used for the standard dilution series is a Human Male Raji cDNA sample of known total concentration.
- Reactions are set up for **2-step RT-PCR**. The Invitrogen VILO Kit is used for reverse transcription; the TaqMan® Gene Expression Master Mix (2✕) is used for PCR.
- Select primer and probe sets from the Life Technologies TaqMan® Gene Expression Assays product line:
  - For the target assay (FAS), select assay ID Hs00907759_m1.
  - For the endogenous control assay (HPRT), select assay ID Hs99999909_m1.

2 Design the Experiment

This chapter explains how to design the example experiment from the Experiment Setup menu.

This chapter covers:

Note: To automatically export the analyzed data to a specified location, select the Auto Export task from the Export screen, before running the experiment. For more information on Auto Export, refer to Chapter 2 in Booklet 1, *Getting Started with QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Experiments*.

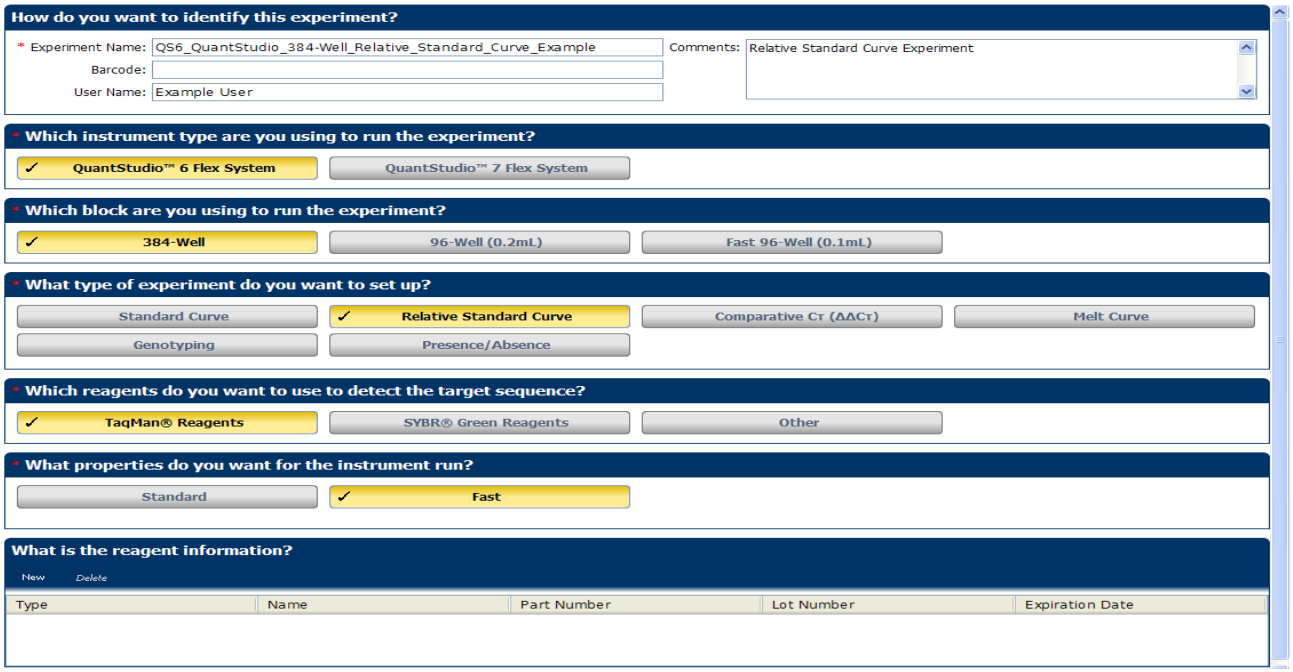
Define the experiment properties

Click **Experiment Setup** ▸ **Experiment Properties** to create a new experiment in the QuantStudio™ 6 and 7 Flex Software. Enter:

Field or Selection	Entry
Experiment Name	QS6_QuantStudio_384-Well_Relative_Standard_Curve_Example
Barcode	Leave field empty
User Name	Example User
Comments	Relative Standard Curve example
Instrument type	QuantStudio™ 6 Flex System
Block	384-Well Block
Experiment Type	Relative Standard Curve
Reagents	TaqMan® Reagents
Ramp speed	Standard
Reagent information	NA

Save the experiment.

Your Experiment Properties screen should look like this:



Define targets, samples, and biological replicates

Click **Define** to access the Define screen. Enter:

1. Targets

Target name	Reporter	Quencher	Color
FAS	FAM	NFQ-MGB	
HPRT	VIC	NFQ-MGB	

2. Samples

Sample name	Color
Brain	
Heart	
Kidney	
Liver	

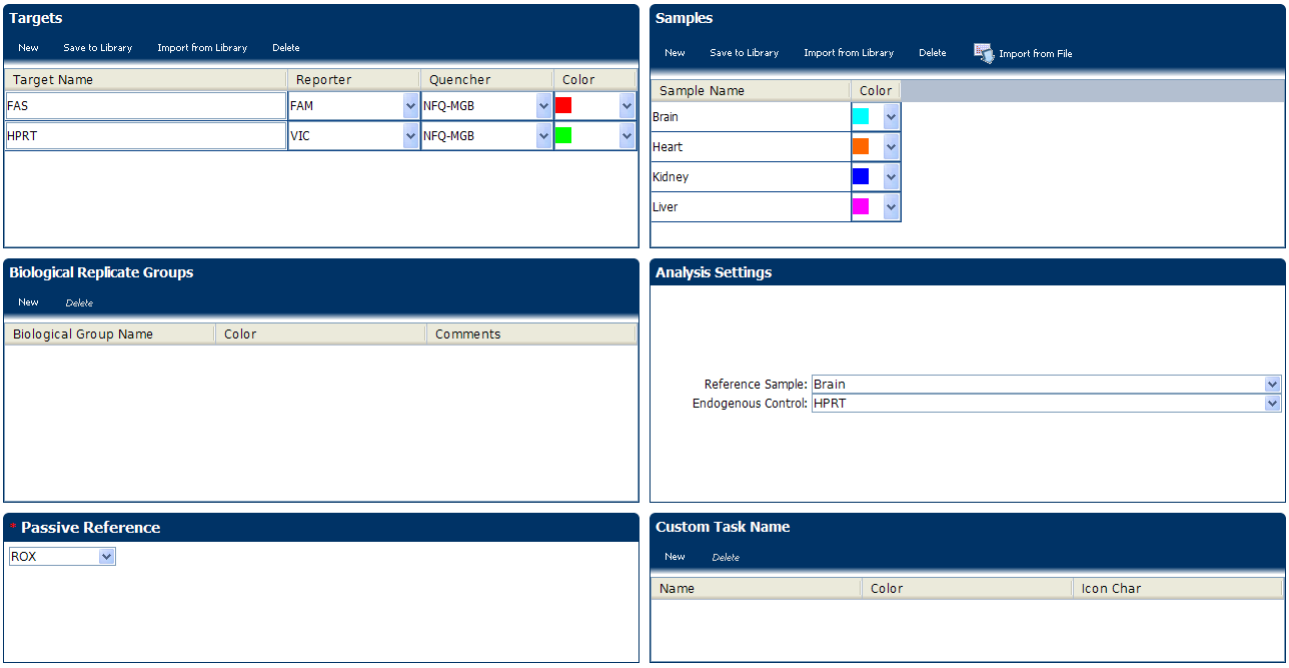
3. Dye to be used as a Passive Reference
   ROX

4. Custom Task Name
   Not applicable

5. Analysis Settings

Field	Select
Reference Sample	Brain
Endogenous Control	HPRT

Your Define screen should look like this:



Note: This example experiment does not define biological replicate groups. Leave Biological Replicate Groups blank.

Assign targets, samples, and biological groups

Click **Assign** to access the Assign screen.

Note: To automatically set up and assign standards, click 

to open the Define and Set Up Standards dialog box.

To assign the targets and samples:

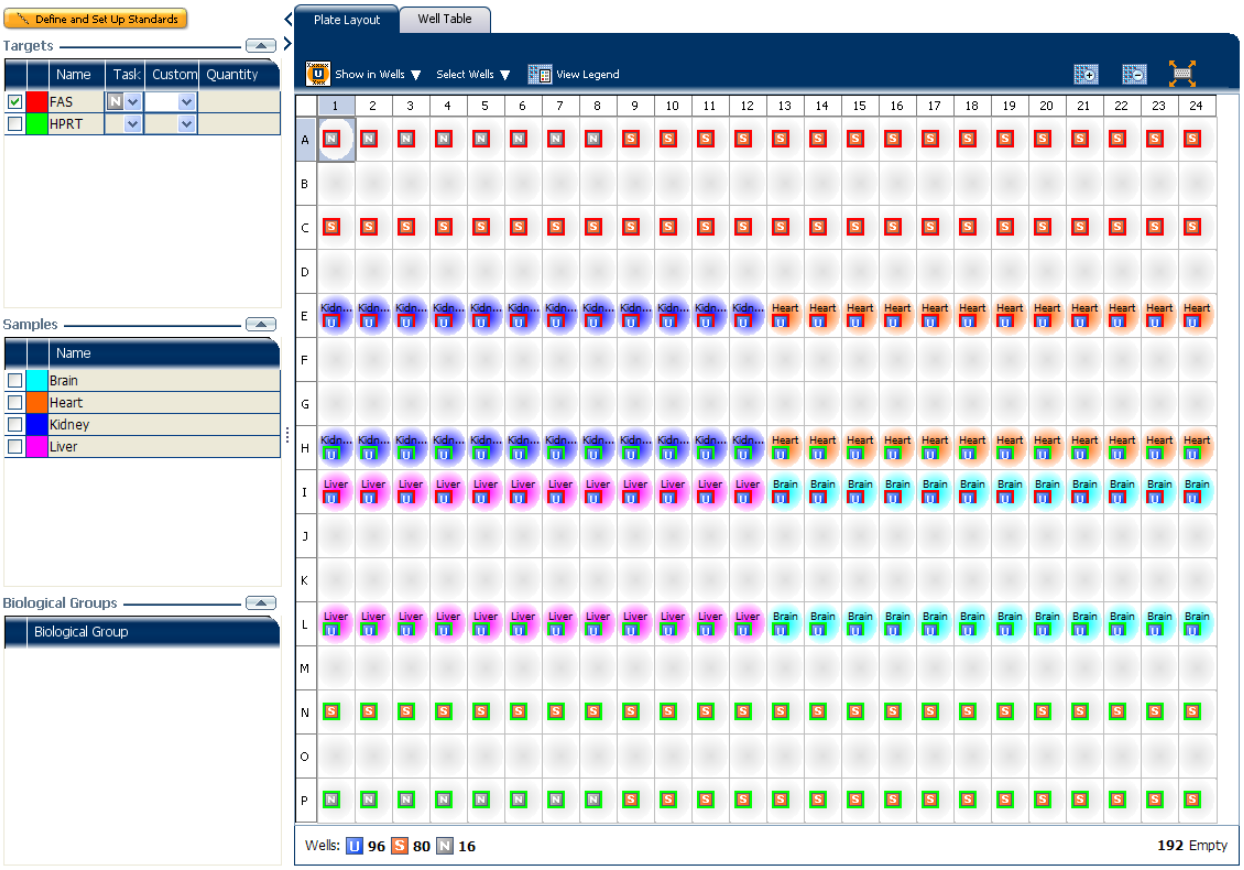
1. Set up the standards.
2. For the first standard for the FAS target:
   a. Click-drag to select wells A9-A16.
   b. Check check box next to FAS in the Targets list.
   c. Select S in the Task drop-down menu.
   d. Enter 625 in the Quantity column.

e. Repeat steps a through c for each of the standards for the FAS target, selecting the wells listed in the table below, and entering the corresponding quantity.

3. Repeat step 2 for each standard for the HPRT target.

Target name	Well number	Task	Quantity	Sample name
FAS	A1 - A8	Negative	None	None
	E1 - E12	Unknown	Determined by run	Kidney
	E13 - E24	Unknown	Determined by run	Heart
	I1 - 112	Unknown	Determined by run	Liver
	I13 - I24	Unknown	Determined by run	Brain
	A9 - A16	Standard	625	None
	A17 - A24	Standard	2,500	None
	C1 - C8	Standard	312.50	None
	C9 - C16	Standard	1,250	None
	C17 - C24	Standard	5,000	None
HPRT	P1 - P8	Negative	None	None
	H1 - H12	Unknown	Determined by run	Kidney
	H13 - H 24	Unknown	Determined by run	Heart
	L1 - L12	Unknown	Determined by run	Liver
	L13 - L24	Unknown	Determined by run	Brain
	N1 - N8	Standard	165.25	None
	N9 - N16	Standard	625	None
	N17 - N24	Standard	2,500	None
	P9 - P16	Standard	312.50	None
	P17 - P24	Standard	1,250	None

Your Assign screen should look like this:



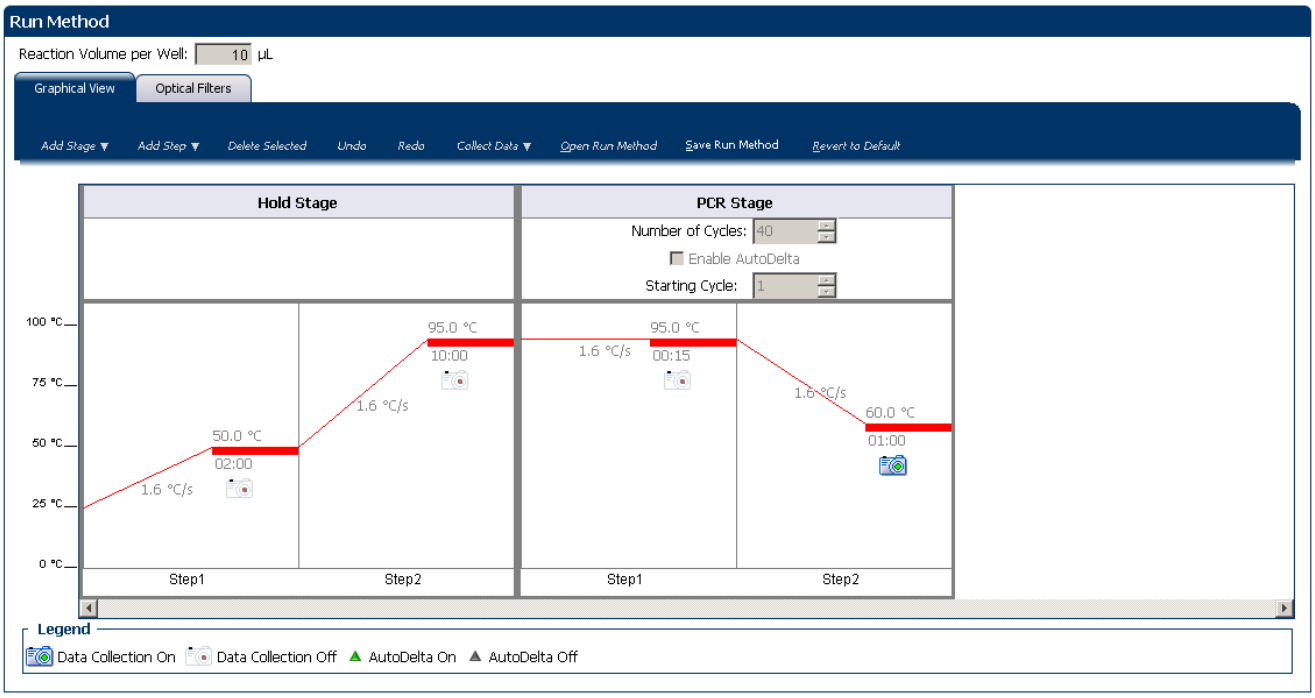
Set up the run method

Click **Run Method** to access the Run Method screen. Set the thermal profile under the Graphical View tab. Enter:

- Reaction Volume Per Well: 10 µL
- Thermal Profile

Stage	Step	Ramp rate	Temperature	Time
Hold Stage	Step 1	1.6°C/s	50°C	2 minutes
	Step 2	1.6°C/s	95°C	10 minutes
PCR Stage	Step 1	1.6°C/s	95°C	15 seconds
Number of Cycles: 40 Enable AutoDelta: Unchecked (default) Starting Cycle: Disabled when Enable AutoDelta is unchecked	Step 2	1.6°C/s	60°C	1 minute

Your Run Method screen should look like this:



Tips for designing your own experiment

Life Technologies recommends that you:

- Set up a standard curve for each target assay in the reaction plate.
- Identify each target assay with a unique name and color. You can enter up to 100 characters in the Target Name field.
- Identify each sample using a unique name and color. You can enter up to 100 characters in the Sample Name field.
- Select an endogenous control for each sample. The endogenous control is a target that is present in all samples under investigation. It should be expressed equally in all sample types, regardless of treatment or tissue origin (examples of endogenous controls are β-actin, GAPDH, and 18S ribosomal RNA [18S rRNA]). The endogenous control is used to normalize the PCR results; the endogenous control corrects for variable sample mass, nucleic acid extraction efficiency, reverse transcription efficiency, and pipette calibration errors.

  Note: Each sample type (for example, each tissue in a study comparing multiple tissues) requires an endogenous control.
- Enter at least five dilution points for each standard curve in the reaction plate.
- Enter at least three replicates (identical reactions) for each point in the standard curve and for each sample reaction.
- Enter at least three negative control reactions for each target assay.

- Carefully consider the appropriate range of standard quantities for your assay because the range of standard quantities affects the amplification efficiency calculations:
  - For more accurate measurements of amplification efficiency, use a broad range of standard quantities, spanning between 5 and 6 logs. If you do so, use a PCR product or a highly concentrated template, such as a cDNA clone.
  - If you have a limited amount of cDNA template and/or if the target is a low-copy number transcript, or known to fall within a given range, a narrow range of standard quantities may be necessary.
- Minimally run a five-point curve of 1:10 dilutions to minimize the effects of small pipetting errors.
- Select a reference sample from your previously defined samples. Amplification results from the samples and from the reference sample are compared to determine relative expression.
- Select an endogenous control from your previously defined target assays. Amplification results from the endogenous control are used to normalize the amplification results from the target for differences in the amount of template added to each reaction.

For more information

For more information on...	Refer to...	Publication number
Consumables	Chapter 1 in Booklet 1, *Getting Started with QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Experiments* Appendix A in Booklet 7, *QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Experiments - Appendixes*	4489822
Using other quantification methods	Booklet 2, *Running Standard Curve Experiments* and Part 2 of Booklet 3, *Running Relative Standard Curve and Comparative C_T Experiments*.	4489822
Amplification efficiency	*Amplification Efficiency of TaqMan® Gene Expression Assays Application Note*	127AP05-03
Using Alternative Setup	Chapter 3 in Booklet 1, *Getting Started with QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Experiments*	4489822

3 Prepare the Reactions

This chapter explains how to prepare the PCR reactions for the Relative Standard Curve example experiment.

This chapter covers:

Assemble required materials

- Items listed in Booklet 1, *Getting Started with QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Experiments*
- Samples - Total RNA isolated from kidney, heart, liver, and brain tissues.
- Example experiment reaction mix components:
  - TaqMan® Gene Expression Master Mix (2×)
  - FAS Assay Mix (20×)
  - HPRT Assay Mix (20×)

Note: Fast Universal Master Mix is not recommended to be used with the 96-well (0.2 mL) reaction plates or reaction tubes and tube strips sealed with caps.

Prepare the template

Prepare the template for the PCR reactions (both samples and standards) using one of the Invitrogen VILO kits, SuperScript® VILO™ cDNA Synthesis Kit (Part no. 4453650).

Example experiment settings

For the Relative Standard Curve example experiment, the template for the PCR reactions is cDNA reverse-transcribed from total RNA samples using the Invitrogen VILO kits.

To prepare the template

Use the Invitrogen VILO kits to reverse-transcribe cDNA from the total RNA samples. Follow the procedures in the *Invitrogen VILO Kits Protocol* (Pub. no. 100002284) to:

1. Isolate total RNA from cells using an Ambion® sample preparation kit appropriate to the tissue or cell type.
2. Quantify and perform quality control on the RNA.
3. Convert the RNA to cDNA via reverse transcription.

Prepare the sample dilutions

For the Relative Standard Curve example experiment, no more than 10% of your reaction should consist of the undiluted cDNA.

1. Label a separate microcentrifuge tube for each diluted sample:
   - Kidney
   - Heart
   - Liver
   - Brain
2. Add the required volume of water (diluent) to each empty tube:

Tube	Sample name	Diluent volume (µL)
1	Kidney	76
2	Heart	76
3	Liver	76
4	Brain	76

3. Add the required volume of sample stock (100 ng/µL) to each empty tube:

Tube	Sample name	Diluent volume (µL)
1	Kidney	4
2	Heart	4
3	Liver	4
4	Brain	4

4. Vortex each diluted sample for 3 to 5 seconds, then centrifuge the tubes briefly.
5. Place the diluted samples on ice until you prepare the reaction plate.

Prepare the standard dilution series for FAS and HPRT assays

The same standard materials are used to prepare the exact same dilutions for both the target genes. The prepared standards are then used to generate the two standard curves.

- The stock concentration for cDNA is 100 ng/µL.
- The volumes calculated for both the FAS and HPRT assays are:

Standard name (labeled tube)	Dilution point	Source	Source volume (µL)	Diluent volume (µL)	Total volume (µL)	Standard concentration (ng/µL)
Std. 1	1	Stock	20	20	40	100
Std. 2	2	Dilution 1	20	20	40	50
Std. 3	3	Dilution	20	20	40	25
Std. 4	4	Dilution 3	20	20	40	12.5
Std. 5	5	Dilution 4	20	20	40	6.25

1. Label ten separate microcentrifuge tubes for each diluted standard:
   - FAS (FAS Std. 1 - FAS Std. 5)
   - HPRT (HPRT Std. 1 - HPRT Std. 5)
2. Prepare five standard dilutions each for FAS and HPRT:

   Note: For dilution 1, first vortex the stock for 3 to 5 seconds, then centrifuge both the Std. 1 tubes briefly before pipetting 10 µL stock into each Std. 1 tube.
3. For each subsequent dilution, add source to the standard:
   a. Use a new pipette tip to add 10 µL of source to the FAS and HPRT tubes containing the standard.
   b. Vortex the tubes for 3 to 5 seconds, then centrifuge the tubes briefly.
4. Place the standards on ice until you prepare the reaction plate.

Prepare the reaction mix ("cocktail mix")

1. Label an appropriately sized tube for each reaction mix:
   - FAS Reaction Mix
   - HPRT Reaction Mix
2. For the FAS assay, add the required volumes of each component to the FAS Reaction Mix tube:

Component	Volume (µL) for 1 reaction	Volume (µL) for 96 reactions (plus 10% excess)
TaqMan® Gene Expression Master Mix (2X)	5	530
FAS Assay Mix (20X)	0.5	53
Water	3.5	371
Total Reaction Mix Volume	9	954

3. For the HPRT assay, add the required volumes of each component to the HPRT Reaction Mix tube:

Component	Volume (µL) for 1 reaction	Volume (µL) for 96 reactions (plus 10% excess)
TaqMan® Gene Expression Master Mix (2X.)	5	530
HPRT Assay Mix (20X	0.5	53
Water	3.5	371
Total Reaction Mix Volume	9	954

4. Mix the reaction in each tube by gently pipetting up and down, then cap each tube.
5. Centrifuge the tubes briefly to remove air bubbles.
6. Place the reaction mixes on ice until you prepare the reaction plate.

   Note: Do not add the sample or standard at this time.

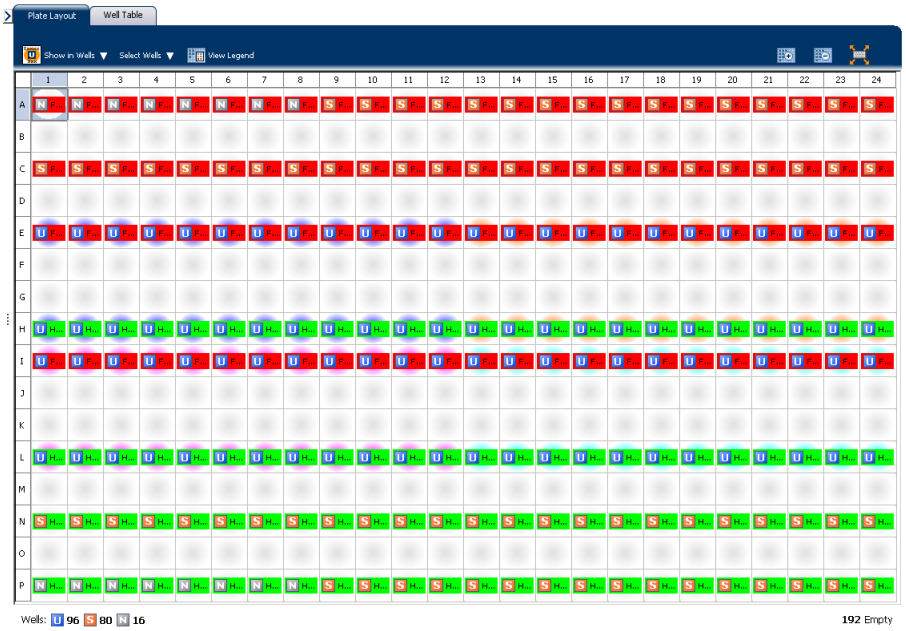
Prepare the reaction plate

Example experiment reaction plate components

The reaction plate for the Relative Standard Curve example experiment contains:

- A MicroAmp® Optical 384-Well Reaction Plate
- Reaction volume: 10 µL/well
- 96 Unknown wells U
- 80 Standard wells S
- 16 Negative control wells N
- 192 Empty wells

The following is an image of the plate layout for the example experiment:



To prepare the reaction plate components

1. For each target, prepare the negative control reactions:
   a. To an appropriate volume tube, add the volumes of reaction mix and water listed below.

Tube	Reaction mix	Reaction mix volume (µL)	Water volume (µL)
1	FAS Reaction Mix	79.2	8.8
2	HPRT Reaction Mix	79.2	8.8

   b. Mix the reaction by gently pipetting up and down, then cap the tube.
   c. Centrifuge the tube briefly to remove air bubbles.
   d. Add 10 µL of the negative control reaction to the appropriate wells in the reaction plate.

2. For each replicate group, prepare the standard reactions:
   a. To appropriately sized tubes, add the volumes of reaction mix and standard listed below.

Tube	Standard reaction	Reaction mix	Reaction mix volume (µL)	Standard	Standard volume (µL)
1	FAS Std. 1	FAS Reaction Mix	79.2	FAS Std. 1	8.8
2	FAS Std. 2	FAS Reaction Mix	79.2	FAS Std. 2	8.8
3	FAS Std. 3	FAS Reaction Mix	79.2	FAS Std. 3	8.8
4	FAS Std. 4	FAS Reaction Mix	79.2	FAS Std. 4	8.8
5	FAS Std. 5	FAS Reaction Mix	79.2	FAS Std. 5	8.8
6	HPRT Std. 1	HPRT Reaction Mix	79.2	HPRT Std. 1	8.8
7	HPRT Std. 2	HPRT Reaction Mix	79.2	HPRT Std. 2	8.8
8	HPRT Std. 3	HPRT Reaction Mix	79.2	HPRT Std. 3	8.8
9	HPRT Std. 4	HPRT Reaction Mix	79.2	HPRT Std. 4	8.8
10	HPRT Std. 5	HPRT Reaction Mix	79.2	HPRT Std. 5	8.8

   b. Mix the reactions by gently pipetting up and down, then cap the tubes.
   c. Centrifuge the tubes briefly to remove air bubbles.
   d. Add 10 µL of the standard reaction to the appropriate wells in the reaction plate.

3. For each replicate group, prepare the reactions for the unknowns:
   a. To appropriately sized tubes, add the volumes of reaction mix and sample listed below:

Tube	Unknown reaction	Reaction mix	Reaction mix volume (µL)	Sample	Sample volume (µL)
1	FAS Kidney	FAS Reaction Mix	118.8	Kidney	13.2
2	FAS Heart	FAS Reaction Mix	118.8	Heart	13.2
3	FAS Liver	FAS Reaction Mix	118.8	Liver	13.2
4	FAS Brain	FAS Reaction Mix	118.8	Brain	13.2
5	HPRT Kidney	HPRT Reaction Mix	118.8	Kidney	13.2
6	HPRT Heart	HPRT Reaction Mix	118.8	Heart	13.2
7	HPRT Liver	HPRT Reaction Mix	118.8	Liver	13.2
8	HPRT Brain	HPRT Reaction Mix	118.8	Brain	13.2

   b. Mix the reactions by gently pipetting up and down, then cap the tubes.
   c. Centrifuge the tubes briefly to remove air bubbles.
   d. Add 10 µL of the unknown (sample) reaction to the appropriate wells in the reaction plate.
4. Seal the reaction plate with optical adhesive film.
5. Centrifuge the reaction plate briefly to remove air bubbles.
6. Confirm that the liquid is at the bottom of each well of the reaction plate. If not, centrifuge the reaction plate again at a higher speed and for a longer period of time.
7. Until you are ready to perform the run, place the reaction plate at 4°C, in the dark.

Tips for preparing reactions for your own experiments

Tips for preparing templates

When you prepare your own Relative Standard Curve experiment, Life Technologies recommends the following templates:

- **Complementary DNA (cDNA)** – cDNA reverse-transcribed from total RNA samples.
- **Genomic DNA (gDNA)** – Purified gDNA already extracted from tissue or sample

Tips for preparing sample dilutions

When you prepare your own Relative Standard Curve experiment, for optimal performance of TaqMan® Gene Expression Assays or Custom TaqMan® Gene Expression Assays, use 10 to 100 ng of cDNA template per 10µL reaction.

Tips for preparing the reaction mix

If your experiment includes more than one target assay, prepare the reaction mix for each target assay separately.

Tips for preparing the reaction plate

When you prepare your own Relative Standard Curve experiment, make sure the arrangement of the PCR reactions matches the plate layout displayed in the QuantStudio™ 6 and 7 Flex Software.

For more information

For more information on...	Refer to...	Publication number
Assigning the reaction plate components	Chapter 2 in Booklet 1, *Getting Started with QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Experiments*	4489822
Sealing the reaction plate	Chapter 2 in Booklet 1, *Getting Started with QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Experiments*	4489822

4 Run the Experiment

This chapter explains how to run the example experiment on the QuantStudio™ 6 or 7 Instrument.

This chapter covers:

IMPORTANT! Run the experiment at the same ambient temperature at which you calibrated the instrument. Extreme variations in ambient temperature can affect the heating and cooling of the instrument and influence experimental results.

IMPORTANT! Do not attempt to open the access door during the run. The door is locked while the QuantStudio™ 6 or 7 Instrument is in operation.

Start the run

1. Open the Relative Standard Curve example file that you created using instructions in Chapter 2.
2. Load the reaction plate into the instrument.
3. Start the run.

Monitor the run

Monitor the example experiment run:

- From the QuantStudio™ 6 and 7 Flex Real-Time PCR System Software using the Run screen, while the experiment is in progress.
- From the Instrument Console of the QuantStudio™ 6 and 7 Flex Real-Time PCR System Software (to monitor an experiment started from another computer or from the QuantStudio™ 6 or 7 Instrument touchscreen).
- From the QuantStudio™ 6 or 7 Instrument touchscreen.

From the Instrument Console of the QuantStudio™ 6 and 7 Flex Real-Time PCR System Software

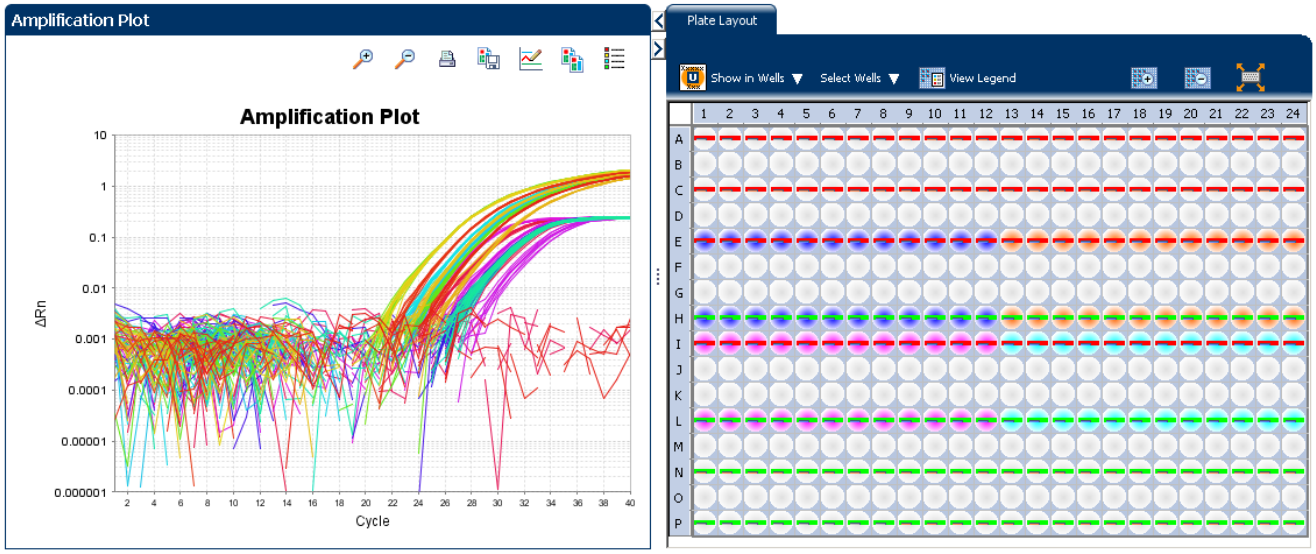
1. In the Instrument Console screen, select the instrument icon.
2. Click **Manage Instrument** or double-click on the instrument icon.
3. In the Manage Instrument screen, click **Monitor Running Experiment** to access the Run screen.

View the Amplification Plot

You can view the progress of the run in real time. During the run, periodically view all the three plots available from the QuantStudio™ 6 and 7 Flex Real-Time PCR System Software for potential problems.

To view data in the Amplification Plot screen, click **Amplification Plot** from the Run Experiment Menu, and select the Plate Layout tab, then select the wells that you want to view.

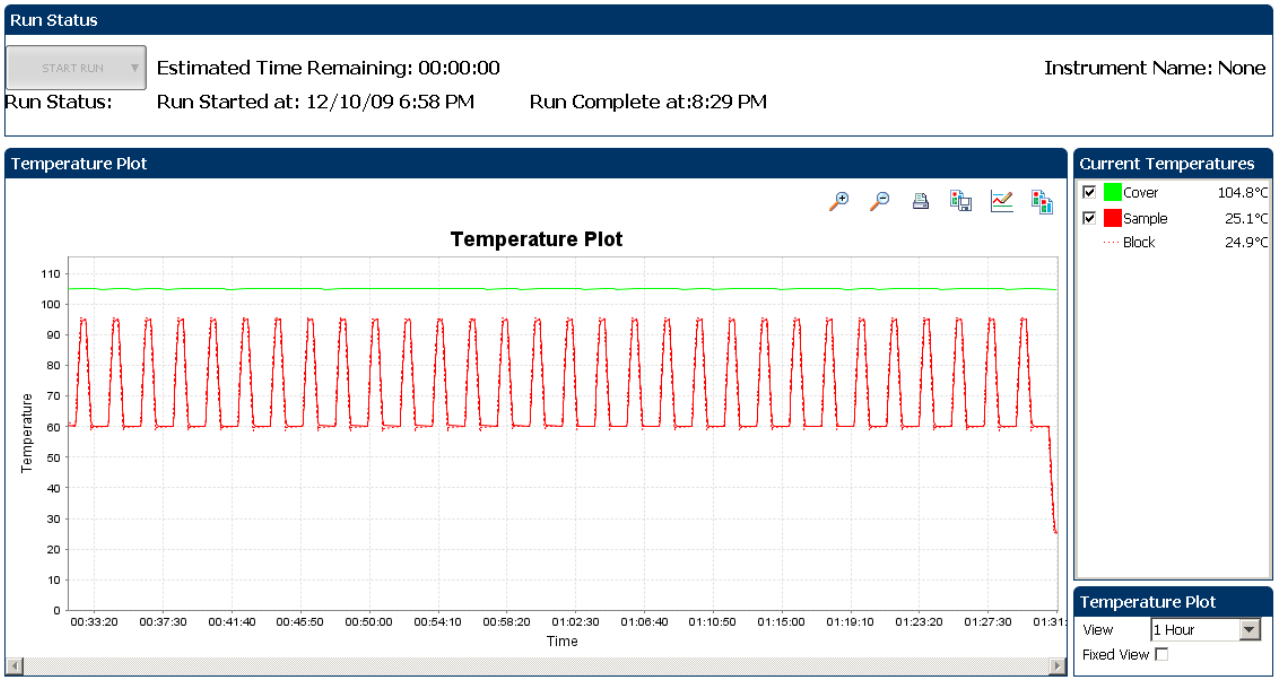
The following is an image of the Amplification Plot screen as it appears at the end of the example experiment.



View the Temperature Plot

To view data in the Temperature Plot screen, click **Temperature Plot** from the Run Experiment Menu.

The following is an image of the Temperature Plot screen as it appears during the example experiment.

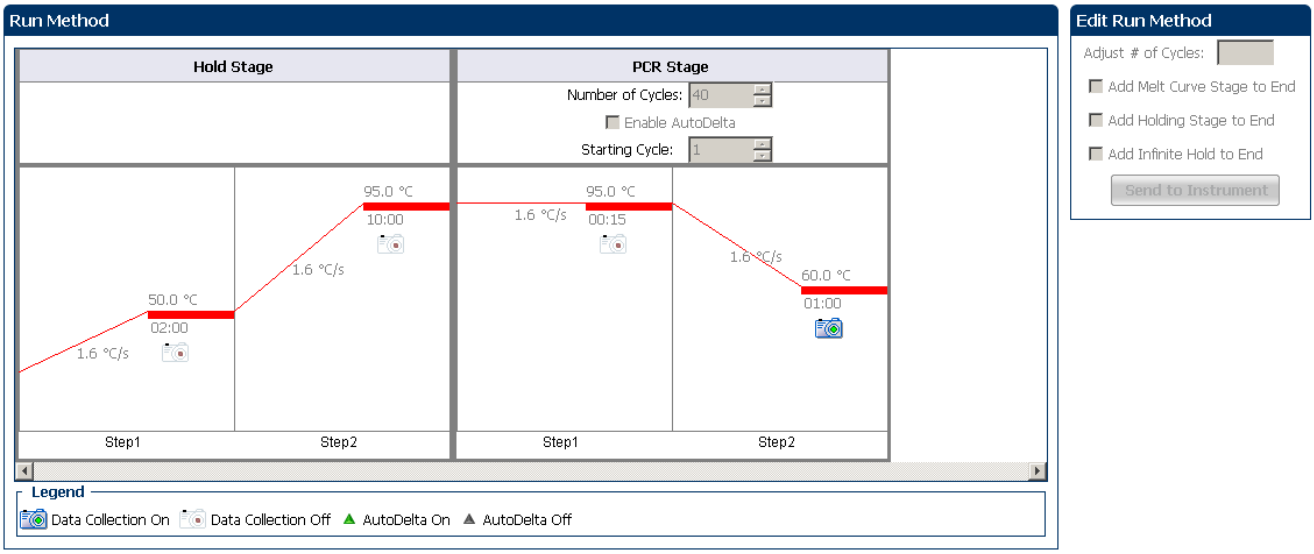


Note: The Sample temperature displayed in the Current Temperatures group is an estimated value.

View the Run Method

To view data in the Run Method screen, click **Run Method** from the Run Experiment Menu.

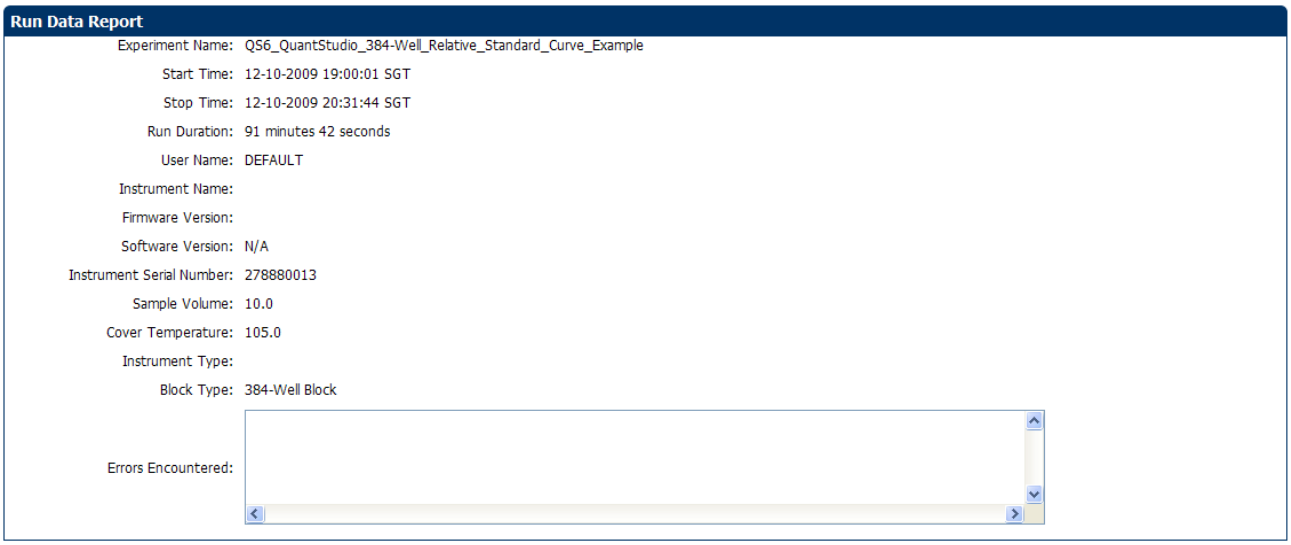
The following is an image of the Run Method screen as it appears in the example experiment.



View run data

To view the run data, click **View Run Data** from the Run Experiment Menu.

The following is an image of the View Run Data screen as it appears in the example experiment:

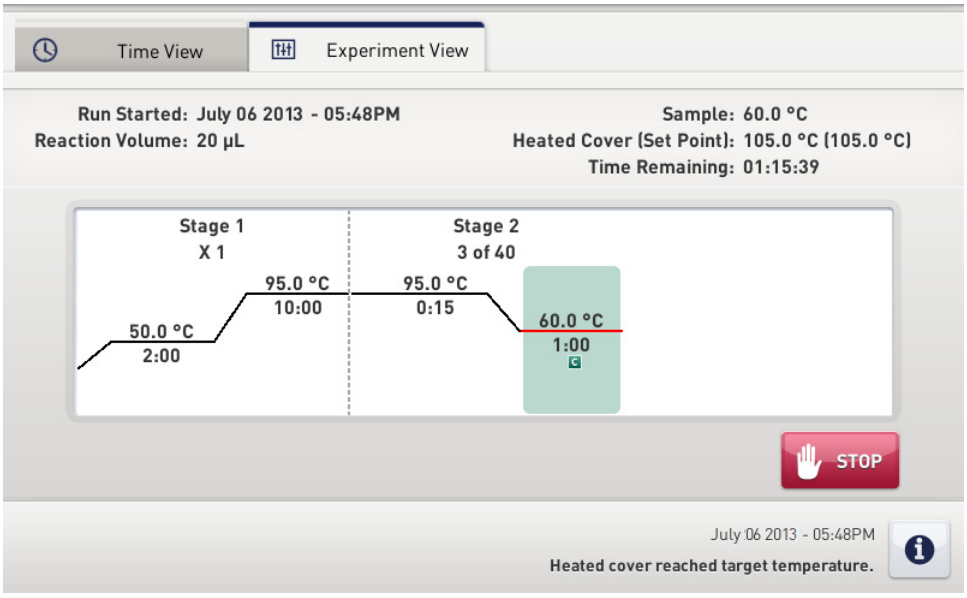


From the QuantStudio™ 6 or 7 Instrument touchscreen

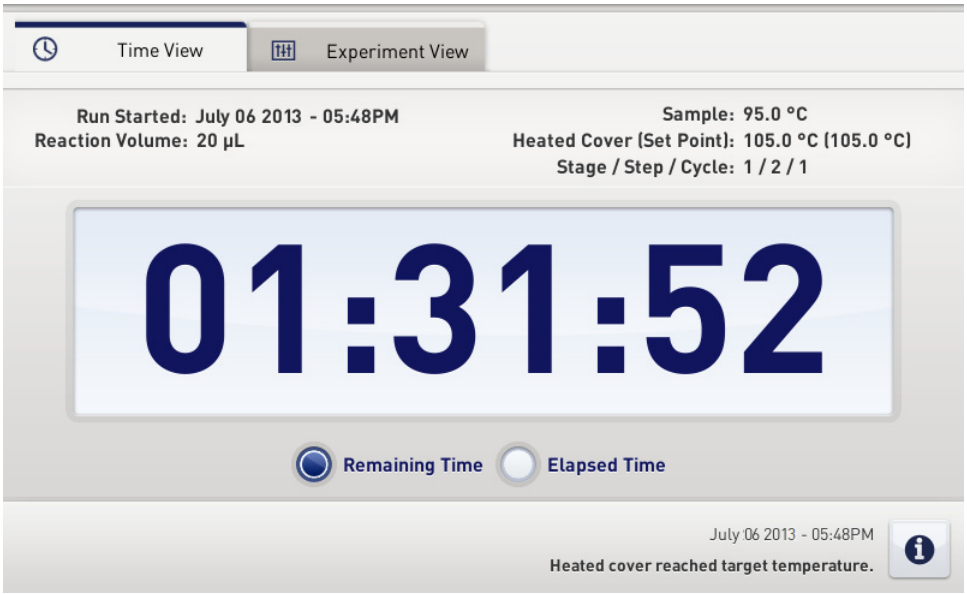
You can also view the progress of the run from the QuantStudio™ 6 or 7 Instrument touchscreen.

The following are images of the Run Method screen on the QuantStudio™ 6 or 7 Instrument touchscreen:

Experiment view



Time view



5 Review Results and Adjust Experiment Parameters

In Section 5.1 of this chapter you review the analyzed data using several of the analysis screens and publish the data. Section 5.2 of this chapter walks you through the process of modifying experiment parameters to troubleshoot problems with experiment results before rerunning an experiment.

This chapter covers:

Section 5.1 Review Results

Analyze the example experiment

1. Open the example experiment file that you ran in Chapter 4.
2. Click **Analyze**. The software analyzes the data using the default analysis settings. You can also access the experiment to analyze from the Home screen.

View the Standard Curve Plot

The Standard Curve Plot screen displays the standard curve for samples designated as standards. The QuantStudio™ 6 and 7 Flex Real-Time PCR System Software calculates the quantity of an unknown target from the standard curve.

Example experiment standard curve values

In the standard curve example experiment, you review the Standard Curve Plot screen for the following regression coefficient values:

- Slope/amplification efficiency
- R^2 value (correlation coefficient)
- C_T values

To view and assess the Standard Curve plot

1. From the Experiment Menu pane, select **Analysis ▸ Standard Curve**.

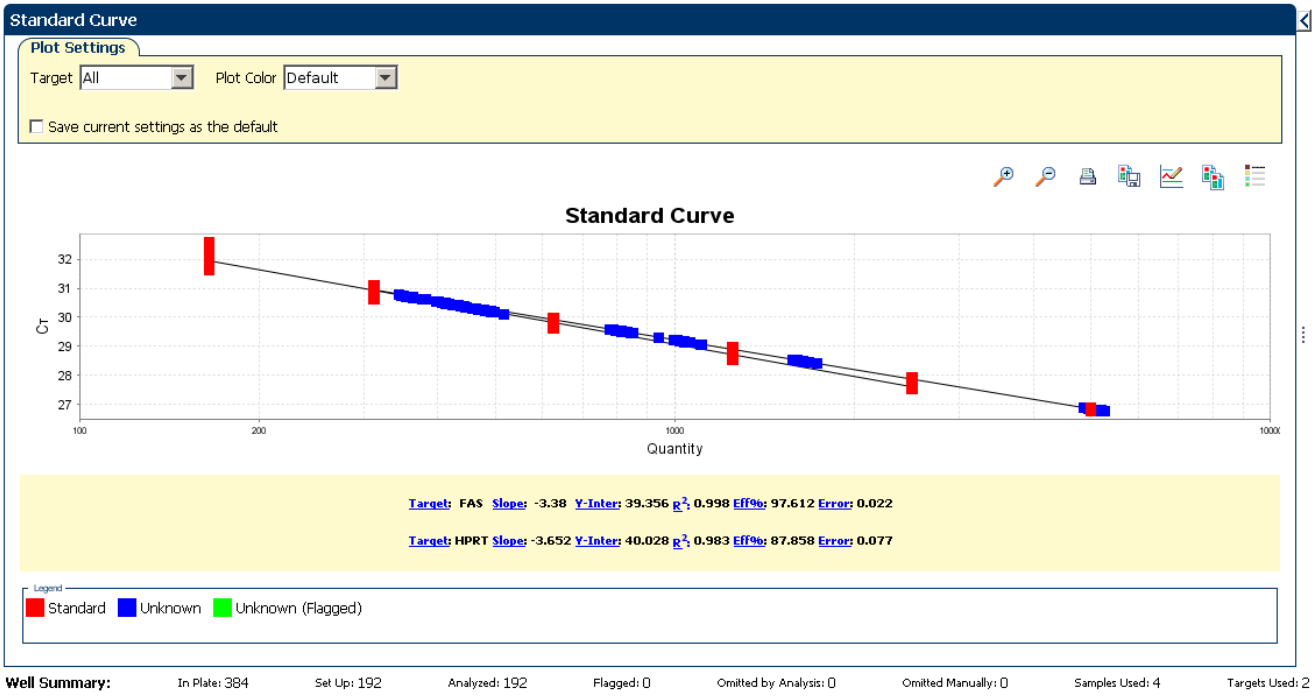
   Note: If no data are displayed, click **Analyze**.
2. Display all 384 wells in the Standard Curve Plot screen by clicking the upper left corner of the plate layout in the Plate Layout tab.
3. Enter the Plot Settings:

Menu	Selection
Target	All
Plot Color	Default
 (This is a toggle button. When the legend is displayed, the button changes to Hide the plot legend.)	Check (default)

4. View the values displayed below the standard curve. In the example experiment, the values for each target fall within the acceptable ranges:

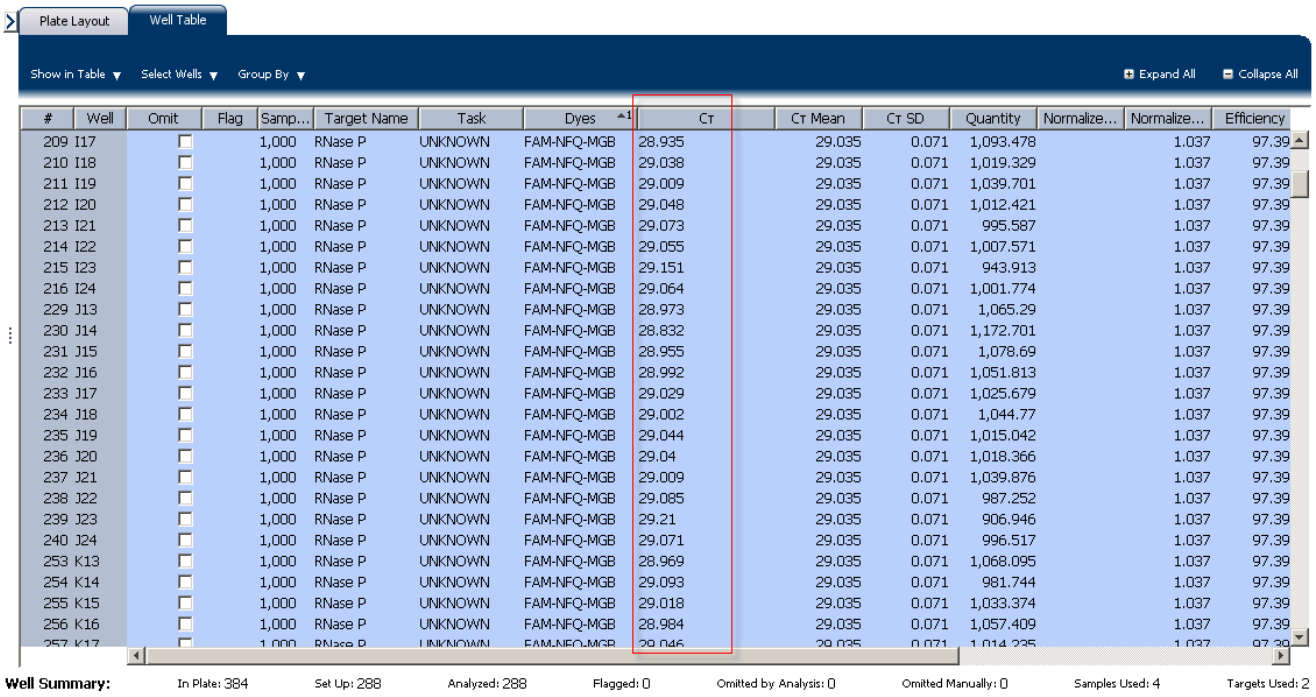
Target	Slope	R^2 Value	Amplification efficiency (Eff%)
FAS	–3.38	0.998	97.612
HPRT	–3.652	0.983	87.858

5. Check that all samples are within the standard curve. In the example experiment, all samples (blue dots) are within the standard curve (red dots).



6. Check the C_T values:
7. Click the **Well Table** tab.
8. From the Group By drop-down menu, select **Replicate**.

9. Look at the values in the C_T column. In the example experiment, the C_T values fall within the expected range (>8 and <35).



#	Well	Omit	Flag	Samp...	Target Name	Task	Dyes	CT	CT Mean	CT SD	Quantity	Normalize...	Normalize...	Efficiency
209	I17			1,000	RNase P	UNKNOWN	FAM-NFQ-MGB	28.935	29.035	0.071	1,093.478		1.037	97.39
210	I18			1,000	RNase P	UNKNOWN	FAM-NFQ-MGB	29.038	29.035	0.071	1,019.329		1.037	97.39
211	I19			1,000	RNase P	UNKNOWN	FAM-NFQ-MGB	29.009	29.035	0.071	1,039.701		1.037	97.39
212	I20			1,000	RNase P	UNKNOWN	FAM-NFQ-MGB	29.048	29.035	0.071	1,012.421		1.037	97.39
213	I21			1,000	RNase P	UNKNOWN	FAM-NFQ-MGB	29.073	29.035	0.071	995.587		1.037	97.39
214	I22			1,000	RNase P	UNKNOWN	FAM-NFQ-MGB	29.055	29.035	0.071	1,007.571		1.037	97.39
215	I23			1,000	RNase P	UNKNOWN	FAM-NFQ-MGB	29.151	29.035	0.071	943.913		1.037	97.39
216	I24			1,000	RNase P	UNKNOWN	FAM-NFQ-MGB	29.064	29.035	0.071	1,001.774		1.037	97.39
229	J13			1,000	RNase P	UNKNOWN	FAM-NFQ-MGB	28.973	29.035	0.071	1,065.29		1.037	97.39
230	J14			1,000	RNase P	UNKNOWN	FAM-NFQ-MGB	28.832	29.035	0.071	1,172.701		1.037	97.39
231	J15			1,000	RNase P	UNKNOWN	FAM-NFQ-MGB	28.955	29.035	0.071	1,078.69		1.037	97.39
232	J16			1,000	RNase P	UNKNOWN	FAM-NFQ-MGB	28.992	29.035	0.071	1,051.813		1.037	97.39
233	J17			1,000	RNase P	UNKNOWN	FAM-NFQ-MGB	29.029	29.035	0.071	1,025.679		1.037	97.39
234	J18			1,000	RNase P	UNKNOWN	FAM-NFQ-MGB	29.002	29.035	0.071	1,044.77		1.037	97.39
235	J19			1,000	RNase P	UNKNOWN	FAM-NFQ-MGB	29.044	29.035	0.071	1,015.042		1.037	97.39
236	J20			1,000	RNase P	UNKNOWN	FAM-NFQ-MGB	29.04	29.035	0.071	1,018.366		1.037	97.39
237	J21			1,000	RNase P	UNKNOWN	FAM-NFQ-MGB	29.009	29.035	0.071	1,039.876		1.037	97.39
238	J22			1,000	RNase P	UNKNOWN	FAM-NFQ-MGB	29.085	29.035	0.071	987.252		1.037	97.39
239	J23			1,000	RNase P	UNKNOWN	FAM-NFQ-MGB	29.21	29.035	0.071	906.946		1.037	97.39
240	J24			1,000	RNase P	UNKNOWN	FAM-NFQ-MGB	29.071	29.035	0.071	996.517		1.037	97.39
253	K13			1,000	RNase P	UNKNOWN	FAM-NFQ-MGB	28.969	29.035	0.071	1,068.095		1.037	97.39
254	K14			1,000	RNase P	UNKNOWN	FAM-NFQ-MGB	29.093	29.035	0.071	981.744		1.037	97.39
255	K15			1,000	RNase P	UNKNOWN	FAM-NFQ-MGB	29.018	29.035	0.071	1,033.374		1.037	97.39
256	K16			1,000	RNase P	UNKNOWN	FAM-NFQ-MGB	28.984	29.035	0.071	1,057.409		1.037	97.39
257	K17			1,000	RNase P	UNKNOWN	FAM-NFQ-MGB	29.046	29.035	0.071	1,014.235		1.037	97.39

Tips for analyzing your own experiments

When you analyze your own Relative Standard Curve experiment, look for:

- **Slope/amplification efficiency values** – The amplification efficiency is calculated using the slope of the regression line in the standard curve. A slope close to –3.3 indicates optimal, 100% PCR amplification efficiency. Factors that affect amplification efficiency:
  - Range of standard quantities – For more accurate and precise efficiency measurements, use a broad range of standard quantities, 5 to 6 logs (10^5 to 10^6 fold).
  - Number of standard replicates – For more accurate efficiency measurements, include replicates to decrease the effects of pipetting inaccuracies.
  - PCR inhibitors – PCR inhibitors in the reaction can alter amplification efficiency.
- **R^2 values (correlation coefficient)** – The R^2 value is a measure of the closeness of fit between the regression line and the individual C_T data points of the standard reactions. A value of 1.00 indicates a perfect fit between the regression line and the data points. An R^2 value >0.99 is desirable.
- **C_T values** – The threshold cycle (C_T) is the PCR cycle number at which the fluorescence level meets the threshold.
  - A C_T value >8 and <35 is desirable.
  - A C_T value <8 indicates that there is too much template in the reaction.
  - A C_T value >35 indicates a low amount of target in the reaction; for C_T values >35, expect a higher standard deviation.

If your experiment does not meet the guidelines above, troubleshoot as follows:

- Omit wells (see "Improve CT precision by omitting wells" on page 65).
  Or
- Rerun the experiment.

Assess amplification results using the Amplification Plot

Amplification plots available for viewing

The Amplification Plot screen displays amplification of all samples in the selected wells. There are three plots available:

- **ΔRn vs Cycle** – ΔRn is the magnitude of normalized fluorescence signal generated by the reporter at each cycle during the PCR amplification. This plot displays ΔRn as a function of cycle number. Use this plot to identify and examine irregular amplification and to view threshold and baseline values for the run.
- **Rn vs Cycle** – Rn is the fluorescence signal from the reporter dye normalized to the fluorescence signal from the passive reference. This plot displays Rn as a function of cycle number. Use this plot to identify and examine irregular amplification.
- **C_T vs Well** – C_T is the PCR cycle number at which the fluorescence meets the threshold in the amplification plot. This plot displays C_T as a function of well position. Use this plot to locate outlying amplification (outliers).

Each plot can be viewed as a linear or log10 graph type.

Purpose

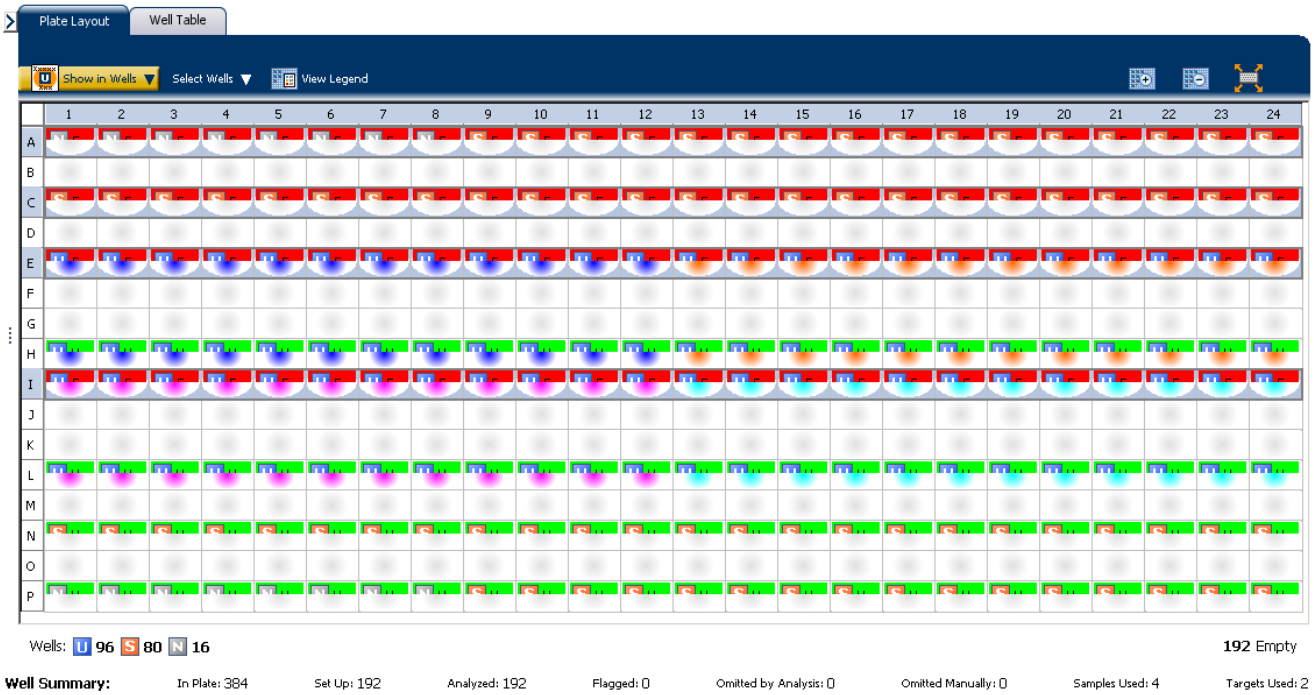
The purpose of viewing the amplification plot for the example experiment is to identify:

- Correct baseline and threshold values
- Outliers

View the Amplification Plot

1. From the Experiment Menu pane, select **Analysis ▸ Amplification Plot**.
   Note: If no data are displayed, click **Analyze**.
2. Display the FAS wells in the Amplification Plot screen:
   a. Click the **Plate Layout** tab.
   b. From the Select Wells drop-down menu, select **Target**, then **FAS**.

The following is an image of the Plate Layout screen:

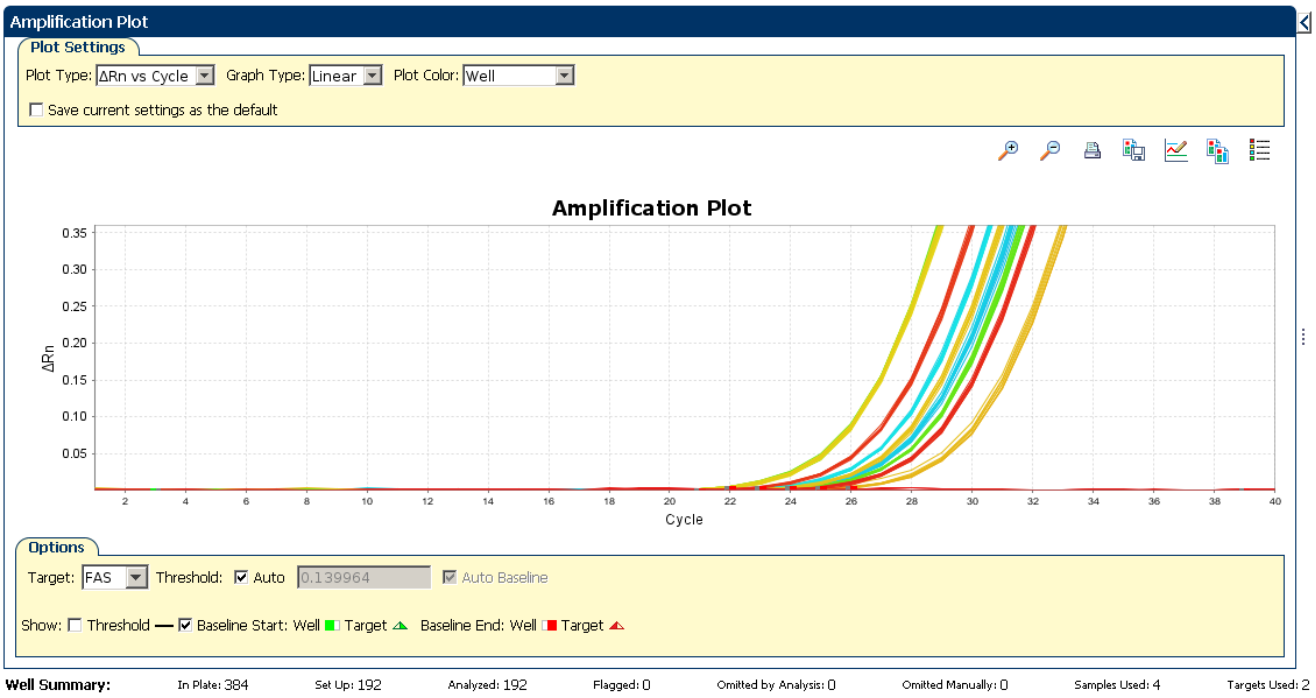


3. In the Amplification Plot screen, enter:

Menu	Selection
Plot Type	ΔRn vs Cycle (default)
Plot Color	Well (default)
 (This is a toggle button. When the legend is displayed, the button changes to **Hide the plot legend**.)	Check (default)

4. View the baseline values:
   a. From the Graph Type drop-down menu, select **Linear**.
   b. Select the **Baseline** check box to show the start cycle and end cycle.

c. Verify that the baseline is set correctly: The end cycle should be set a few cycles before the cycle number where significant fluorescent signal is detected. In the example experiment, the baseline is set correctly.

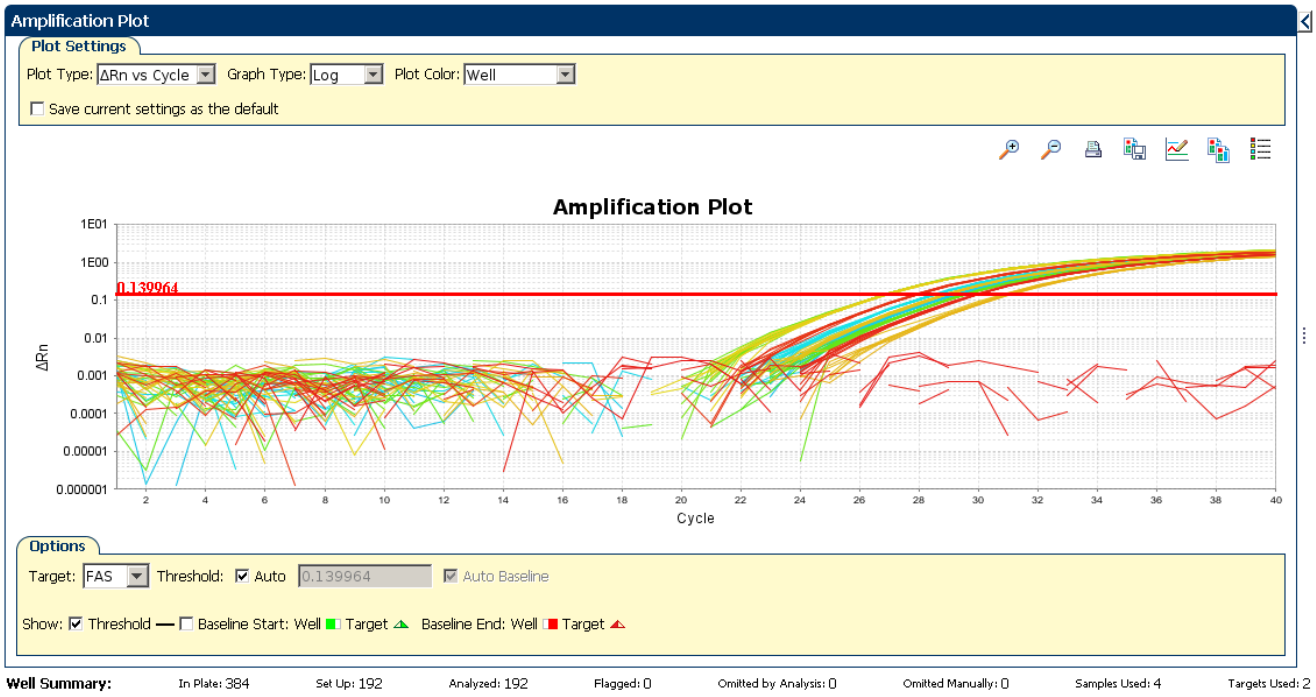


5. View the threshold values:

Menu	Select
Graph Type	Log
Target	FAS

a. Select the **Threshold** check box to show the threshold.

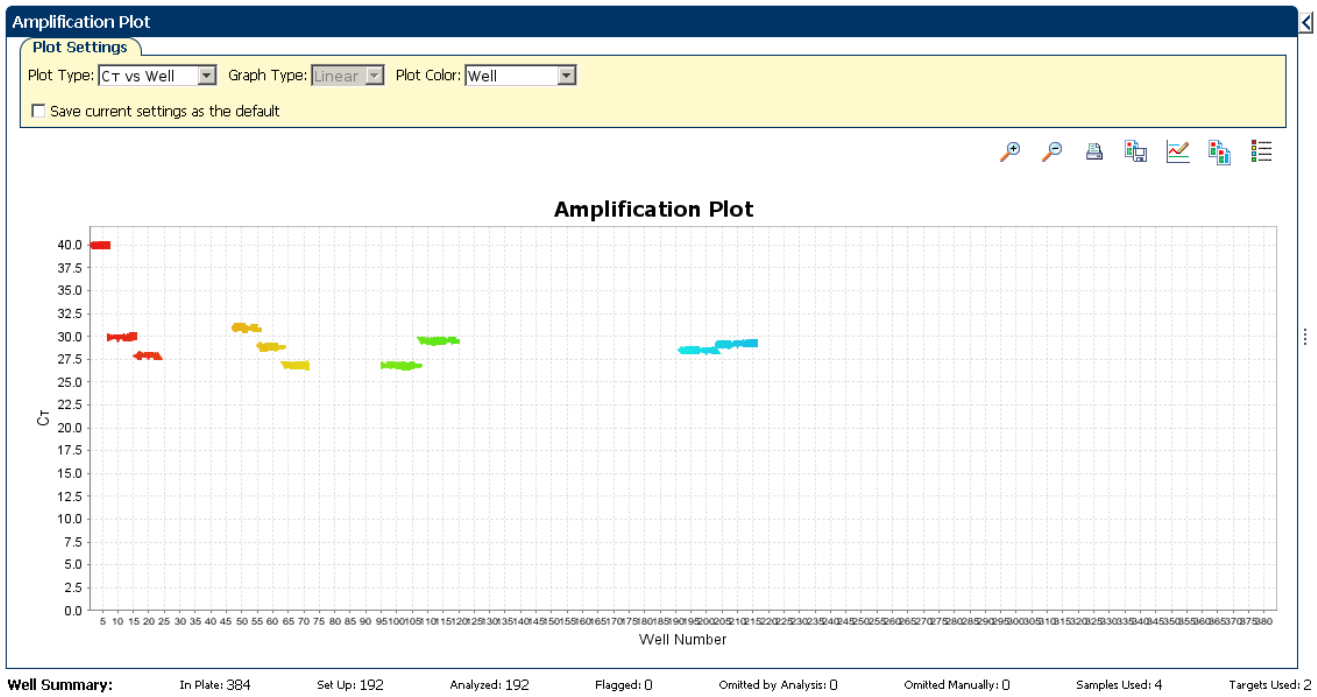
b. Verify that the threshold is set correctly. In the example experiment, the threshold is in the exponential phase.



6. Locate outliers:

   a. From the Plot Type drop-down menu, select **C_T vs Well**.

   b. Look for outliers from the amplification plot. In the example experiment, there are no outliers for FAS.



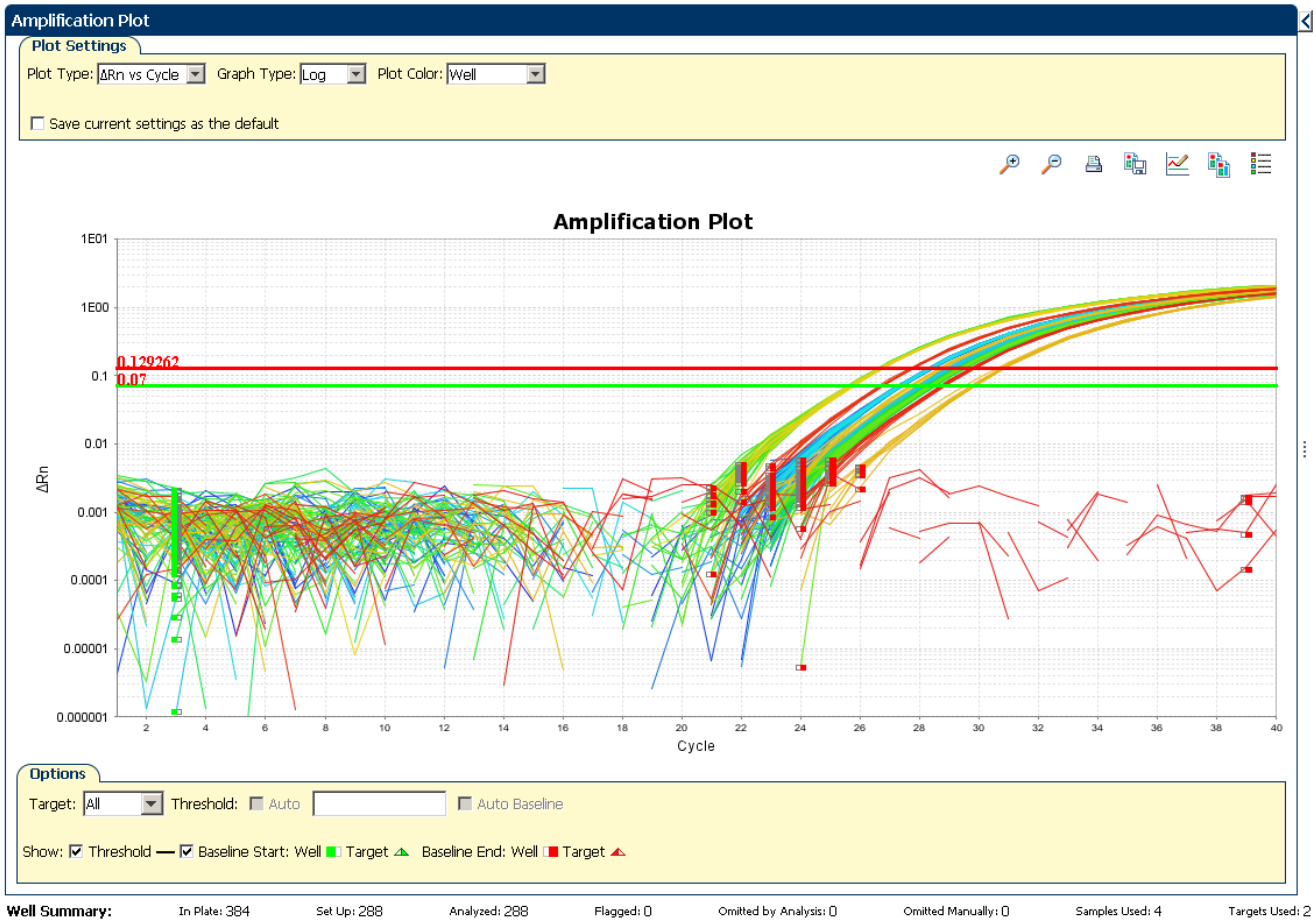
7. Repeat steps 2 through 6 for the HPRT wells.

Tips for analyzing your own experiments

When you analyze your own Relative Standard Curve experiment, look for:

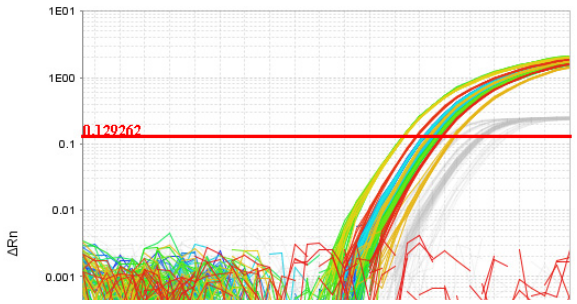
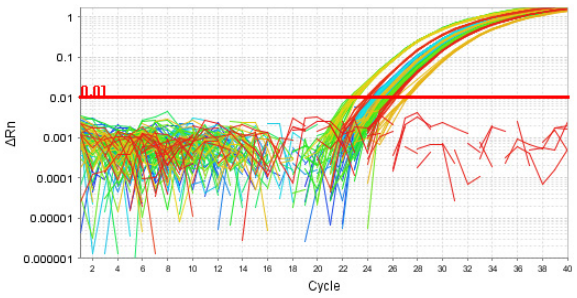
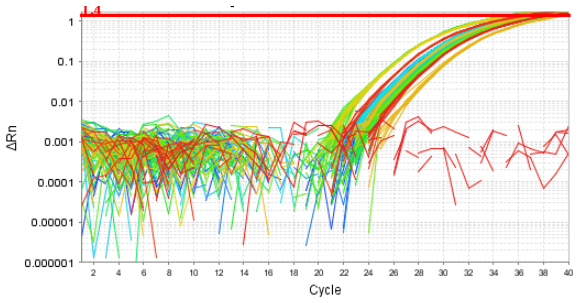
- **Outliers**
- **A typical amplification plot** – The QuantStudio™ 6 and 7 Flex Real-Time PCR System Software automatically calculates baseline and threshold values based on the assumption that the data exhibit a *typical* amplification plot. A typical amplification plot has four distinct sections:
  - Plateau phase
  - Linear phase
  - Exponential (geometric phase)
  - Baseline

  A following is an image of a typical amplification plot:

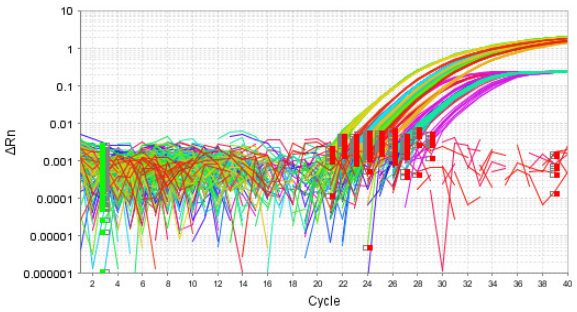


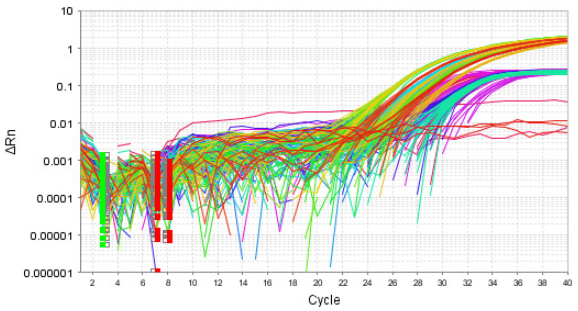
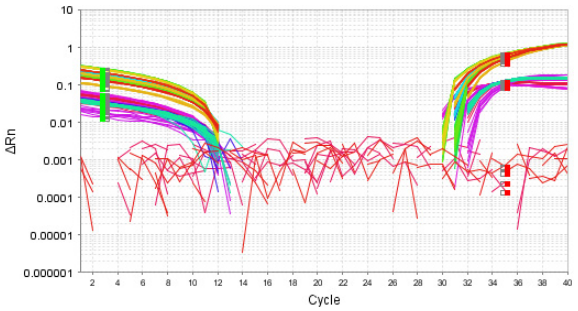
IMPORTANT! Experimental error (such as contamination or pipetting errors) can produce atypical amplification curves that can result in incorrect baseline and threshold value calculations by the QuantStudio™ 6 and 7 Flex Real-Time PCR System Software. Therefore, Life Technologies recommends that you examine the Amplification Plot screen and review the assigned baseline and threshold values for each well after analysis completes.

- **Correct threshold values.**

Threshold Set Correctly The threshold is set in the exponential phase of the amplification curve. Threshold settings above or below the optimum increase the standard deviation of the replicate groups.	
Threshold Set Too Low The threshold is set below the exponential phase of the amplification curve. The standard deviation is significantly higher than that for a plot where the threshold is set correctly. Drag the threshold bar up into the exponential phase of the curve.	
Threshold Set Too High The threshold is set above the exponential phase of the amplification curve. The standard deviation is significantly higher than that for a plot where the threshold is set correctly. Drag the threshold bar down into the exponential phase of the curve.	

- **Correct baseline values**

Baseline Set Correctly The amplification curve begins after the maximum baseline.	

Baseline Set Too Low The amplification curve begins too far to the right of the maximum baseline. Increase the End Cycle value.	
Baseline Set Too High The amplification curve begins before the maximum baseline. Decrease the End Cycle value.	

If your experiment does not meet the guidelines above, troubleshoot as follows:

- Omit wells (see "Improve CT precision by omitting wells" on page 65).

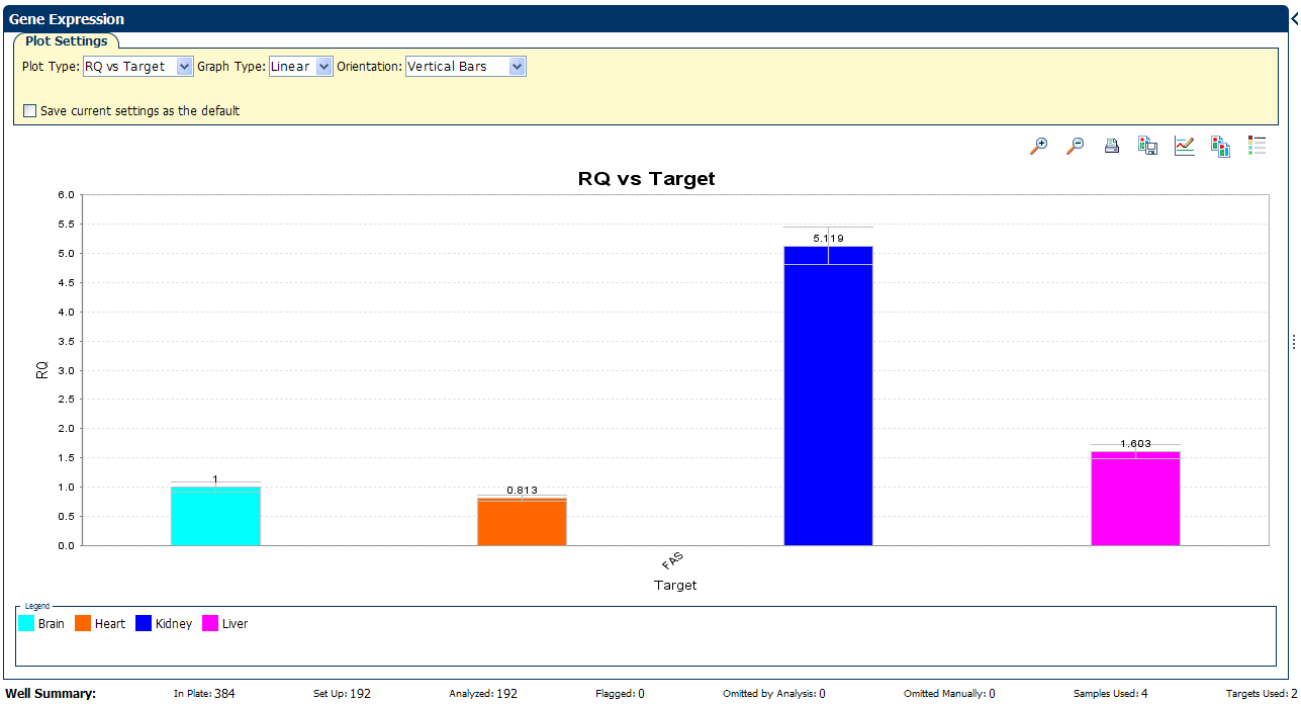
  Or

- Manually adjust the baseline and/or threshold (see "Adjust analysis settings" on page 61).

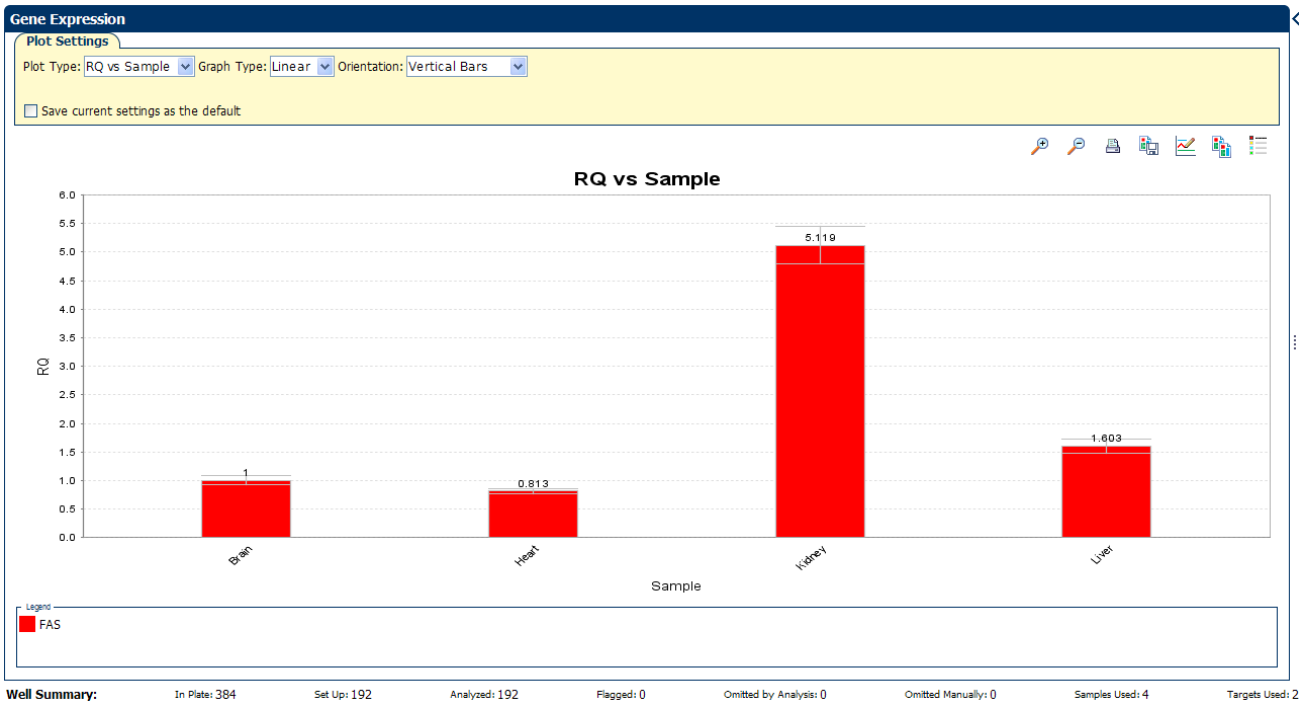
Assess the gene expression profile using the Gene Expression Plot

The Gene Expression Plot screen displays the results of relative quantification calculations in the gene expression profile. There are two plots available:

- **RQ vs Target** – Groups the relative quantification (RQ) values by target. Each sample is plotted for each target. You can view the plot as the linear, log10, Ln, and log2 graph types. The Gene Expression plot when viewed as a linear graph type looks like this:



- **RQ vs Sample** – Groups the relative quantification (RQ) values by sample. Each target is plotted for each sample. You can view the plot as the linear, log10, Ln, and log2 graph types. The Gene Expression plot when viewed as a linear graph type looks like this:



Example experiment values

Review each target in the Gene Expression Plot screen for the expression level (or fold change) of the target sample relative to the reference sample.

View the Gene Expression Plot

1. From the Experiment Menu pane, select **Analysis** ▸ **Gene Expression**.

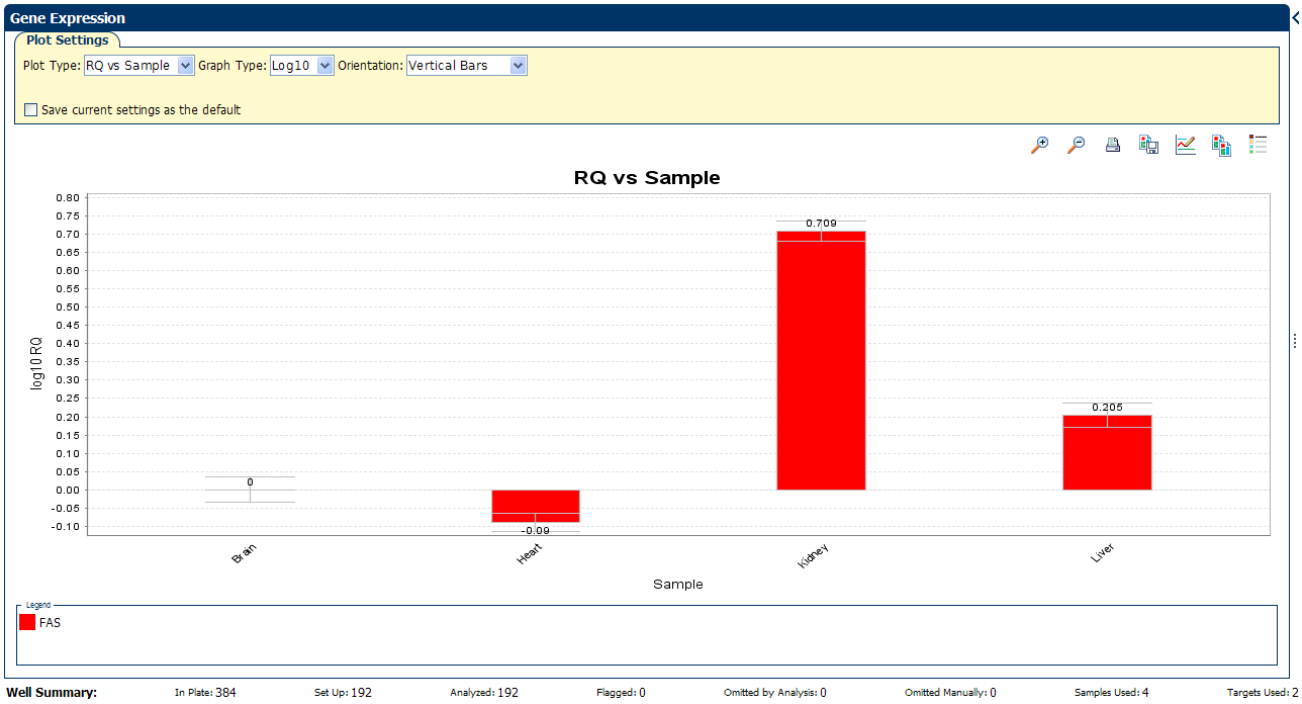
   Note: If no data are displayed, click **Analyze**.

2. In the Gene Expression Plot screen:
   a. From the Plot Type drop-down menu, select **RQ vs Sample**.
   b. From the Graph Type drop-down menu, select **Log10**.
   c. From the Orientation drop-down menu, select **Vertical Bars**.

3. Click **Show a legend for the plot** (default).

   Note: This is a toggle button. When the legend is displayed, the button changes to Hide the plot legend.

In the example experiment, the expression level of FAS in heart, kidney, and liver is displayed relative to its expression level in the reference sample (brain). Since the reference sample is compared to itself, the relative expression level is 1. When the result is displayed in the Log10 graph type, the expression level of the reference sample appears as 0 in the graph (log10 of 1 = 0).



Assessing the gene expression plot in your own experiments

When you analyze your own Relative Standard Curve experiment, look for differences in gene expression (as a fold change) relative to the reference sample.

Identify well problems using the Well Table

The Well Table displays data for each well in the reaction plate, including:

- The sample name, target name, task, and dyes
- The calculated threshold cycle (C_T), normalized fluorescence (Rn), and quantity values
- Flags

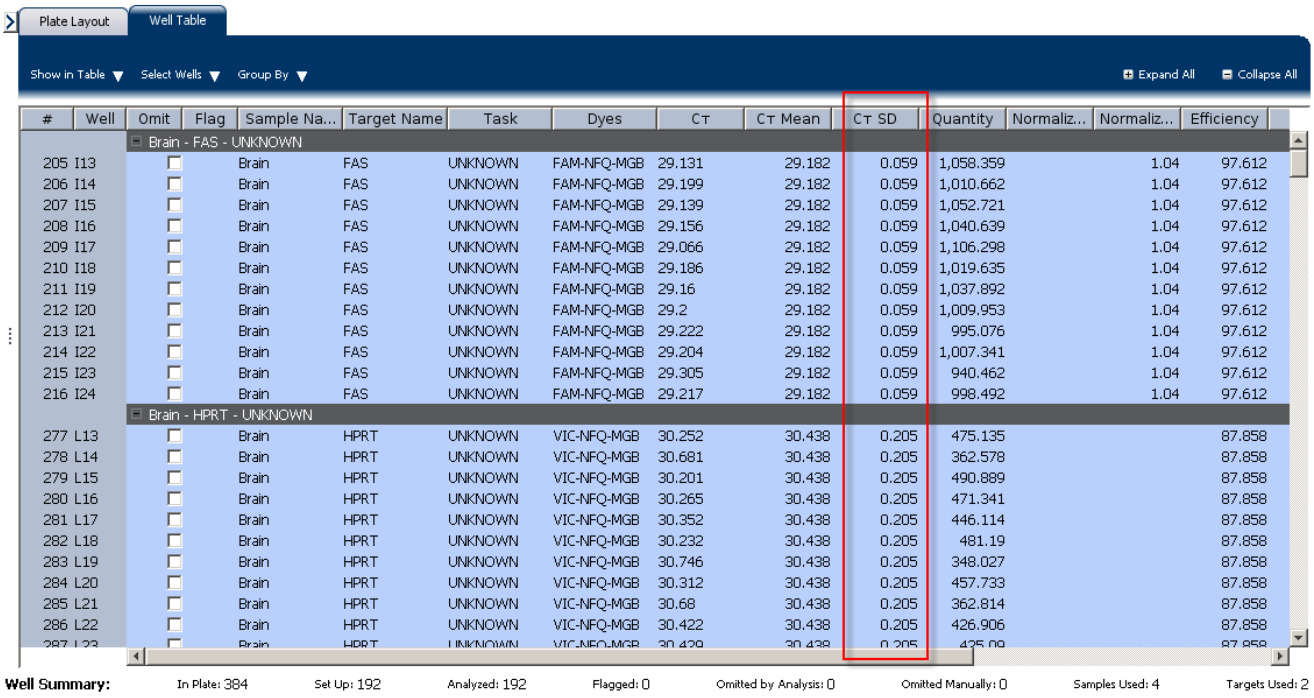
Example experiment values and flags

Review the Well Table to evaluate the C_T precision of the replicate groups.

View the well table

1. From the Experiment Menu pane, select **Analysis** ▸ **Amplification Plot**, then click the **Well Table** tab.

2. From the Group By drop-down menu, select **Replicate**.
3. Look at the C_T SD column to evaluate the C_T precision of the replicate groups. In the example experiment, the C_T SD have the expected value of < 0.5.



#	Well	Omit	Flag	Sample Na...	Target Name	Task	Dyes	CT	CT Mean	CT SD	Quantity	Normaliz...	Normaliz...	Efficiency
		Brain - FAS - UNKNOWN												
205	I13			Brain	FAS	UNKNOWN	FAM-NFQ-MGB	29.131	29.182	0.059	1,058.359		1.04	97.612
206	I14			Brain	FAS	UNKNOWN	FAM-NFQ-MGB	29.199	29.182	0.059	1,010.662		1.04	97.612
207	I15			Brain	FAS	UNKNOWN	FAM-NFQ-MGB	29.139	29.182	0.059	1,052.721		1.04	97.612
208	I16			Brain	FAS	UNKNOWN	FAM-NFQ-MGB	29.156	29.182	0.059	1,040.639		1.04	97.612
209	I17			Brain	FAS	UNKNOWN	FAM-NFQ-MGB	29.066	29.182	0.059	1,106.298		1.04	97.612
210	I18			Brain	FAS	UNKNOWN	FAM-NFQ-MGB	29.186	29.182	0.059	1,019.635		1.04	97.612
211	I19			Brain	FAS	UNKNOWN	FAM-NFQ-MGB	29.16	29.182	0.059	1,037.892		1.04	97.612
212	I20			Brain	FAS	UNKNOWN	FAM-NFQ-MGB	29.2	29.182	0.059	1,009.953		1.04	97.612
213	I21			Brain	FAS	UNKNOWN	FAM-NFQ-MGB	29.222	29.182	0.059	995.076		1.04	97.612
214	I22			Brain	FAS	UNKNOWN	FAM-NFQ-MGB	29.204	29.182	0.059	1,007.341		1.04	97.612
215	I23			Brain	FAS	UNKNOWN	FAM-NFQ-MGB	29.305	29.182	0.059	940.462		1.04	97.612
216	I24			Brain	FAS	UNKNOWN	FAM-NFQ-MGB	29.217	29.182	0.059	998.492		1.04	97.612
		Brain - HPRT - UNKNOWN												
277	L13			Brain	HPRT	UNKNOWN	VIC-NFQ-MGB	30.252	30.438	0.205	475.135			87.858
278	L14			Brain	HPRT	UNKNOWN	VIC-NFQ-MGB	30.681	30.438	0.205	362.578			87.858
279	L15			Brain	HPRT	UNKNOWN	VIC-NFQ-MGB	30.201	30.438	0.205	490.889			87.858
280	L16			Brain	HPRT	UNKNOWN	VIC-NFQ-MGB	30.265	30.438	0.205	471.341			87.858
281	L17			Brain	HPRT	UNKNOWN	VIC-NFQ-MGB	30.352	30.438	0.205	446.114			87.858
282	L18			Brain	HPRT	UNKNOWN	VIC-NFQ-MGB	30.232	30.438	0.205	481.19			87.858
283	L19			Brain	HPRT	UNKNOWN	VIC-NFQ-MGB	30.746	30.438	0.205	348.027			87.858
284	L20			Brain	HPRT	UNKNOWN	VIC-NFQ-MGB	30.312	30.438	0.205	457.733			87.858
285	L21			Brain	HPRT	UNKNOWN	VIC-NFQ-MGB	30.68	30.438	0.205	362.814			87.858
286	L22			Brain	HPRT	UNKNOWN	VIC-NFQ-MGB	30.422	30.438	0.205	426.906			87.858

Well Summary: In Plate: 384 Set Up: 192 Analyzed: 192 Flagged: 0 Omitted by Analysis: 0 Omitted Manually: 0 Samples Used: 4 Targets Used: 2

Note: To show or hide columns in the Well Table, select or deselect respectively the column name from the Show in Table drop-down menu.

Assessing the well table in your own experiments

When you analyze your own Relative Standard Curve experiment, look for standard deviation in the replicate groups (C_T SD values). If needed, omit outliers ("Improve CT precision by omitting wells" on page 65).

Confirm accurate dye signal using the Multicomponent Plot

The Multicomponent Plot screen displays the complete spectral contribution of each dye in a selected well over the duration of the PCR run.

Purpose

In the Relative Standard Curve example experiment, you review the Multicomponent Plot screen for:

- ROX™ dye (passive reference)
- FAM™ dye (reporter for RNase P)
- VIC® dye (reporter for HPRT)
- Spikes, dips, and/or sudden changes
- Amplification in the negative control wells

View the Multicomponent Plot

1. From the Experiment Menu pane, select **Analysis** ▸ **Multicomponent Plot**.

   Note: If no data are displayed, click **Analyze**.

2. Display the unknown and standard wells one at a time in the Multicomponent Plot screen:

   a. Click the **Plate Layout** tab.

   b. Select one well in the plate layout; the well is shown in the Multicomponent Plot screen.

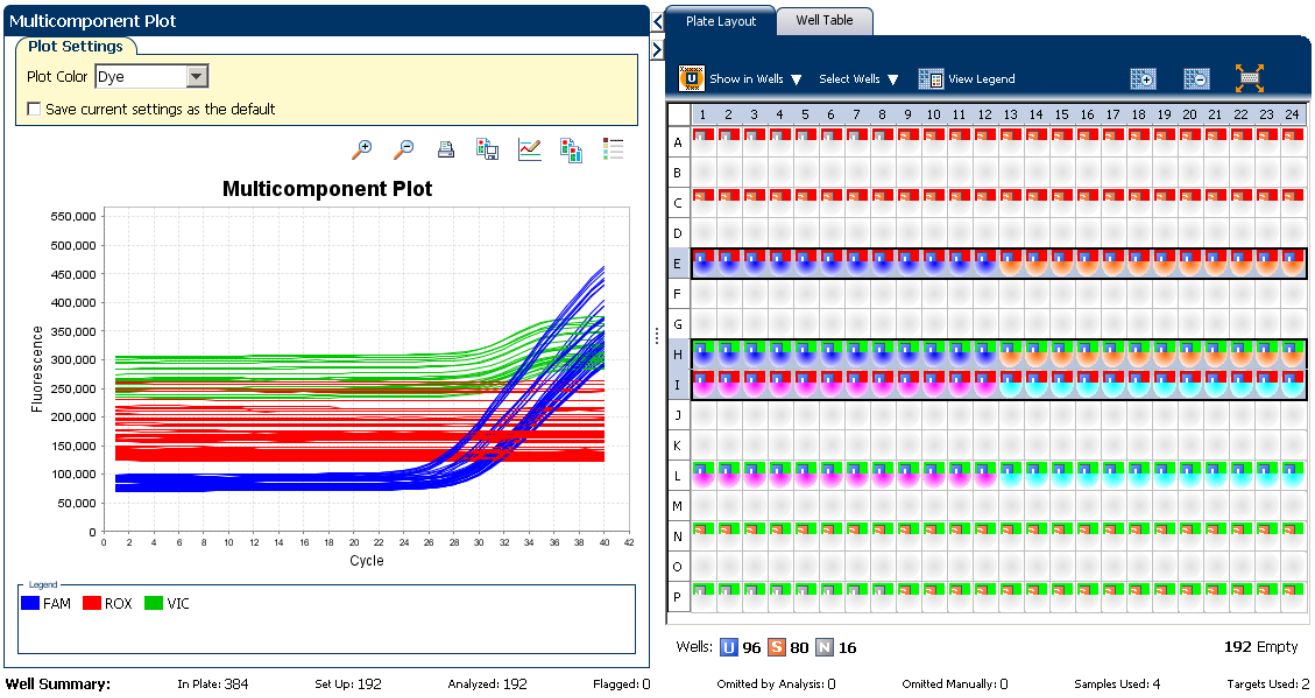
      Note: If you select multiple wells, the Multicomponent Plot screen displays the data for all selected wells simultaneously.

3. From the Plot Color drop-down menu, select **Dye**.

4. Click **Show a legend for the plot** (default).

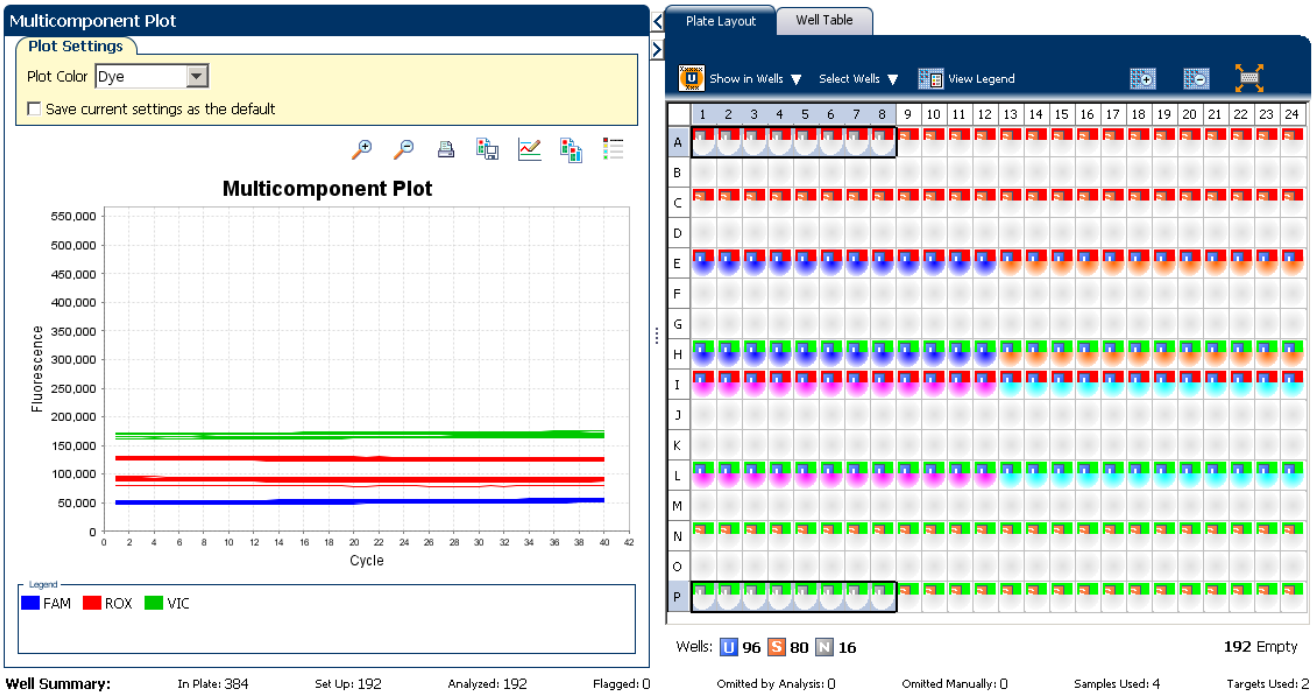
   Note: This is a toggle button. When the legend is displayed, the button changes to Hide the plot legend.

5. Check the FAM and VIC dye signals. In the example experiment, the FAM and VIC dyes signal increase throughout the PCR process, indicating normal amplification.



6. Check the ROX dye signal. In the example experiment, the ROX dye signal remains constant throughout the PCR process indicating typical data.

7. Select the negative control wells one at time and check for amplification. In the example experiment, there is no amplification in any of the negative control wells.



Tips for confirming dye accuracy in your own experiment

When you analyze your own Relative Standard Curve experiment, look for:

- **Passive reference** – The passive reference dye fluorescence level should remain relatively constant throughout the PCR process.
- **Reporter dye** – The reporter dye fluorescence level should display a flat region corresponding to the baseline, followed by a rapid rise in fluorescence as the amplification proceeds.
- **Irregularities in the signal** – There should not be any spikes, dips, and/or sudden changes in the fluorescent signal.
- **Negative control wells** – There should not be any amplification in the negative control wells.

Determine signal accuracy using the Raw Data Plot

The Raw Data Plot screen displays the raw fluorescence signal (not normalized) for each optical filter for the selected wells during each cycle of the real-time PCR.

About the example experiment

In the Relative Standard Curve example experiment, you review the Raw Data Plot screen for a stable increase in signal (no abrupt changes or dips) from the appropriate filter.

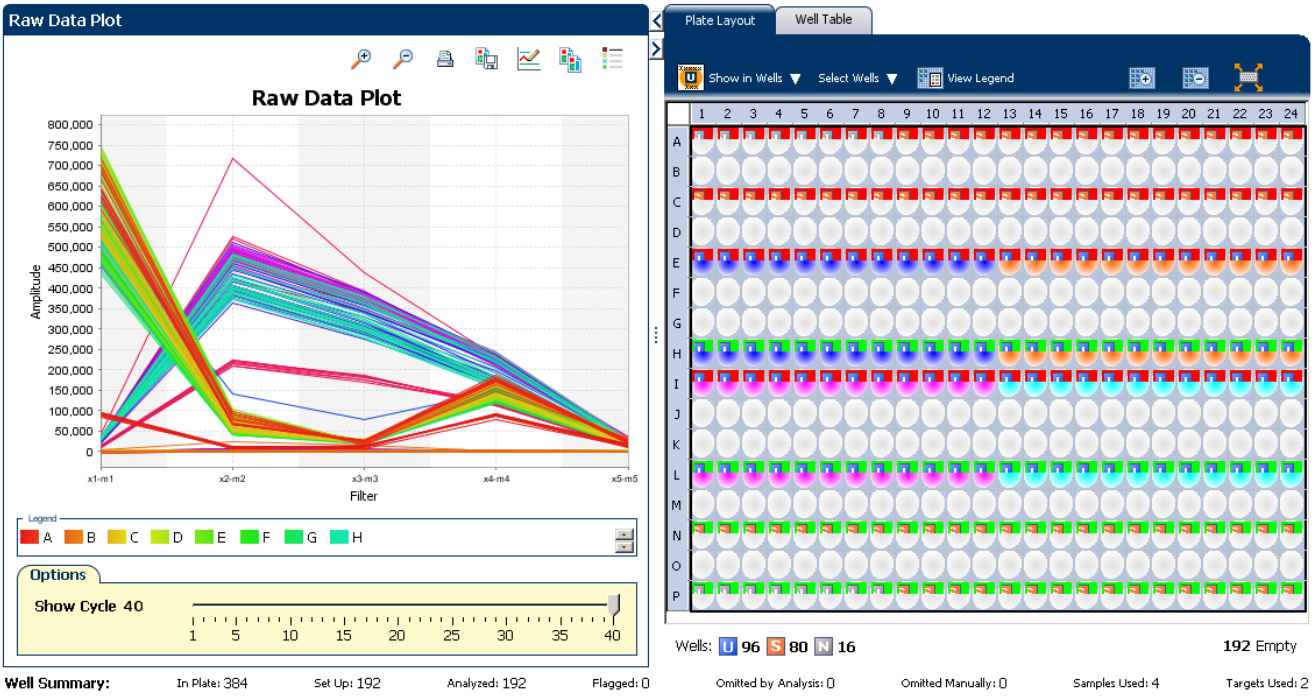
View the Raw Data Plot

1. From the Experiment Menu pane, select **Analysis ▸ Raw Data Plot**.

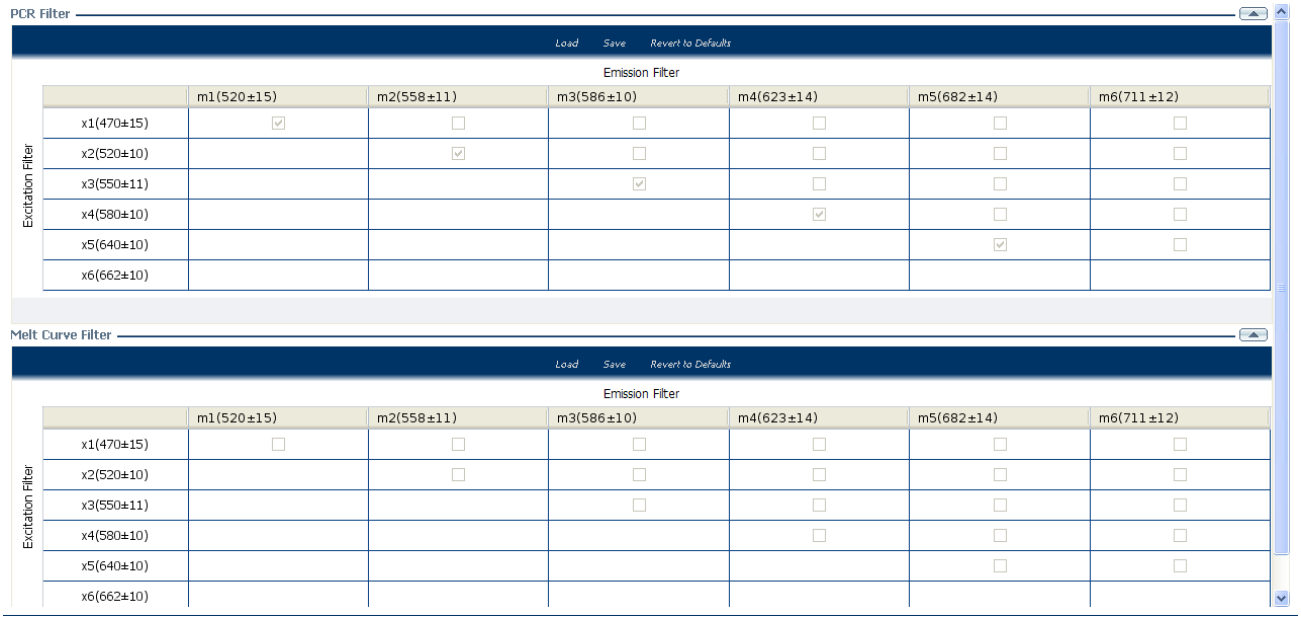
   Note: If no data are displayed, click **Analyze**.

2. Display all 384 wells in the Raw Data Plot screen by clicking the upper left corner of the plate layout in the Plate Layout tab.
3. Click  **Show a legend for the plot** (default). The legend displays the color code for each row of the reaction plate (see the legend in the Raw Data Plot shown below).

4. Click and drag the Show Cycle pointer from cycle 1 to cycle 40. In the example experiment, there is a stable increase in signal from filter 1, which corresponds to the FAM™ dye filter.



The filters used for the example experiment are:



Tips for determining signal accuracy in your own experiment

When you analyze your own Relative Standard Curve experiment, look for the following in each filter:

- Characteristic signal growth
- No abrupt changes or dips

View the endogenous control profile using the QC Plot

In the Relative Standard Curve experiment, the QC Plot screen displays the Endogenous Control Profile plot for all the targets in the experiment. The QC Plot serves as a tool to help users choose the best endogenous control for that experiment. The endogenous control profile plot is a visual display of the C_T level of the endogenous control across the sample. You can view up to four endogenous controls at a time. The sample is plotted on the X-axis, and the C_T is plotted on the Y-axis. The expression is viewed as a color and shape combination in the plot. Endogenous controls are also known as reference genes.

To view the QC Plot:

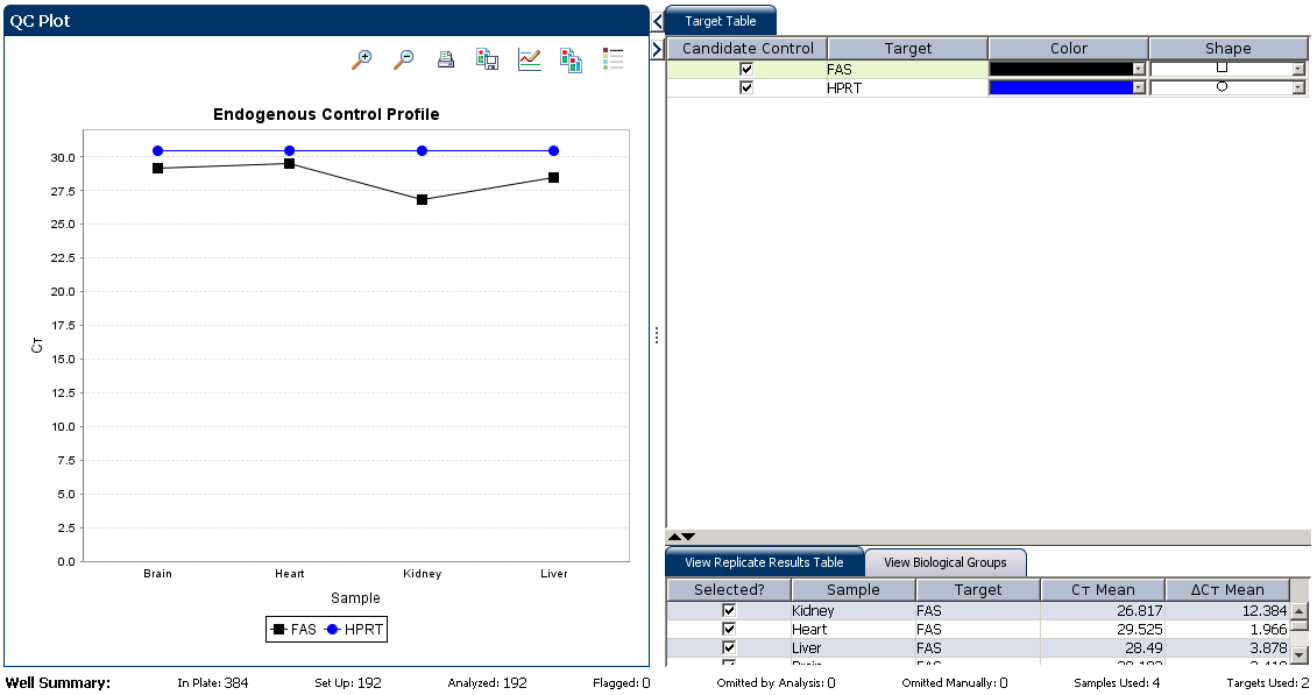
1. From the Experiment Menu pane, select **Analysis ▸ QC Plot**.

   Note: If no data are displayed, click **Analyze**.

2. In the QC Plot screen, click **Target Table** to select a target to profile:
   a. In the Candidate Control column, select the check box of the target of the endogenous control profile to plot. In the example experiment, the endogenous control is HPRT.
   b. Select a color from the Color drop-down menu.
   c. Select a shape from the Shape drop-down menu.
3. Click the **View Replicate Results Table**.
4. Select the check box of the samples you want to plot.
5. Click **Show a legend for the plot** (default).

   Note: This is a toggle button. When the legend is displayed, the button changes to Hide the plot legend.

The QC Plot in the Relative Standard Curve example experiment looks like this. Note that the endogenous control, HPRT is expressed at the same level in all the four samples:



This example experiment does not define Biological Groups.

Review the QC flags in the QC Summary

The QC Summary screen displays a list of the QuantStudio™ 6 and 7 Flex Real-Time PCR System Software flags, including the flag frequency and location for the open experiment.

View the QC Summary

1. From the Experiment Menu pane, select **Analysis** ▸ **QC Summary**.

   Note: If no data are displayed, click **Analyze**.

2. Review the Flags Summary.

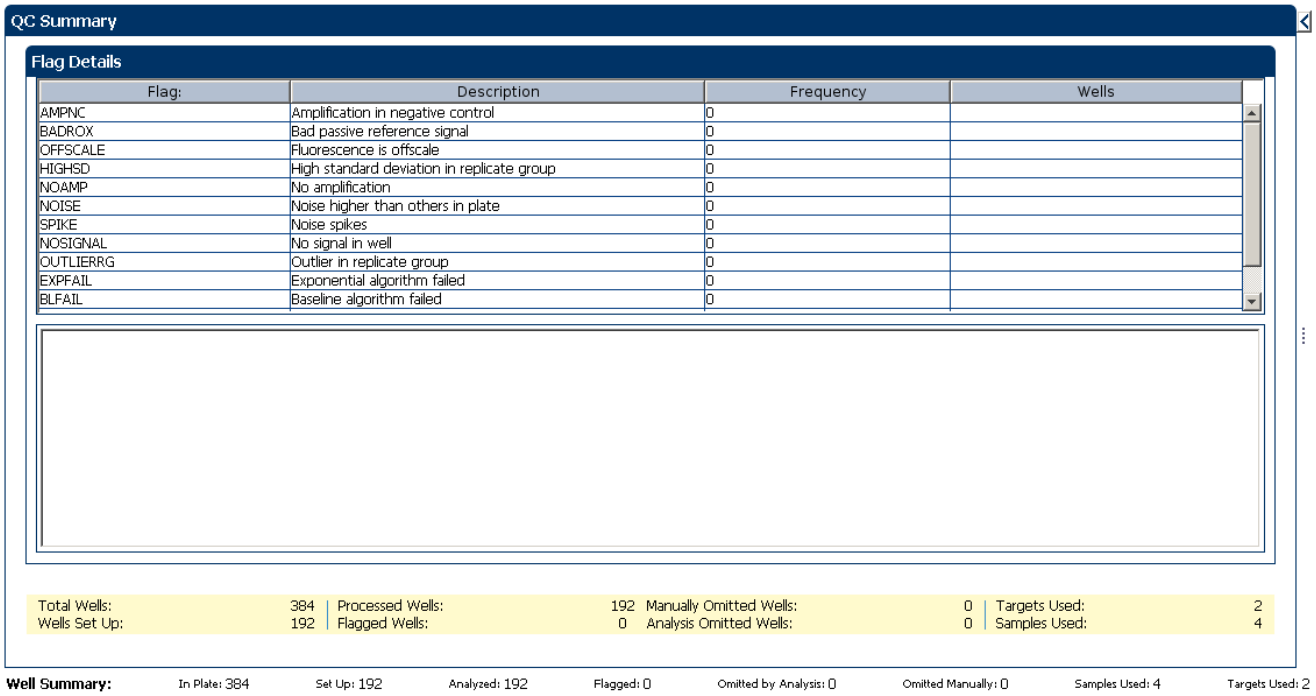
   Note: A 0 displayed in the Frequency column indicates that the flag does not appear in the experiment. If the frequency is >0, the flag appears somewhere in the experiment; the well position is listed in the Wells column.

   In the example experiment, there are no flagged wells.

3. In the Flag Details table, click each flag with a frequency > 0 to display detailed information about the flag. In the example experiment, the Frequency column displays 0 for all flags.

4. *(Optional)* For those flags with frequency > 0, click the troubleshooting link to view information on correcting the flag.

The following is an image of the QC Summary screen for the example experiment:



Possible flags

The flags listed below may be triggered by the experiment data.

Flag	Description
Pre-processing flag	
OFFSCALE	Fluorescence is offscale
Primary analysis flags	
BADROX	Bad passive reference signal
NOAMP	No amplification
NOISE	Noise higher than others in plate
SPIKE	Noise spikes
NOSIGNAL	No signal in well
EXPFAIL	Exponential algorithm failed
BLFAIL	Baseline algorithm failed
THOLDFAIL	Thresholding algorithm failed
CTFAIL	C_T algorithm failed
Secondary analysis flags	
OUTLIERRG	Outlier in replicate group
AMPNC	Amplification in negative control
HIGHSD	High standard deviation in replicate group

For more information

For more information on...	Refer to...	Publication number
Publishing data	Chapter 2 in Booklet 1, *Getting Started with QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Experiments*.	4489822

Section 5.2 Adjust parameters for re-analysis of your own experiments

Adjust analysis settings

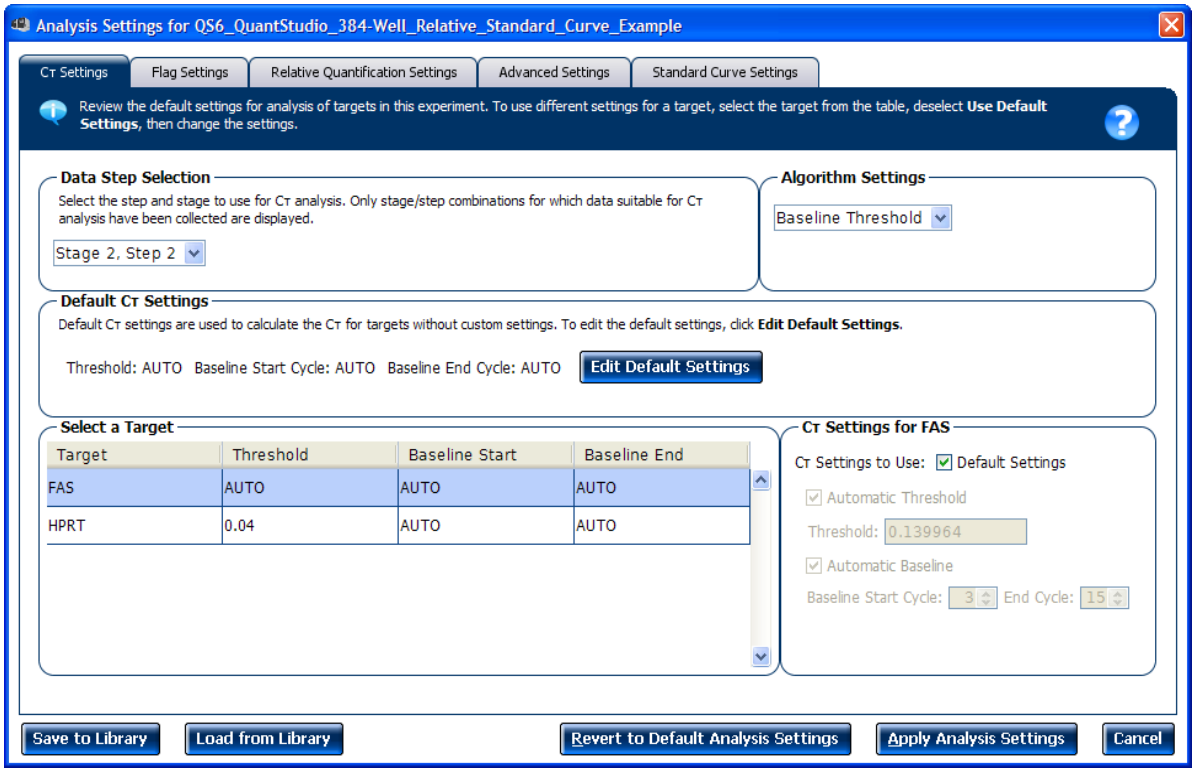
The Analysis Settings dialog box displays the analysis settings for the threshold cycle (C_T), flags, and advanced options.

If the default analysis settings in the QuantStudio™ 6 and 7 Flex Real-Time PCR System Software are not suitable for your own experiment, you can change the settings in the Analysis Settings dialog box, then reanalyze your experiment.

View the analysis settings

1. From the Experiment Menu pane, select **Analysis**.
2. Click **Analysis** ▸ **Analysis Settings** to open the Analysis Settings dialog box.
   In the example experiment, the default analysis settings are used for each tab:
   - C_T Settings
   - Flag Settings
   - Relative Quantification Settings
   - Advanced Settings
   - Standard Curve Settings

   The following is an image of the Analysis Settings dialog box for a Relative Standard Curve experiment:



3. View and, if necessary, change the analysis settings (see "Adjust analysis settings" below).

   Note: You can save the changes to the analysis settings to the Analysis Settings Library for later use. For more information, see About the Analysis Settings Library in Booklet 1, *Getting Started with QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Experiments*.

4. Click **Apply Analysis Settings** to apply the current analysis settings.

   Note: You can go back to the default analysis settings, by clicking **Revert to Default Analysis Settings**.

Adjust analysis settings

C_T Settings

- **Data Step Selection**

  Use this feature to select one stage/step combination for C_T analysis when there is more than one data collection point in the run method**.**

- **Algorithm Settings**

  Use the Baseline Threshold algorithm to determine the C_T values.

  The Baseline Threshold algorithm is an expression estimation algorithm that subtracts a baseline component and sets a fluorescent threshold in the exponential region for gene quantification.

- **Default C_T Settings**

  Use the default C_T settings feature to calculate C_T for the targets that do not have custom settings. To edit the default settings, click **Edit Default Settings**.

- **C_T Settings for Target**

  When you manually set the threshold and baseline, Life Technologies recommends:

Setting	Recommendation
Threshold	Enter a value for the threshold so that the threshold is: • Above the background. • Below the plateau and linear regions of the amplification curve. • Within the exponential phase of the amplification curve.
Baseline	Select the Start Cycle and End Cycle values so that the baseline ends before significant fluorescent signal is detected.

  Note: This setting is applicable only to the Baseline Threshold algorithm.

  Note: Selecting Automatic Threshold implies selection of automatic setting of the baseline. However, if Automatic Threshold is deselected, then you can choose between setting the baseline either automatically or manually.

Flag Settings

Use the Flag Settings tab to:

- Adjust the sensitivity so that more wells or fewer wells are flagged.
- Change the flags that are applied by the QuantStudio™ 6 and 7 Flex Real-Time PCR System Software.

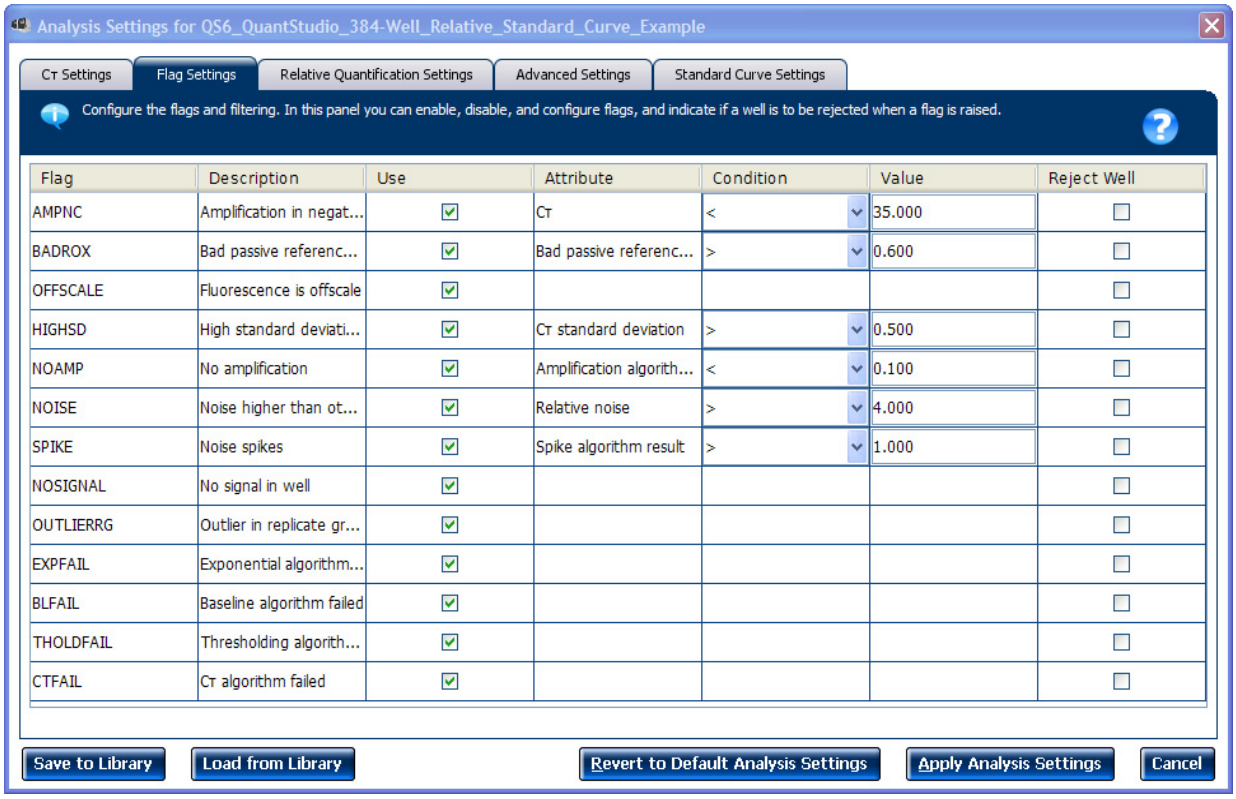
To adjust the flag settings

1. In the Use column, select the check boxes for flags to apply during analysis.
2. *(Optional)* If an attribute, condition, and value are listed for a flag, specify the setting for applying the flag.

   Note: If you choose to adjust the setting for applying a flag, make minor adjustments as you evaluate the appropriate setting.
3. In the Reject Well column, select the check boxes if you want the software to reject wells with the flag.

   Note: After you have rejected the flagged wells, analysis results depend on factors such as the experiment type and flag type. For example, rejecting wells flagged by HIGHSD in experiments using the Standard Deviation calculations may change the result of C_T SD. For some flags, analysis results calculated before the well is rejected are maintained.
4. Click **Apply Analysis Settings** in the Analysis Settings dialog box. If the run status is complete, the data are reanalyzed.

The following is an image of the Flag Settings tab:



Relative Quantification Settings

Use the Relative Quantification Settings tab to:

- Change the type of analysis, singleplex or multiplex.
- Change the reference sample and/or endogenous control.

- Reject Outliers with ΔC_T values less than or equal to the entered value.

  Note: The Outlier Rejection settings apply only to multiplex reactions.

- Select the algorithm to use to determine the relative quantification minimum and maximum values (error bars):
  - **Confidence Level** - Select to calculate the RQ minimum and maximum values based on the selected confidence level. Select the confidence level to use.
  - **Standard Deviations** - Select to calculate the RQ minimum and maximum values based on the selected number of standard deviations. Select the number of standard deviations to use.

Advanced Settings

Use the Advanced Settings tab to change baseline settings well-by-well.

To use custom baseline settings for a well-target combination:

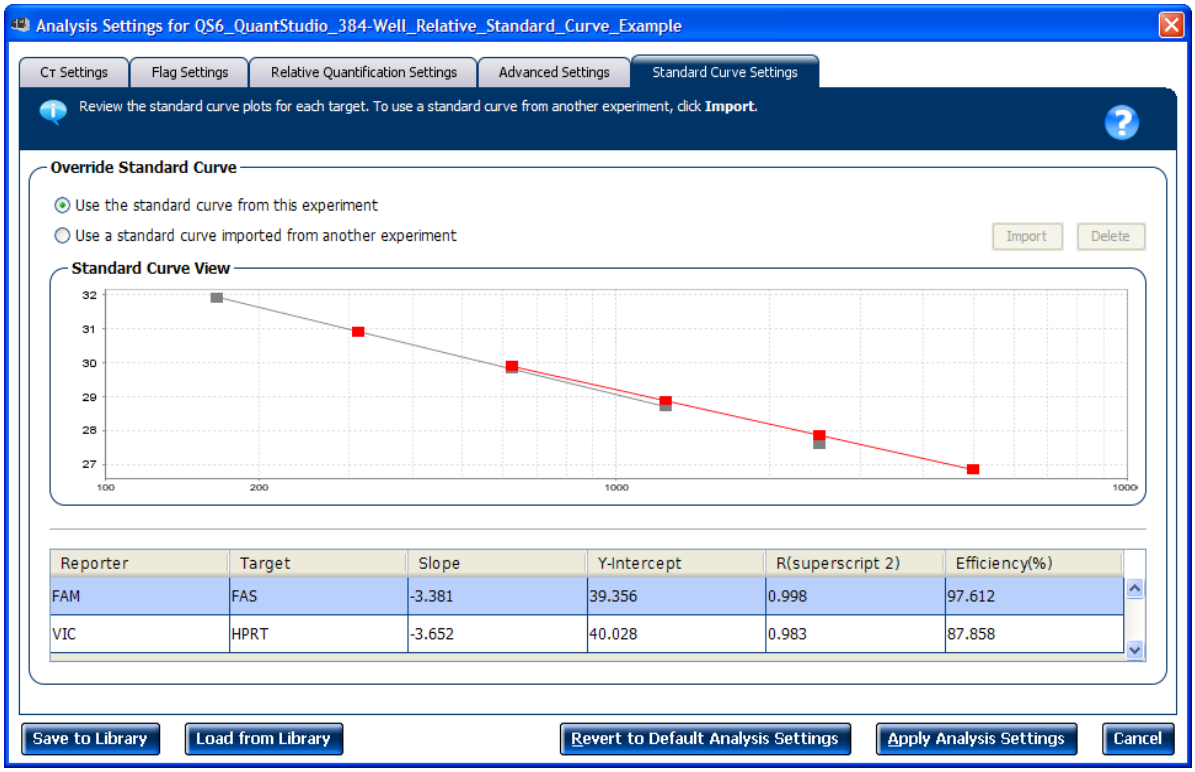
1. Select one or more well-target combinations in the table.
2. Deselect the **Use C_T Settings Defined for Target** check box.
3. Define the custom baseline settings:
   - For automatic baseline calculations, select the **Automatic Baseline** check box.
   - To define the baseline, deselect the **Automatic Baseline** check box, then enter the baseline start cycle and baseline end cycle.

Standard Curve Settings

Use this tab to review the settings of the current standard curve experiment or to import the standard curve from an external experiment (with the same samples and targets) and apply it to this current experiment.

Note: The run method must be the same. Life Technologies recommends using the standard curve from the current experiment.

For the example experiment, the settings from the current experiment have been used.



Improve C_T precision by omitting wells

Experimental error may cause some wells to be amplified insufficiently or not at all. These wells typically produce C_T values that differ significantly from the average for the associated replicate wells. If included in the calculations, these outliers can result in erroneous measurements; to ensure C_T precision, omit the outliers from the analysis.

Note: In the Relative Standard Curve example experiment, there are no outliers. No wells need to be removed from analysis.

1. From the Experiment Menu pane, select **Analysis ▸ Amplification Plot**.

   Note: If no data are displayed, click **Analyze**.

2. In the Amplification Plot screen, select **C_T vs. Well** from the Plot Type drop-down menu.

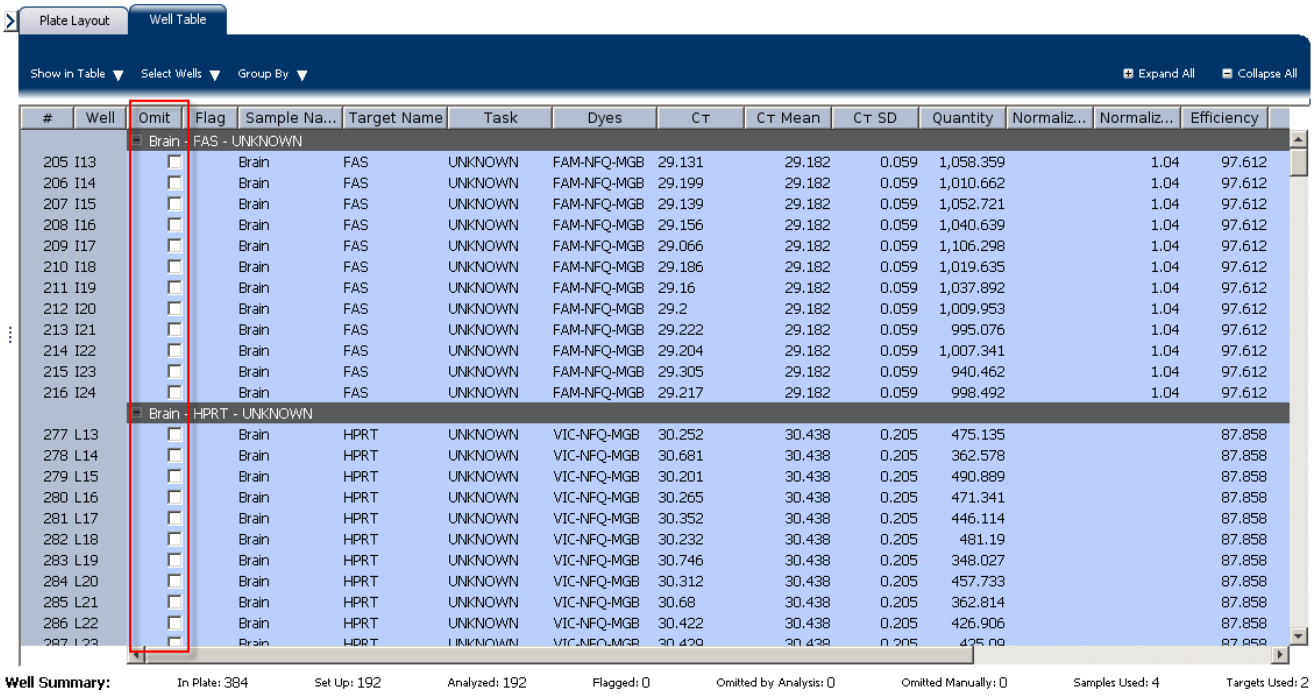
3. Select the **Well Table** tab, select replicates to omit:

4. In the Well Table:

   a. From the Group By drop-down menu, select **Replicate**.

   b. Look for outliers in the replicate group (make sure they are flagged).

c. Select the **Omit** check box next to outlying well(s), as shown in the following image.



#	Well	Omit	Flag	Sample Na...	Target Name	Task	Dyes	CT	CT Mean	CT SD	Quantity	Normaliz...	Normaliz...	Efficiency
			Brain - FAS - UNKNOWN											
205	I13			Brain	FAS	UNKNOWN	FAM-NFQ-MGB	29.131	29.182	0.059	1,058.359		1.04	97.612
206	I14			Brain	FAS	UNKNOWN	FAM-NFQ-MGB	29.199	29.182	0.059	1,010.662		1.04	97.612
207	I15			Brain	FAS	UNKNOWN	FAM-NFQ-MGB	29.139	29.182	0.059	1,052.721		1.04	97.612
208	I16			Brain	FAS	UNKNOWN	FAM-NFQ-MGB	29.156	29.182	0.059	1,040.639		1.04	97.612
209	I17			Brain	FAS	UNKNOWN	FAM-NFQ-MGB	29.066	29.182	0.059	1,106.298		1.04	97.612
210	I18			Brain	FAS	UNKNOWN	FAM-NFQ-MGB	29.186	29.182	0.059	1,019.635		1.04	97.612
211	I19			Brain	FAS	UNKNOWN	FAM-NFQ-MGB	29.16	29.182	0.059	1,037.892		1.04	97.612
212	I20			Brain	FAS	UNKNOWN	FAM-NFQ-MGB	29.2	29.182	0.059	1,009.953		1.04	97.612
213	I21			Brain	FAS	UNKNOWN	FAM-NFQ-MGB	29.222	29.182	0.059	995.076		1.04	97.612
214	I22			Brain	FAS	UNKNOWN	FAM-NFQ-MGB	29.204	29.182	0.059	1,007.341		1.04	97.612
215	I23			Brain	FAS	UNKNOWN	FAM-NFQ-MGB	29.305	29.182	0.059	940.462		1.04	97.612
216	I24			Brain	FAS	UNKNOWN	FAM-NFQ-MGB	29.217	29.182	0.059	998.492		1.04	97.612
			Brain - HPRT - UNKNOWN											
277	L13			Brain	HPRT	UNKNOWN	VIC-NFQ-MGB	30.252	30.438	0.205	475.135			87.858
278	L14			Brain	HPRT	UNKNOWN	VIC-NFQ-MGB	30.681	30.438	0.205	362.578			87.858
279	L15			Brain	HPRT	UNKNOWN	VIC-NFQ-MGB	30.201	30.438	0.205	490.889			87.858
280	L16			Brain	HPRT	UNKNOWN	VIC-NFQ-MGB	30.265	30.438	0.205	471.341			87.858
281	L17			Brain	HPRT	UNKNOWN	VIC-NFQ-MGB	30.352	30.438	0.205	446.114			87.858
282	L18			Brain	HPRT	UNKNOWN	VIC-NFQ-MGB	30.232	30.438	0.205	481.19			87.858
283	L19			Brain	HPRT	UNKNOWN	VIC-NFQ-MGB	30.746	30.438	0.205	348.027			87.858
284	L20			Brain	HPRT	UNKNOWN	VIC-NFQ-MGB	30.312	30.438	0.205	457.733			87.858
285	L21			Brain	HPRT	UNKNOWN	VIC-NFQ-MGB	30.68	30.438	0.205	362.814			87.858
286	L22			Brain	HPRT	UNKNOWN	VIC-NFQ-MGB	30.422	30.438	0.205	426.906			87.858

5. Click **Analyze** to reanalyze the experiment data with the outlying well(s) removed from the analysis.

   Note: You can also omit undesirable wells in an experiment from the Plate Layout screen. To omit a well from the Plate Layout screen, right-click the well and select **Omit**.

For more information

For more information on	Refer to	Publication number
Amplification efficiency	*Amplification Efficiency of TaqMan® Gene Expression Assays Application Note*	127AP05-03

6 Export Analysis Results

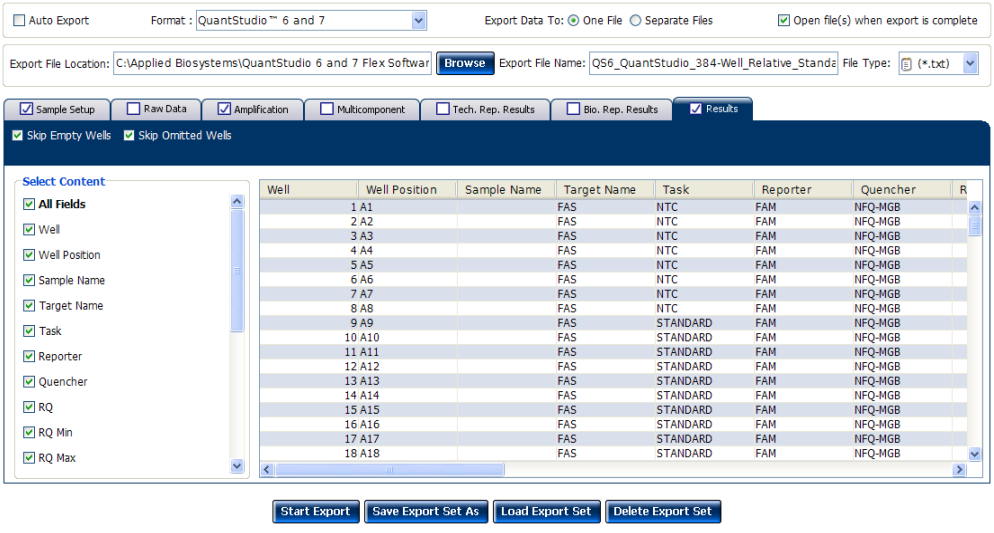
1. Open the Relative Standard Curve example experiment file that you analyzed in Chapter 5.
2. In the Experiment Menu, click  **Export.**

   Note: To export data automatically after analysis, select the **Auto Export** check box during experiment setup or before running the experiment. Auto export is unchecked for the example experiment.
3. Select **QuantStudio™ 6 and 7** format.

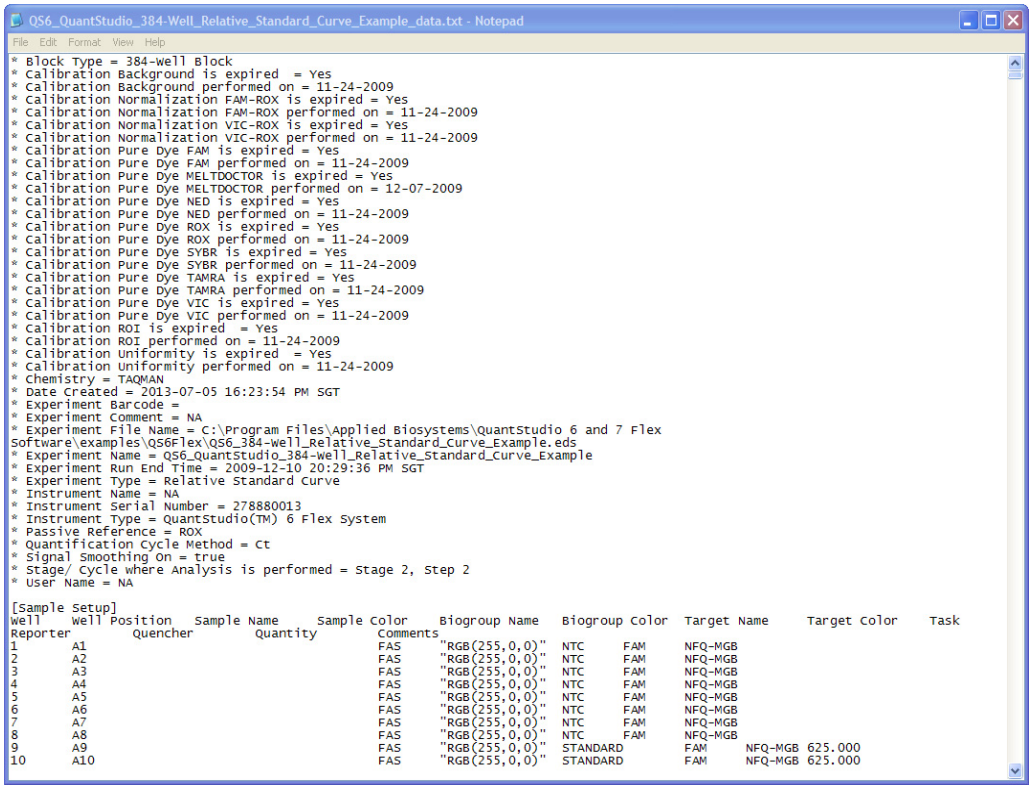
   Note: Select **7900** Format if you want to export the Clipped Data.
4. Complete the Export dialog box as shown below:

Field or Selection	Entry
Select Data to export/ Select Content	Results
Export Data To	One File
Export File Name	QS6_QuantStudio_384-Well_Relative_Standard_Curve_Example_data
File Type	*.txt
Export File Location	C:\Applied Biosystems\QuantStudio 6 and 7 Flex Software\User Files\Export

Your Export screen should look like this:



Your exported file when opened in Notepad should look like this:



QS6_QuantStudio_384-Well_Relative_Standard_Curve_Example_data.txt - Notepad

File Edit Format View Help

```
* Block Type = 384-Well Block
* Calibration Background is expired  = Yes
* Calibration Background performed on = 11-24-2009
* Calibration Normalization FAM-ROX is expired = Yes
* Calibration Normalization FAM-ROX performed on = 11-24-2009
* Calibration Normalization VIC-ROX is expired = Yes
* Calibration Normalization VIC-ROX performed on = 11-24-2009
* Calibration Pure Dye FAM is expired = Yes
* Calibration Pure Dye FAM performed on = 11-24-2009
* Calibration Pure Dye MELTDOCTOR is expired = Yes
* Calibration Pure Dye MELTDOCTOR performed on = 12-07-2009
* Calibration Pure Dye NED is expired = Yes
* Calibration Pure Dye NED performed on = 11-24-2009
* Calibration Pure Dye ROX is expired = Yes
* Calibration Pure Dye ROX performed on = 11-24-2009
* Calibration Pure Dye SYBR is expired = Yes
* Calibration Pure Dye SYBR performed on = 11-24-2009
* Calibration Pure Dye TAMRA is expired = Yes
* Calibration Pure Dye TAMRA performed on = 11-24-2009
* Calibration Pure Dye VIC is expired = Yes
* Calibration Pure Dye VIC performed on = 11-24-2009
* Calibration ROI is expired  = Yes
* Calibration ROI performed on = 11-24-2009
* Calibration Uniformity is expired  = Yes
* Calibration Uniformity performed on = 11-24-2009
* Chemistry = TAQMAN
* Date Created = 2013-07-05 16:23:54 PM SGT
* Experiment Barcode =
* Experiment Comment = NA
* Experiment File Name = C:\Program Files\Applied Biosystems\QuantStudio 6 and 7 Flex
Software\examples\QS6Flex\QS6_384-Well_Relative_Standard_Curve_Example.eds
* Experiment Name = QS6_QuantStudio_384-Well_Relative_Standard_Curve_Example
* Experiment Run End Time = 2009-12-10 20:29:36 PM SGT
* Experiment Type = Relative Standard Curve
* Instrument Name = NA
* Instrument Serial Number = 278880013
* Instrument Type = QuantStudio(TM) 6 Flex System
* Passive Reference = ROX
* Quantification Cycle Method = Ct
* Signal Smoothing On = true
* Stage/ Cycle where Analysis is performed = Stage 2, Step 2
* User Name = NA

[Sample Setup]
Well    Well Position   Sample Name     Sample Color    Biogroup Name   Biogroup Color  Target Name     Target Color    Task
Reporter        Quencher        Quantity        Comments
1       A1                                      FAS     "RGB(255,0,0)"  NTC     FAM     NFQ-MGB
2       A2                                      FAS     "RGB(255,0,0)"  NTC     FAM     NFQ-MGB
3       A3                                      FAS     "RGB(255,0,0)"  NTC     FAM     NFQ-MGB
4       A4                                      FAS     "RGB(255,0,0)"  NTC     FAM     NFQ-MGB
5       A5                                      FAS     "RGB(255,0,0)"  NTC     FAM     NFQ-MGB
6       A6                                      FAS     "RGB(255,0,0)"  NTC     FAM     NFQ-MGB
7       A7                                      FAS     "RGB(255,0,0)"  NTC     FAM     NFQ-MGB
8       A8                                      FAS     "RGB(255,0,0)"  NTC     FAM     NFQ-MGB
9       A9                                      FAS     "RGB(255,0,0)"  STANDARD        FAM     NFQ-MGB 625.000
10      A10                                     FAS     "RGB(255,0,0)"  STANDARD        FAM     NFQ-MGB 625.000
```

PART 2
Running Comparative C_T Experiments

7 About Comparative C_T Experiments

This chapter covers:

IMPORTANT! First-time users of the QuantStudio™ 6 and 7 Flex Real-Time PCR System Software, please read Booklet 1, *Getting Started with QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Experiments* and Booklet 7, *QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Experiments - Appendixes* of this binder thoroughly. The booklets provide information and general instructions that are applicable to all the experiments described in this binder.

Note: For more information about any of the topics discussed in this guide, access the Help from within QuantStudio™ 6 and 7 Flex Software by pressing **F1**, clicking in the toolbar, or selecting **Help** ▸ QuantStudio™ 6 and 7 Flex Software **Help**.

About Comparative C_T experiments

The Comparative CT ($\Delta\Delta C_T$) method is used to determine the relative target quantity in samples. With the comparative C_T method, the QuantStudio™ 6 and 7 Flex Software measures amplification of the target and of the endogenous control in samples and in a reference sample. Measurements are normalized using the endogenous control. The software determines the relative quantity of target in each sample by comparing normalized C_T (ΔC_T) in each sample to normalized C_T (ΔC_T) in the reference sample.

Comparative C_T experiments are commonly used to:

- Compare expression levels of a gene in different tissues.
- Compare expression levels of a gene in a treated sample and an untreated sample.
- Compare expression levels of wild-type alleles and mutated alleles.
- Analyze the gene expression changes over time under specific treatment conditions.

The Relative Standard Curve method determines the relative target quantity in samples. The QuanStudio™ 6 and 7 Flex Software measures amplification of the target and of the endogenous control in samples, in a reference sample, and in a standard dilution series. For more information on Relative Standard Curve experiments, refer to Part I, Running Relative Standard Curve Experiments of this booklet.

Assemble required components

- **Sample** – The tissue group that you are testing for a target gene.
- **Reference sample (also called a calibrator)**– The sample used as the basis for relative quantification results. For example, in a study of drug effects on gene expression, an untreated control is an appropriate reference sample.

- **Endogenous control** – A gene that is used to normalize template input differences, and sample-to-sample or run-to-run variation.
- **Replicates** – The total number of identical reactions containing identical components and identical volumes.
- **Negative controls** – Wells that contain water or buffer instead of sample template. No amplification of the target should occur in negative control wells.

PCR options

When performing real-time PCR, choose between:

- Singleplex and multiplex PCR (below)

  and

- 1-step and 2-step RT-PCR (page 72)

Singleplex and Multiplex PCR

You can perform a PCR reaction using either:

- **Singleplex PCR** – In singleplex PCR a single primer and probe set is present in the reaction tube or well. Only one target or endogenous control can be amplified per reaction.

  Or

- **Multiplex PCR** – In multiplex PCR, two or more primer and probe sets are present in the reaction tube or well. Each set amplifies a specific target or endogenous control. Typically, a probe labeled with FAM™ dye detects the target and a probe labeled with VIC® dye detects the endogenous control.

IMPORTANT! SYBR® Green reagents cannot be used for multiplex PCR.

1- and 2-Step RT-PCR

You can perform reverse transcription (RT) and PCR in a single reaction (1-step) or in separate reactions (2-step). The reagent configuration you use depends on whether you are performing 1- or 2-step RT-PCR:

- **1-step RT-PCR**– In 1-step RT-PCR, RT and PCR take place in one buffer system. Using one buffer system provides the convenience of a single-tube preparation for RT and PCR amplification. However, you cannot use Fast PCR Master Mix or the carryover prevention enzyme, AmpErase® UNG (uracil-N-glycosylase), to perform 1-step RT-PCR.
- **2-step RT-PCR**– 2-step RT-PCR is performed in two separate reactions: First, total RNA is reverse-transcribed into cDNA, then the cDNA is amplified by PCR. This method is useful for detecting multiple transcripts from a single cDNA template or for storing cDNA aliquots for later use. The AmpErase® UNG enzyme can be used to prevent carryover contamination.

About the example experiment

To illustrate how to perform comparative C_T experiment, this guide leads you through the process of designing and performing an example experiment. The example experiment represents a typical setup that you can use to quickly familiarize yourself with the QuantStudio™ 6 and 7 Flex Software.

The objective of the comparative C_T example experiment is to compare the expression of GH1, LPIN1, LIPC, GAPDH, and ACTB in liver, heart, brain, and lung tissues.

- The samples are liver, heart, lung, and brain tissues.
- The targets are GH1, LPIN1, LIPC, GAPDH, and ACTB.
- The reference sample is brain.
- The endogenous control is ACTB.
- The experiment is designed for singleplex PCR, where the targets and endogenous control assays are performed in separate wells.
- Reactions are set up for 2-step RT-PCR. The Invitrogen VILO Kit is used for reverse transcription; the TaqMan® Fast Universal PCR Master Mix is used for PCR.
- Primer and probe sets are selected from the Life Technologies TaqMan® Gene Expression Assays product line:
  - GH1 Assay Mix: Hs00236859_m1
  - LPIN1 Assay Mix: Hs00299515_m1
  - LIPC Assay Mix: Hs00165106_m1
  - GAPDH Assay Mix: Hs99999905_m1
  - ACTB Assay Mix: Hs99999903_m1

8 Design the Experiment

This chapter explains how to design the example experiment from the Experiment Setup menu.

This chapter covers:

Note: To automatically export the analyzed data to a specified location, select the Auto Export task from the Export screen, before running the experiment. For more information on Auto Export, refer to Chapter 2 in Booklet 1, *Getting Started with QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Experiments*.

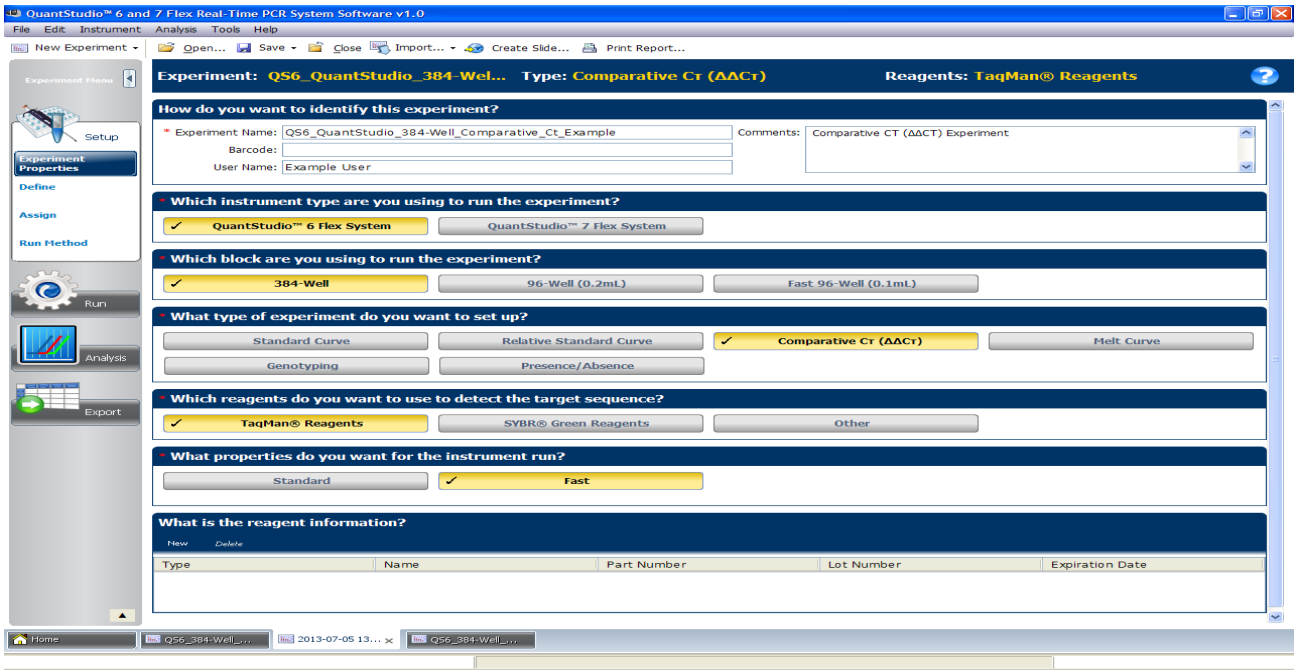
Define the experiment properties

Click **Experiment Setup** ▸ **Experiment Properties** to create a new experiment in the QuantStudio™ 6 and 7 Flex Software. Enter:

Field or Selection	Entry
Experiment Name	QS6_QuantStudio_384-Well_Comparative_Ct_Example
Barcode	Leave field empty
User Name	Example User
Comments	Comparative C_T ($\Delta\Delta C_T$) example
Instrument type	QuantStudio™ 6 Flex System
Block	384-Well Block
Experiment Type	Comparative C_T ($\Delta\Delta C_T$)
Reagents	TaqMan® Reagents
Ramp speed	Fast
Reagent information	NA

Save the experiment.

Your Experiment Properties screen should look like this:



Define targets, samples and biological replicates

Click **Define** to access the Define screen. Enter:

1. Targets

Target name	Reporter	Quencher	Color
ACTB	FAM	NFQ-MGB	
GAPDH	FAM	NFQ-MGB	
GH1	FAM	NFQ-MGB	
LPIN1	FAM	NFQ-MGB	
LIPC	FAM	NFQ-MGB	

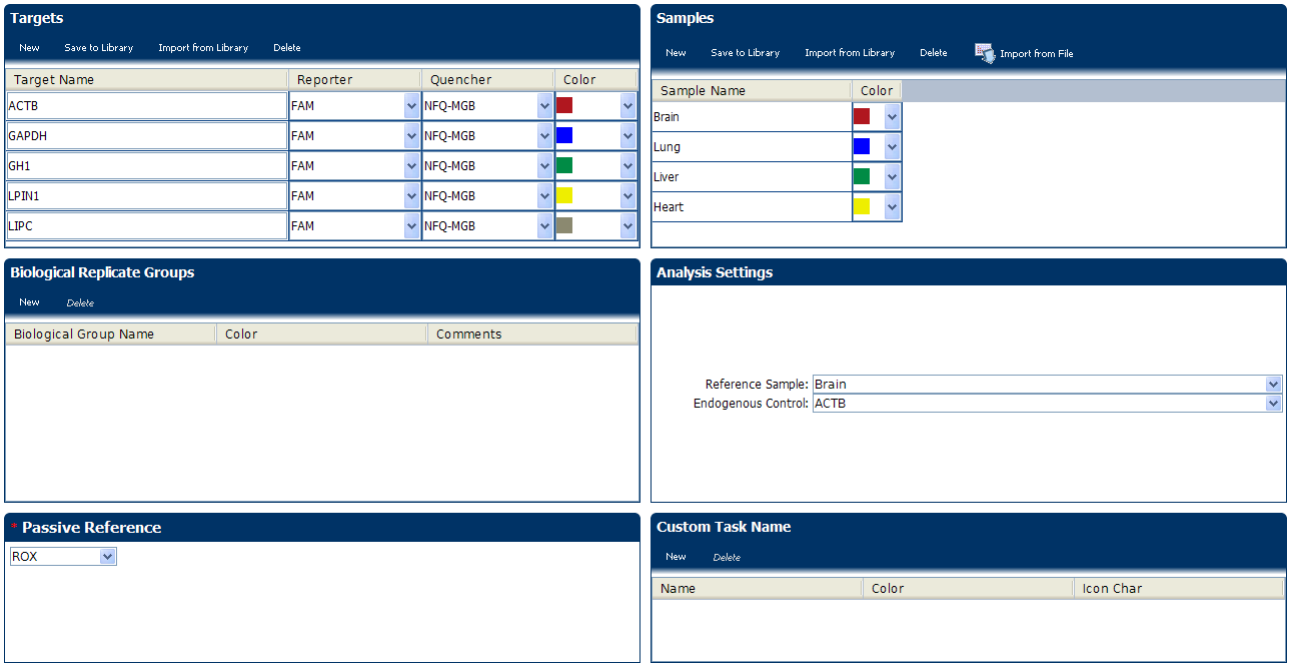
2. Samples

Sample Name	Color
Brain	
Lung	
Liver	
Heart	

3. Dye to be used as a Passive Reference
   ROX
4. Custom Task Name
   Not applicable
5. Analysis Settings

Field	Select
Reference Sample	Brain
Endogenous Control	ACTB

Your Define screen should look like this:



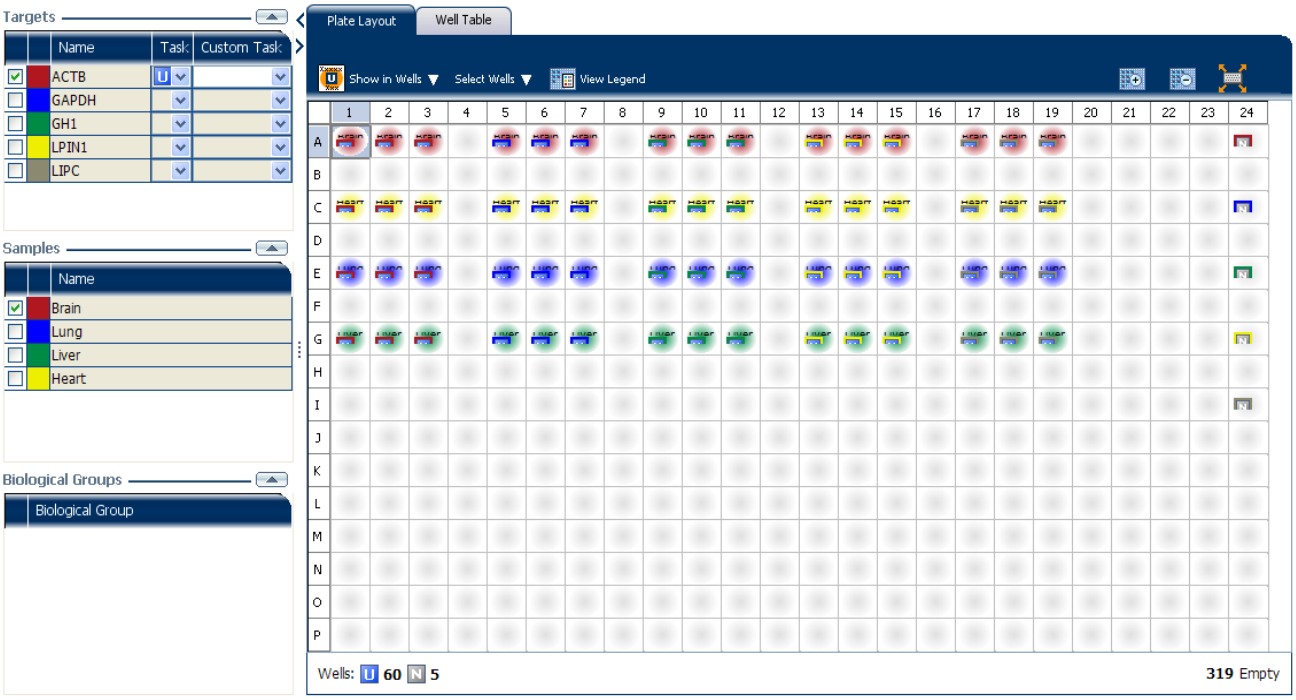
Note: This example experiment does not define biological replicate groups. Leave Biological Replicate Groups blank.

Assign targets, samples and biological groups

Click **Assign** to access the Assign screen. Enter the targets and samples:

Target name	Well number	Task	Sample name
ACTB	A1, A2, A3	Unknown	Brain
	C1, C2, C3	Unknown	Heart
	E1, E2, E3	Unknown	Lung
	G1, G2, G3	Unknown	Liver
	A24	Negative	None
GAPDH	A5, A6, A7	Unknown	Brain
	C5, C6, C7	Unknown	Heart
	E5, E6, E7	Unknown	Lung
	G5, G6, G7	Unknown	Liver
	C24	Negative	None
GH1	A9, A10, A11	Unknown	Brain
	C9, C10, C11	Unknown	Heart
	E9, E10, E11	Unknown	Lung
	G9, G10, G11	Unknown	Liver
	E24	Negative	None
LPIN1	A13, A14, A15	Unknown	Brain
	C13, C14, C15	Unknown	Heart
	E13, E14, E15	Unknown	Lung
	G13, G14, G15	Unknown	Liver
	G24	Negative	None
LIPC	A17, A18, A19	Unknown	Brain
	C17, C18, C19	Unknown	Heart
	E17, E18, E19	Unknown	Lung
	G17, G18, G19	Unknown	Liver
	I24 (Row I, Column 24)	Negative	None

Your Assign screen should look like this:



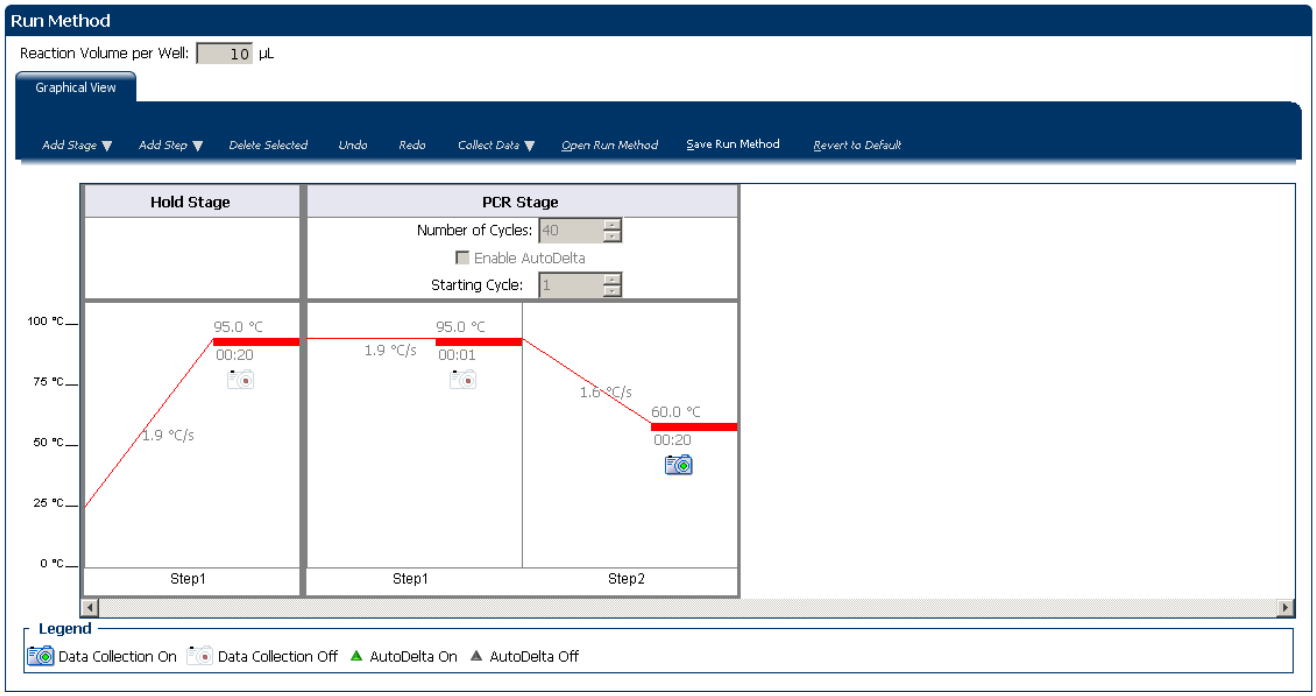
Set up the run method

Click **Run Method** to access the Run Method screen. Set the thermal profile under the Graphical View tab. Enter:

- Reaction Volume Per Well: 10 µL
- Thermal Profile

Stage	Step	Ramp rate	Temperature	Time
Hold Stage	Step 1	1.9°C/s	95°C	20 seconds
PCR Stage	Step 1	1.9°C/s	95°C	1 second
Number of Cycles: 40 Enable AutoDelta: Unchecked (default) Starting Cycle: Disabled when Enable AutoDelta is unchecked	Step 2	1.6°C/s	60°C	20 seconds

Your Run Method screen should look like this:



Tips for designing your own experiment

Life Technologies recommends that you:

- Identify each target assay with a unique name and color. You can enter up to 100 characters in the Target Name field.
- Identify each sample using a unique name and color. You can enter up to 100 characters in the Sample Name field.
- Select an endogenous control for each sample. The endogenous control is a target that is present in all samples under investigation. It should be expressed equally in all sample types, regardless of treatment or tissue origin (examples of endogenous controls are β-actin, GAPDH, and 18S ribosomal RNA [18S rRNA]). The endogenous control is used to normalize the PCR results; the endogenous control corrects for variable sample mass, nucleic acid extraction efficiency, reverse transcription efficiency, and pipette calibration errors. Note that:
  - Each sample type (for example, each tissue in a study comparing multiple tissues) requires an endogenous control.
  - If samples are spread across multiple plates, each plate must have an endogenous control. Additionally, every plate must include an endogenous control for every sample type on the plate.
- Select an endogenous control from your previously defined target assays. Amplification results from the endogenous control are used to normalize the amplification results from the target for differences in the amount of template added to each reaction.
- Select a reference sample from your previously defined samples. Amplification results from the samples and from the reference sample are compared to determine relative expression.

For more information

For more information on	Refer to	Publication number
Consumables	Chapter 1 in Booklet 1, *Getting Started with QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Experiments* Appendix A in Booklet 7, *QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Experiments - Appendixes*	4489822
Using the Standard Curve quantification methods	Booklet 2, *Running Standard Curve Experiments*.	4489822
Using the relative standard curve quantification method	Part 1 of this booklet	4489822
Selecting an endogenous control	Application Note *Using TaqMan® Endogenous Control Assays to Select an Endogenous Control for Experimental Studies*	127AP05-03
Reference samples (also known as calibrators) and endogenous controls	*User Bulletin #2: Relative quantification of Gene Expression*	4303859
Using alternative setup	Chapter 3 in Booklet 1, *Getting Started with QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Experiments*	4489822

9 Prepare the Reactions

This chapter explains how to prepare the PCR reactions for the Comparative C_T ($\Delta\Delta C_T$) example experiment.

This chapter covers:

Assemble required materials

- Items listed in Booklet 1, *Getting Started with QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Experiments*.
- Samples - Total RNA isolated from liver, heart, brain, and lung tissues.
- Example experiment reaction mix components:
  - TaqMan® Fast Universal PCR Master Mix (2X.)
  - ACTB Assay Mix (20X)
  - GAPDH Assay Mix (20X)
  - GH1 Assay Mix (20X)
  - LIPN1 Assay Mix (20X)
  - LIPC Assay Mix (20X)

Note: Fast Universal Master Mix is not recommended to be used with the 96-well (0.2 mL) reaction plates or reaction tubes and tube strips sealed with caps.

Prepare the template

Prepare the template for the PCR reactions using the High-Capacity cDNA Reverse Transcription Kit or one of the Invitrogen VILO kits to carry out the reverse transcription.

Example experiment settings

For the Comparative C_T example experiment, the template for the PCR reactions is cDNA reverse-transcribed from total RNA samples using one of the Invitrogen VILO kits, SuperScript® VILO™ cDNA Synthesis Kit (Part no. 4453650).

Prepare the template

Use the Invitrogen VILO kits to reverse-transcribe cDNA from the total RNA samples. Follow the procedures in the *Invitrogen VILO Kits Protocol* (Pub. no. 100002284) to:

1. Prepare the RT master mix.
2. Prepare the cDNA reactions.
3. Perform reverse transcription on a thermal cycler.

Prepare the sample dilutions

For the Comparative C_T example experiment, no more than 10% of your reaction should consist of the undiluted RT product.

1. Label a separate microcentrifuge tube for each diluted sample:
   - Liver
   - Heart
   - Brain
   - Lung
2. Add the required volume of water (diluent) to each empty tube:

Tube	Sample name	Diluent volume (µL)
1	Liver	19
2	Heart	19
3	Brain	19
4	Lung	19

3. Add the required volume of cDNA sample stock (100 ng/µL) to each empty tube:

Tube	Sample name	Volume (µL)
1	Liver	1.0
2	Heart	1.0
3	Brain	1.0
4	Lung	1.0

4. Vortex each diluted sample for 3 to 5 seconds, then centrifuge the tubes briefly.
5. Place the diluted samples on ice until you prepare the reaction plate.

Prepare the reaction mix ("cocktail mix")

1. Label an appropriately sized tube for each reaction mix:
   - ACTB Reaction Mix
   - GAPDH Reaction Mix
   - GH1 Reaction Mix

- LPIN1 Reaction Mix
- LIPC Reaction Mix

2. For the ACTB assay, add the required volumes of each component to the ACTB Reaction Mix tube:

Component	Volume (µL) for 1 reaction	Volume (µL) for 13 reactions (plus 10% excess)
TaqMan® Fast Universal PCR Master Mix (2X)	5.0	75.0
ACTB Assay Mix (20X)	0.5	7.5
Water	3.5	52.5
Total Reaction Mix Volume	9.0	135.0

3. Mix the reaction mix in each tube by gently pipetting up and down, then cap each tube.
4. Centrifuge the tubes briefly to remove air bubbles.
5. Place the reaction mixes on ice until you prepare the reaction plate.
6. Repeat steps 2 through 5 for the GAPDH, GH1, LPIN1, and LIPC assays.

   Note: Do not add the sample at this time.

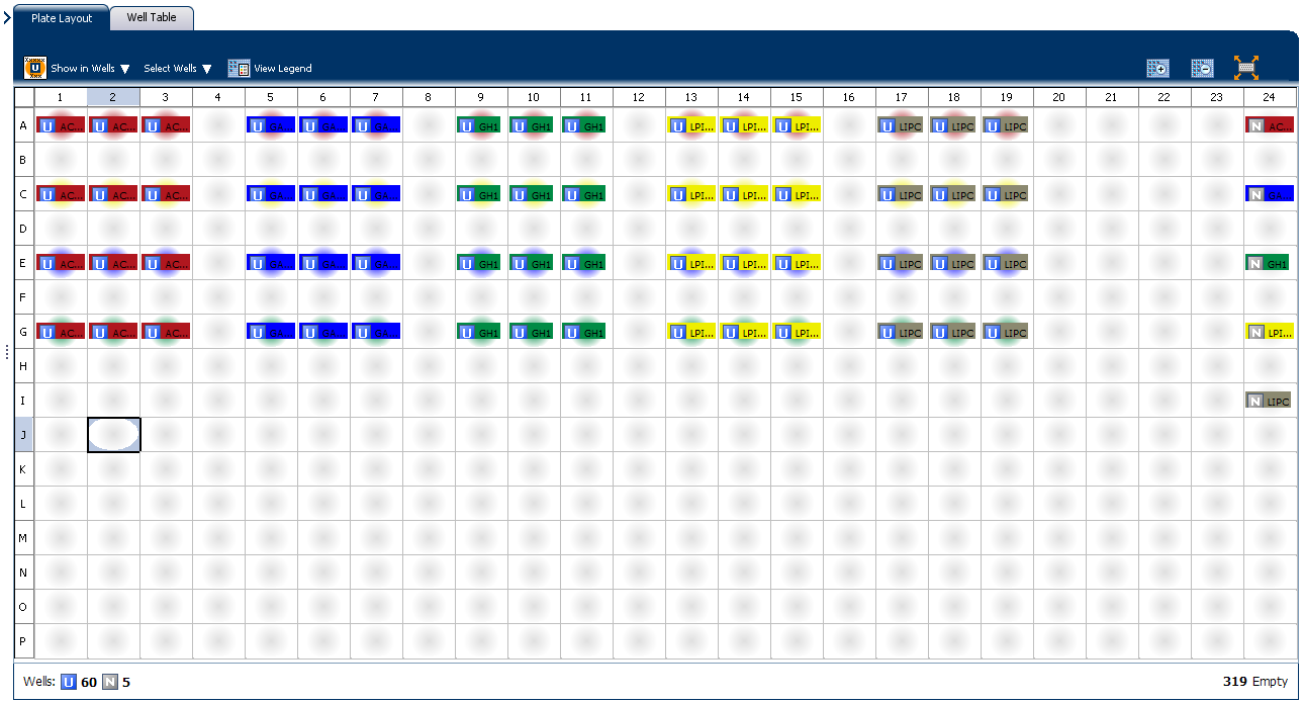
Prepare the reaction plate

Example experiment reaction plate components

The reaction plate for the Comparative C_T example experiment contains:

- A MicroAmp® Optical 384-Well Reaction Plate
- Reaction volume: 10µL/well
- The reaction plate contains:
  - 60 Unknown wells 
  - 5 Negative control wells 
  - 319 Empty wells

The following is an image of the plate layout for the example experiment:



To prepare the reaction plate components

1. Add 1 µL of each cDNA to the appropriate wells.
2. Pipet 1 µL of sterile water into the NTC wells.
3. Add 8 µL of the appropriate assay-specific cocktail to the wells.
4. Seal the reaction plate with optical adhesive film.
5. Centrifuge the reaction plate briefly to remove air bubbles.
6. Confirm that the liquid is at the bottom of each well of the reaction plate. If not, centrifuge the reaction plate again at a higher speed and for a longer period of time.
7. Until you are ready to perform the run, place the reaction plate at 4°C, in the dark.

Tips for preparing reactions for your own experiments

Tips for preparing templates

When you prepare your own Comparative C_T experiment, Life Technologies recommends the following templates:

- **Complementary DNA (cDNA)** – cDNA reverse-transcribed from total RNA samples.
- **Genomic DNA (gDNA)** – Purified gDNA already extracted from tissue or sample.

Tips for preparing the reaction mix

If your experiment includes more than one target assay, prepare the reaction mix for each target assay separately.

Tips for preparing the reaction plate

When you prepare your own Comparative C_T experiment, make sure the arrangement of the PCR reactions matches the plate layout displayed in the QuantStudio™ 6 and 7 Flex Real-Time PCR System Software.

For more information

For more information on...	Refer to...	Publication number
Assigning the reaction plate components	Chapter 2 in Booklet 1, *Getting Started with QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Experiments*	4489822
Sealing the reaction plate	Chapter 2 in Booklet 1, *Getting Started with QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Experiments*	4489822

10 Run the Experiment

This chapter explains how to run the example experiment on the QuantStudio™ 6 or 7 Instrument.

This chapter covers:

IMPORTANT! Run the experiment at the same ambient temperature at which you calibrated the instrument. Extreme variations in ambient temperature can affect the heating and cooling of the instrument and influence experimental results.

IMPORTANT! Do not attempt to open the access door during the run. The door is locked while the QuantStudio™ 6 or 7 Instrument is in operation.

Start the run

1. Open the Comparative C_T example file that you created using instructions in Chapter 8.
2. Load the reaction plate into the instrument.
3. Start the run.

Monitor the run

Monitor the example experiment run:

- From the QuantStudio™ 6 and 7 Flex Real-Time PCR System Software using the Run screen, while the experiment is in progress.
- From the Instrument Console of the QuantStudio™ 6 and 7 Flex Real-Time PCR System Software (to monitor an experiment started from another computer or from the QuantStudio™ 6 or 7 Instrument touchscreen).
- From the QuantStudio™ 6 or 7 Instrument touchscreen.

From the Instrument Console of the QuantStudio™ 6 and 7 Flex Real-Time PCR System Software

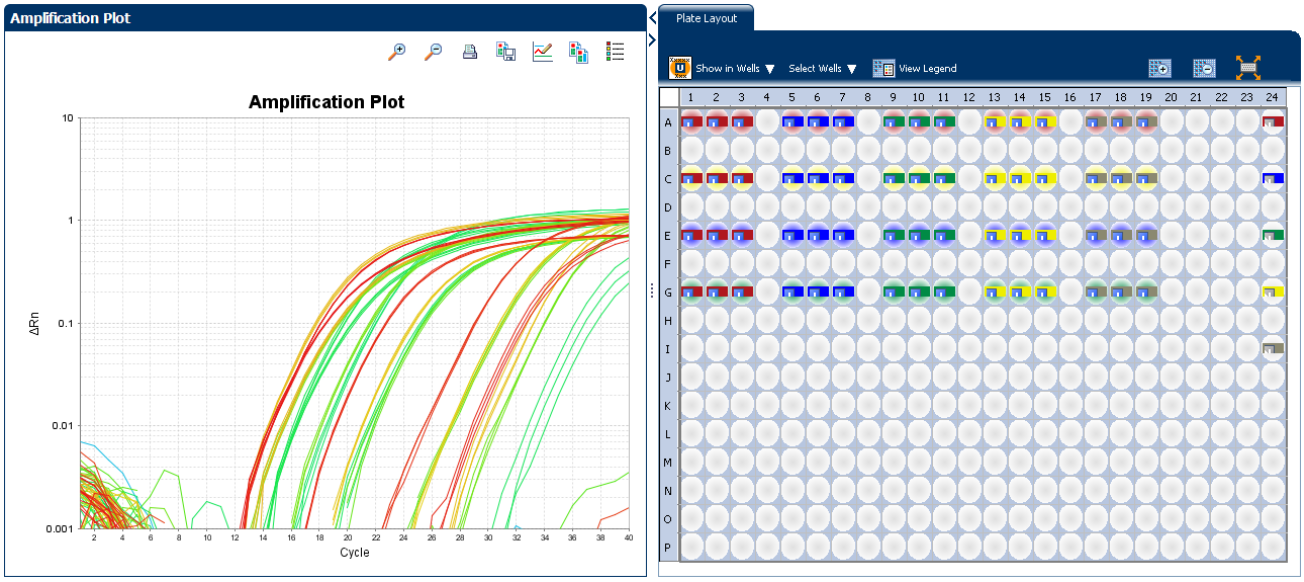
1. In the Instrument Console screen, select the instrument icon.
2. Click **Manage Instrument** or double-click on the instrument icon.
3. In the Manage Instrument screen, click **Monitor Running Experiment** to access the Run screen.

View the Amplification Plot

You can view the progress of the run in real time. During the run, periodically view all the three plots available from the QuantStudio™ 6 and 7 Flex Software for potential problems.

Click **Amplification Plot** from the Run Experiment Menu, select the Plate Layout tab, then select the wells t to view.

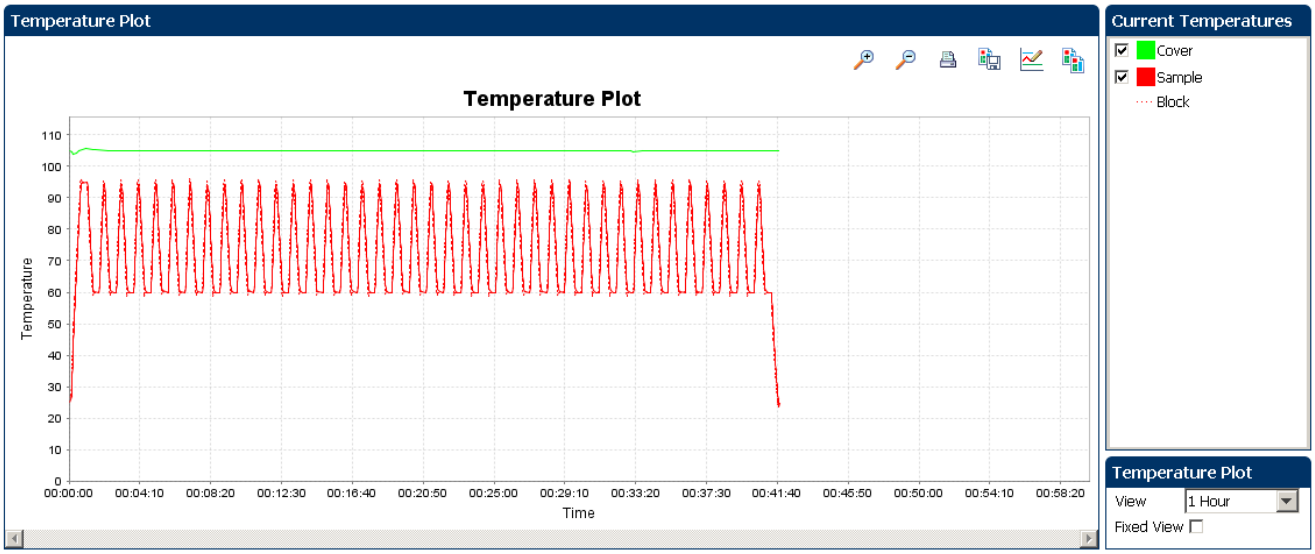
The following is an image of the Amplification Plot screen as it appears at the end of the example experiment.



View the Temperature Plot

Click **Temperature Plot** from the Run Experiment Menu.

The following is an image of the Temperature Plot screen as it appears during the example experiment.

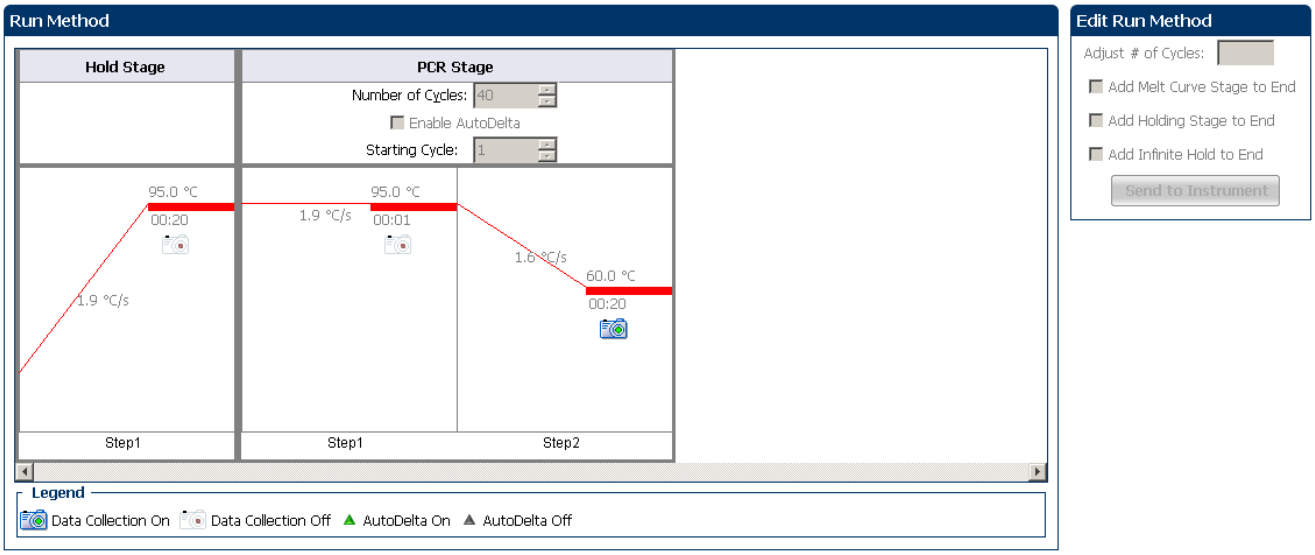


Note: The Sample temperature displayed in the Current Temperatures group is an estimated value.

View the Run Method

Click **Run Method** from the Run Experiment Menu.

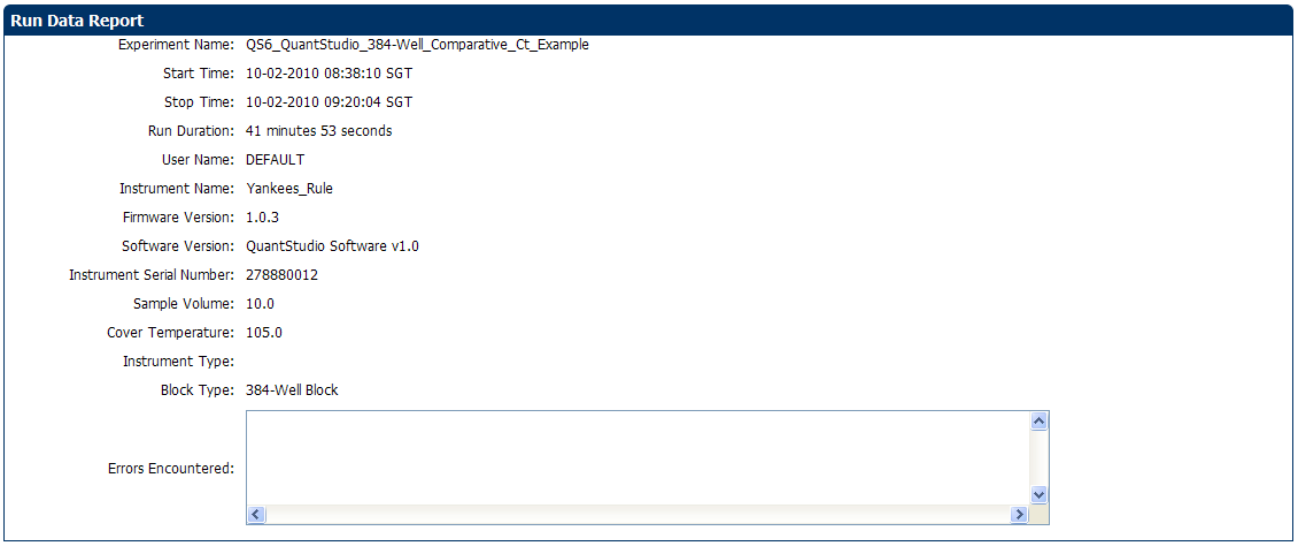
The following is an image of the Run Method screen as it appears in the example experiment.



View Run Data

Click **View Run Data** from the Run Experiment Menu.

The following is an image of the View Run Data screen as it appears in the example experiment:

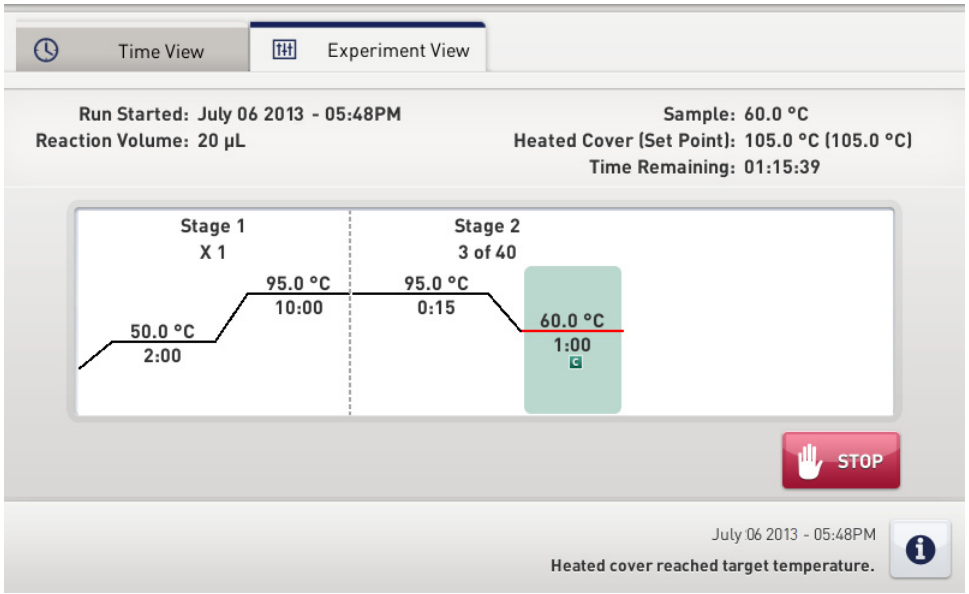


From the QuantStudio™ 6 or 7 Instrument touchscreen

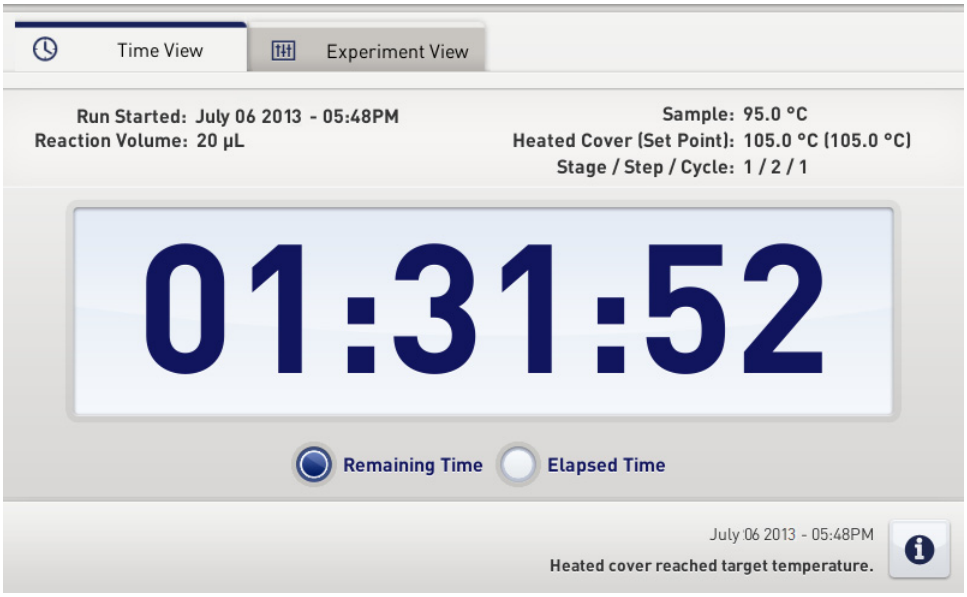
You can also view the progress of the run from the QuantStudio™ 6 or 7 Instrument touchscreen.

The following are images of the Run Method screen on the QuantStudio™ 6 or 7 Instrument touchscreen:

Experiment view



Time View


Time View
Experiment View
Run Started: July 06 2013 - 05:48PM
Reaction Volume: 20 µL
Sample: 95.0 °C
Heated Cover (Set Point): 105.0 °C (105.0 °C)
Stage / Step / Cycle: 1 / 2 / 1
01:31:52
Remaining Time
Elapsed Time
July 06 2013 - 05:48PM
Heated cover reached target temperature.

11 Review Results and Adjust Experiment Parameters

In Section 11.1 of this chapter you review the analyzed data using several of the analysis screens and publish the data. Section 11.2 of this chapter walks you through the process of modifying experiment parameters to troubleshoot problems with experiment results before rerunning an experiment.

This chapter covers:

Section 11.1 Review Results

Analyze the example experiment

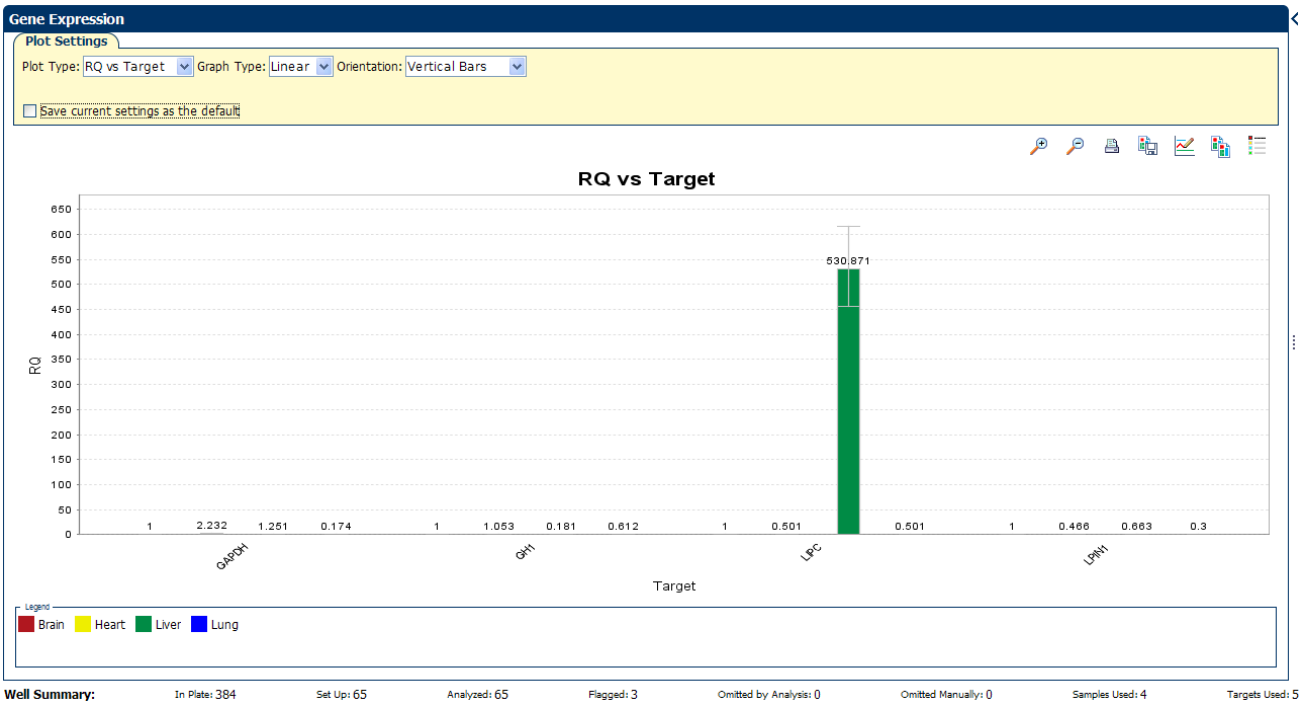
1. Open the example experiment file that you ran in Chapter 10.
2. Click **Analyze**. The software analyzes the data using the default analysis settings.

   Note: You can also access the experiment to analyze from the Home screen.

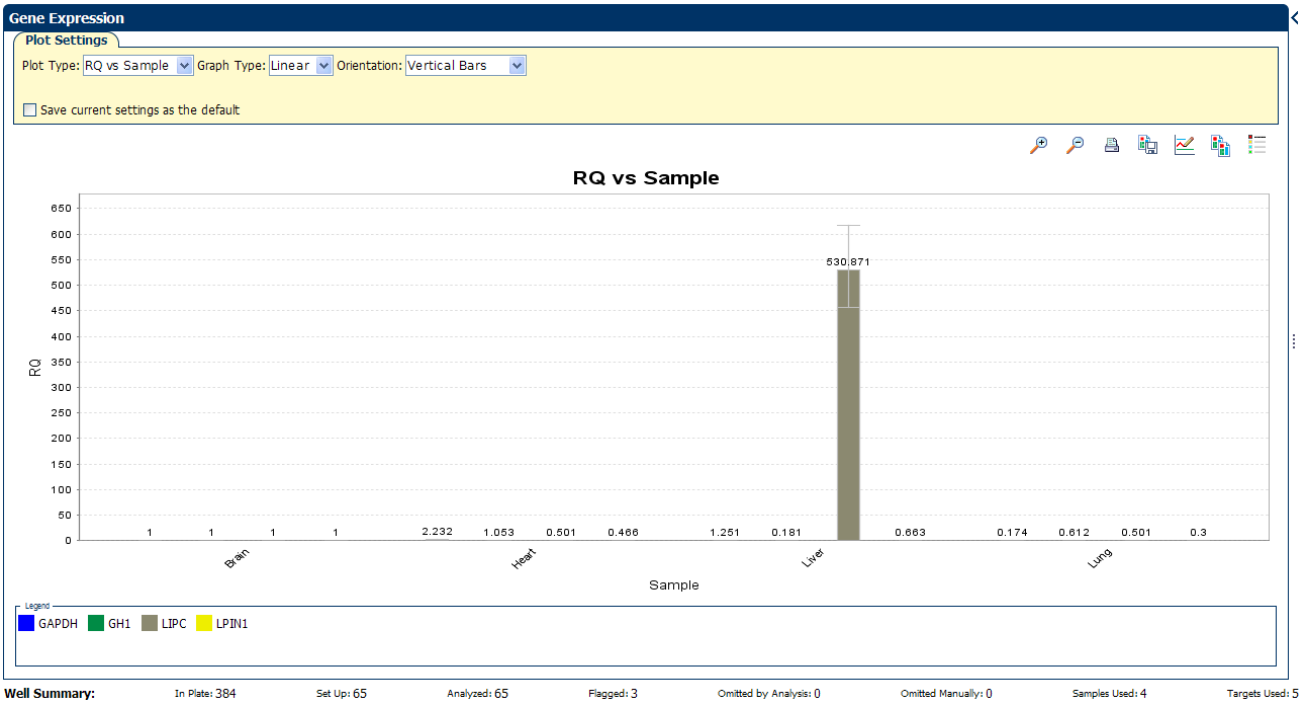
Assess the gene expression profile using the Gene Expression Plot

The Gene Expression Plot screen displays the results of relative quantification calculations in the gene expression profile. There are two plots available:

- **RQ vs Target** – Groups the relative quantification (RQ) values by target. Each sample is plotted for each target. You can view the plot as the linear, log10, Ln, and log2 graph types. The Gene Expression plot when viewed as a linear graph type looks like this:



- **RQ vs Sample** – Groups the relative quantification (RQ) values by sample. Each target is plotted for each sample. You can view the plot as the following graph types: linear, log10, Ln, log2.



Example experiment values

Review each target in the Gene Expression Plot screen for the expression level (or fold change) of the target sample relative to the reference sample.

View the Gene Expression Plot

1. From the Experiment Menu pane, select **Analysis** ▸ **Gene Expression**.

   Note: If no data are displayed, click **Analyze**.

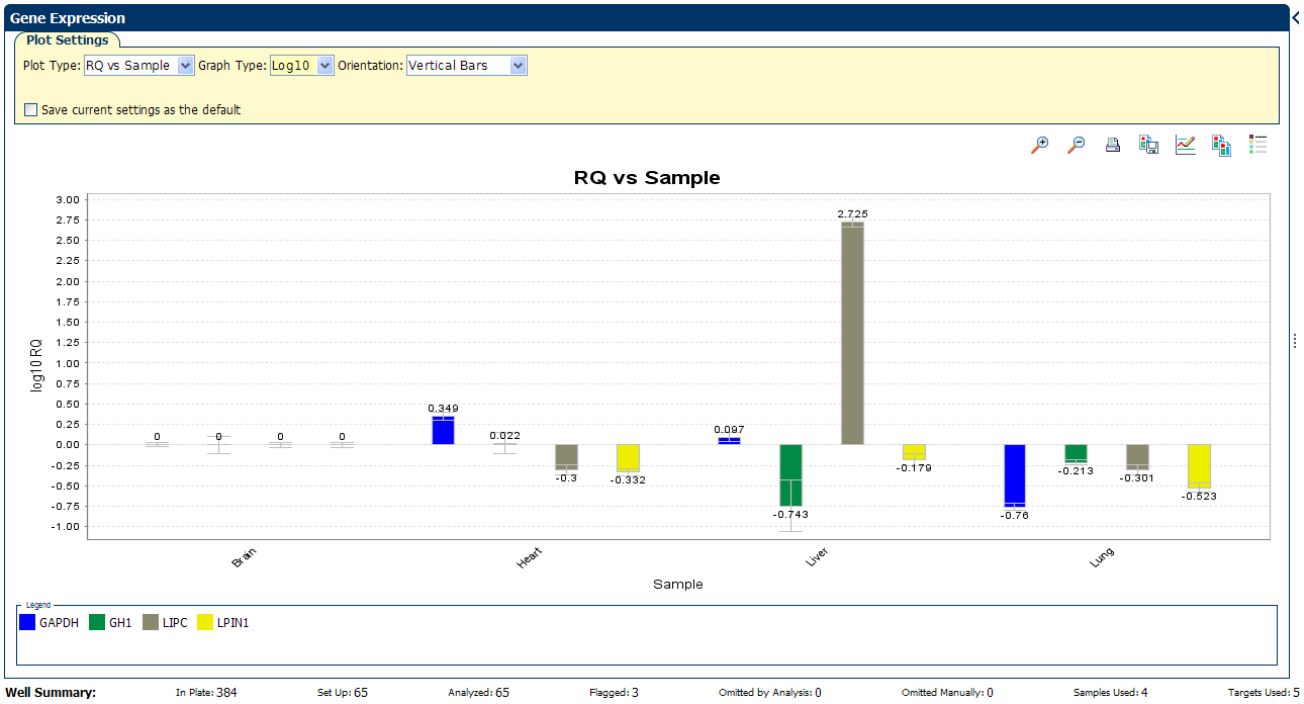
2. In the Gene Expression Plot screen, select:

Menu	Selection
Plot Type	RQ vs Sample (default)
Graph Type	Log10
Orientation	Vertical Bars

3. Click **Show a legend for the plot** (default).

   Note: This is a toggle button. When the legend is displayed, the button changes to Hide the plot legend.

In the example experiment, as shown below, the expression level of each target gene in liver, heart, and lung is displayed relative to its respective expression level in the reference sample (brain). Since the reference sample is compared to itself, the relative expression level is 1. When the result is displayed in the Log10 graph type, the expression level of the reference sample appears as 0 in the graph (log10 of 1 = 0).



Assessing the gene expression plot your own experiments

When you analyze your own Comparative C_T experiment, look for differences in gene expression (as a fold change) relative to the reference sample.

Identify well problems using the Well Table

The Well Table displays data for each well in the reaction plate, including:

- The sample name, target name, task, and dyes
- The calculated threshold cycle (C_T), normalized fluorescence (Rn), and quantity values
- Flags

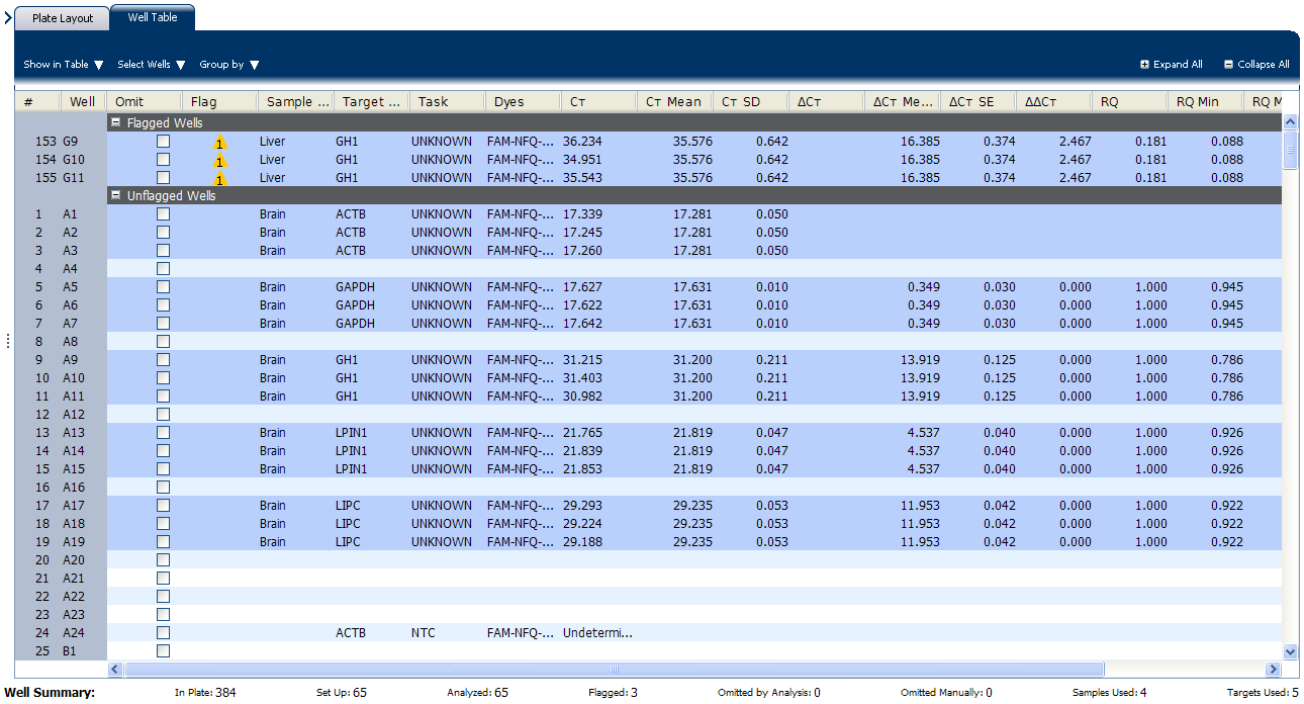
Example experiment values and flags

Review the Well Table to evaluate the C_T precision of the replicate groups.

View the well table

1. From the Experiment Menu pane, select **Analysis ▸ Amplification Plot**, then click the **Well Table** tab.

2. From the Group By drop-down menu, select **Replicate**.
3. Look at the C_T SD column to evaluate the C_T precision of the replicate groups. In the example experiment, there are three outliers. You will omit these wells in the troubleshooting section ("Improve CT precision by omitting wells" on page 120).



#	Well	Omit	Flag	Sample ...	Target ...	Task	Dyes	CT	CT Mean	CT SD	ΔCT	ΔCT Me...	ΔCT SE	ΔΔCT	RQ	RQ Min	RQ M
Flagged Wells																	
153	G9	☐	1	Liver	GH1	UNKNOWN	FAM-NFQ-...	36.234	35.576	0.642		16.385	0.374	2.467	0.181	0.088	
154	G10	☐	1	Liver	GH1	UNKNOWN	FAM-NFQ-...	34.951	35.576	0.642		16.385	0.374	2.467	0.181	0.088	
155	G11	☐	1	Liver	GH1	UNKNOWN	FAM-NFQ-...	35.543	35.576	0.642		16.385	0.374	2.467	0.181	0.088	
Unflagged Wells																	
1	A1	☐		Brain	ACTB	UNKNOWN	FAM-NFQ-...	17.339	17.281	0.050							
2	A2	☐		Brain	ACTB	UNKNOWN	FAM-NFQ-...	17.245	17.281	0.050							
3	A3	☐		Brain	ACTB	UNKNOWN	FAM-NFQ-...	17.260	17.281	0.050							
4	A4	☐															
5	A5	☐		Brain	GAPDH	UNKNOWN	FAM-NFQ-...	17.627	17.631	0.010		0.349	0.030	0.000	1.000	0.945	
6	A6	☐		Brain	GAPDH	UNKNOWN	FAM-NFQ-...	17.622	17.631	0.010		0.349	0.030	0.000	1.000	0.945	
7	A7	☐		Brain	GAPDH	UNKNOWN	FAM-NFQ-...	17.642	17.631	0.010		0.349	0.030	0.000	1.000	0.945	
8	A8	☐															
9	A9	☐		Brain	GH1	UNKNOWN	FAM-NFQ-...	31.215	31.200	0.211		13.919	0.125	0.000	1.000	0.786	
10	A10	☐		Brain	GH1	UNKNOWN	FAM-NFQ-...	31.403	31.200	0.211		13.919	0.125	0.000	1.000	0.786	
11	A11	☐		Brain	GH1	UNKNOWN	FAM-NFQ-...	30.982	31.200	0.211		13.919	0.125	0.000	1.000	0.786	
12	A12	☐															
13	A13	☐		Brain	LPIN1	UNKNOWN	FAM-NFQ-...	21.765	21.819	0.047		4.537	0.040	0.000	1.000	0.926	
14	A14	☐		Brain	LPIN1	UNKNOWN	FAM-NFQ-...	21.839	21.819	0.047		4.537	0.040	0.000	1.000	0.926	
15	A15	☐		Brain	LPIN1	UNKNOWN	FAM-NFQ-...	21.853	21.819	0.047		4.537	0.040	0.000	1.000	0.926	
16	A16	☐															
17	A17	☐		Brain	LIPC	UNKNOWN	FAM-NFQ-...	29.293	29.235	0.053		11.953	0.042	0.000	1.000	0.922	
18	A18	☐		Brain	LIPC	UNKNOWN	FAM-NFQ-...	29.224	29.235	0.053		11.953	0.042	0.000	1.000	0.922	
19	A19	☐		Brain	LIPC	UNKNOWN	FAM-NFQ-...	29.188	29.235	0.053		11.953	0.042	0.000	1.000	0.922	
20	A20	☐															
21	A21	☐															
22	A22	☐															
23	A23	☐															
24	A24	☐			ACTB	NTC	FAM-NFQ-...	Undetermi...									
25	B1	☐															

Note: To show/hide columns in the Well Table, select/deselect the column name from the Show in Table drop-down menu.

Assessing the well table in your own experiments

When you analyze your own Comparative C_T experiment, look for standard deviation in the replicate groups (C_T SD values). If needed, omit outliers (see "Improve CT precision by omitting wells" on page 120).

Assess amplification results using the Amplification Plot

Amplification plots available for viewing

The Amplification Plot screen displays amplification of all samples in the selected wells. There are three plots available:

- **ΔRn vs Cycle** – ΔRn is the magnitude of normalized fluorescence signal generated by the reporter at each cycle during the PCR amplification. This plot displays ΔRn as a function of cycle number. You can use this plot to identify and examine irregular amplification and to view threshold and baseline values for the run.
- **Rn vs Cycle** – Rn is the fluorescence signal from the reporter dye normalized to the fluorescence signal from the passive reference. This plot displays Rn as a function of cycle number. You can use this plot to identify and examine irregular amplification.
- **C_T vs Well** – C_T is the PCR cycle number at which the fluorescence meets the threshold in the amplification plot. This plot displays C_T as a function of well position. You can use this plot to locate outlying amplification (outliers).

Each plot can be viewed as a linear or log10 graph type.

Purpose

The purpose of viewing the amplification plot for the example experiment is to identify:

- Correct baseline and threshold values
- Outliers

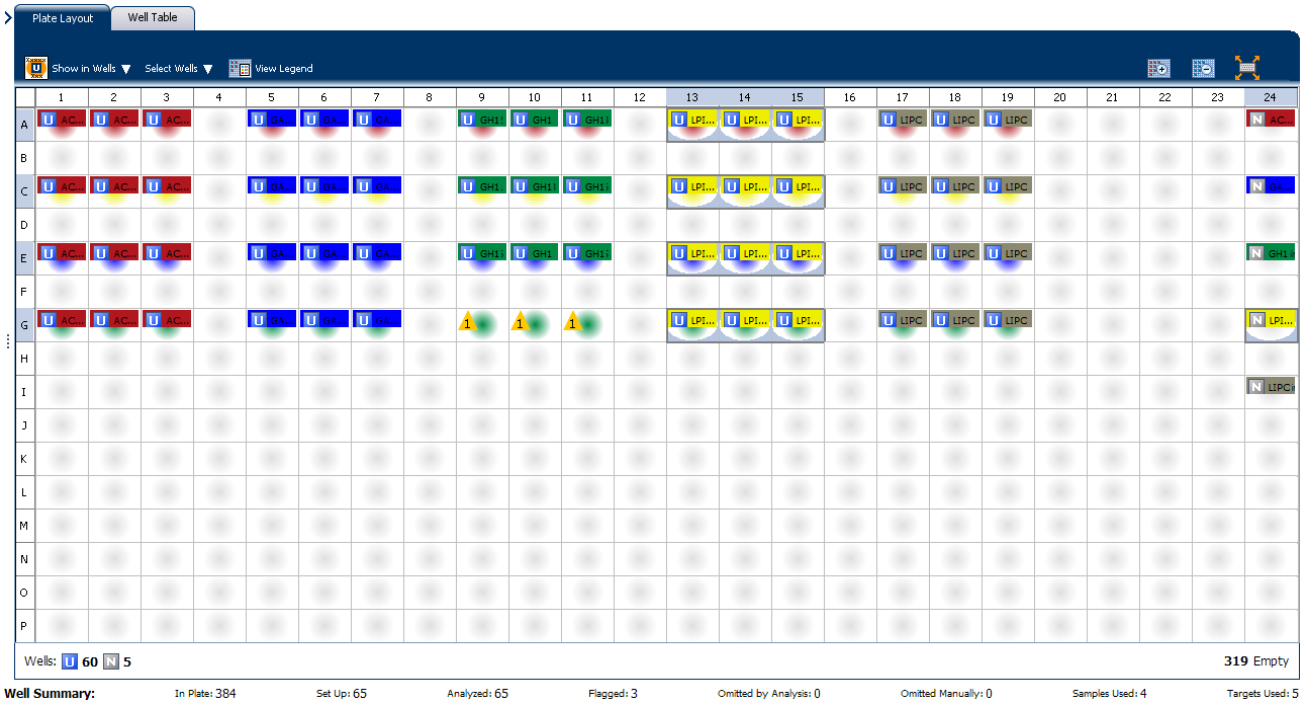
View the Amplification Plot

1. From the Experiment Menu pane, select **Analysis ▸ Amplification Plot**.

   Note: If no data are displayed, click **Analyze**.

2. Display the LP1N1 wells in the Amplification Plot screen:
   a. Click the **Plate Layout** tab.
   b. From the Select Wells drop-down menus, select **Target**, then **LP1N1**.

The following is an image of the Plate Layout screen:



3. In the Amplification Plot screen, enter:

Menu	Selection
Plot Type	ΔRn vs Cycle (default)
Plot Color	Well (default)
 (This is a toggle button. When the legend is displayed, the button changes to **Hide the plot legend**.)	Check (default)

4. View the baseline values:
   a. From the Graph Type drop-down menu, select **Linear**.
   b. Select the **Baseline** check box to show the start cycle and end cycle.

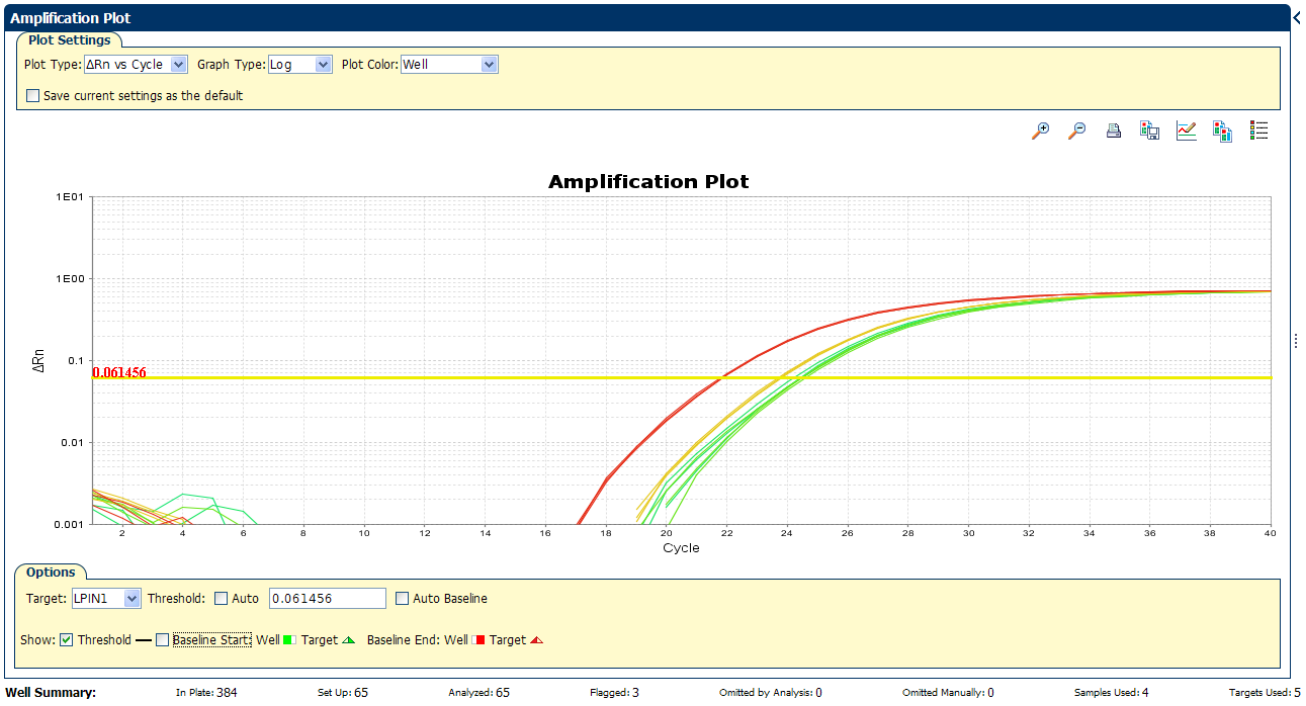
c. Verify that the baseline is set correctly: The end cycle should be set a few cycles before the cycle number where significant fluorescent signal is detected. In the example experiment, the baseline is set correctly.

5. View the threshold values:

Menu	Select
Graph Type	Log
Target	LP1N1

a. Select the **Threshold** check box to show the threshold.

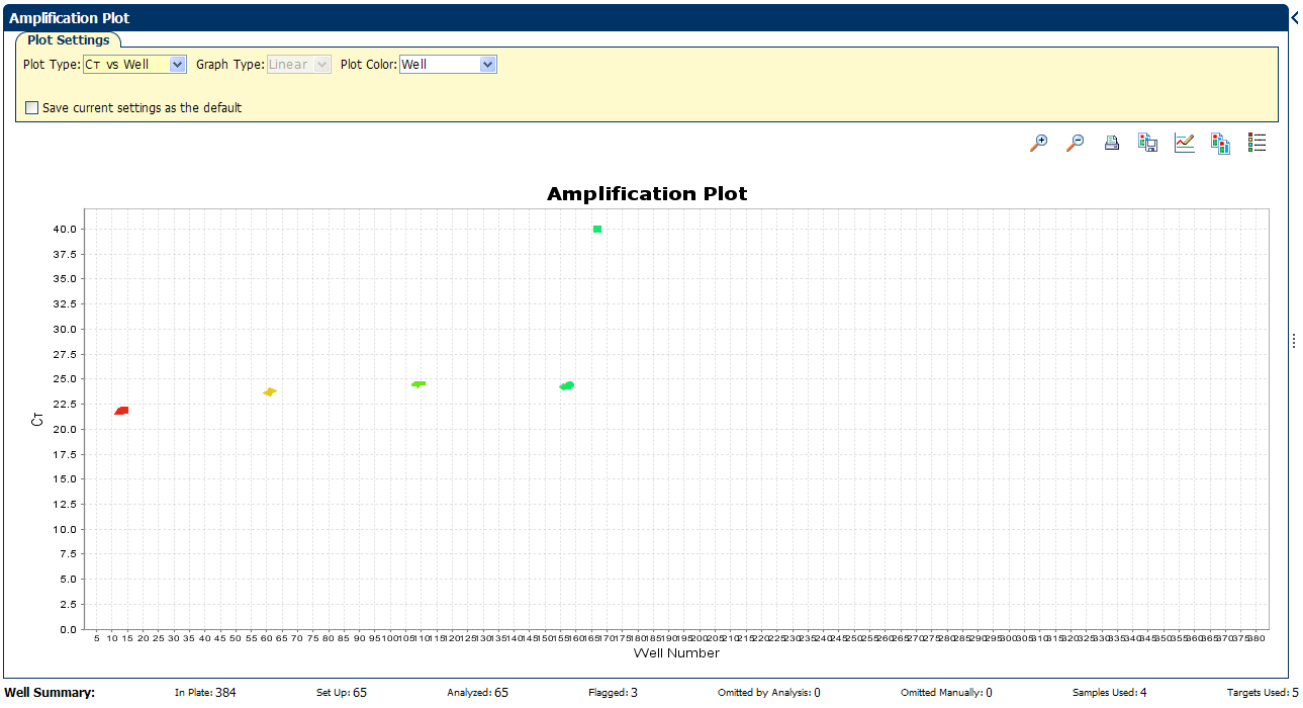
b. Verify that the threshold is set correctly. In the example experiment, the threshold is in the exponential phase.



6. Locate any outliers:

   a. From the Plot Type drop-down menu, select **C_T vs Well**.

   b. Look for outliers from the amplification plot. In the example experiment, there are no outliers for LP1N1.



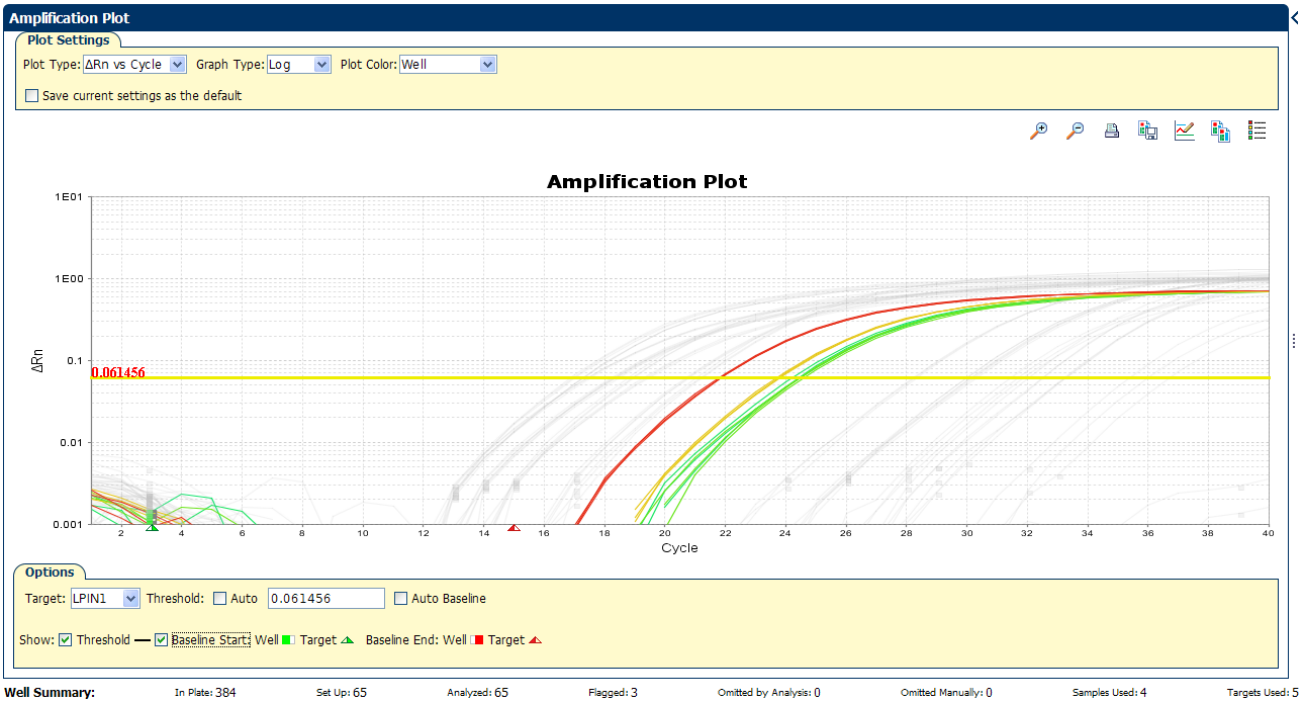
7. Repeat steps 2 through 6 for the GH1, TGFB1, LIPC, GAPDH, 18S, and ACTB wells. In the example experiment, there is one outlier for 18S. You will omit these wells in the troubleshooting section ("Improve CT precision by omitting wells" on page 120).

Tips for analyzing your own experiments

When you analyze your own Comparative C_T experiment, look for:

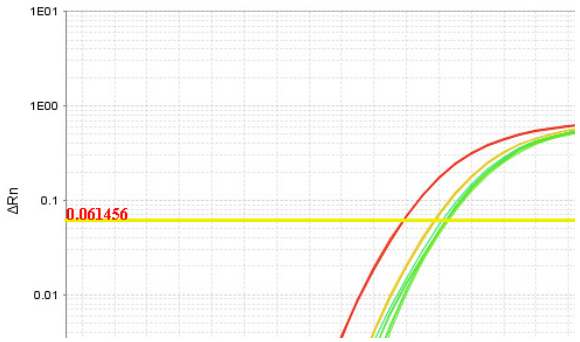
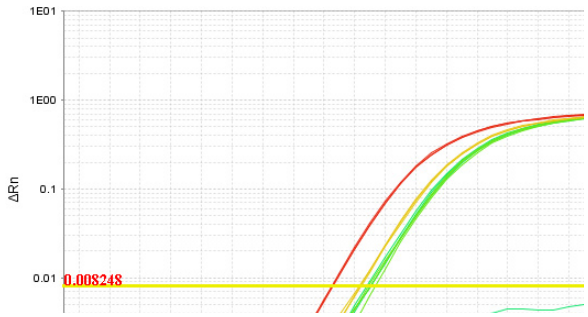
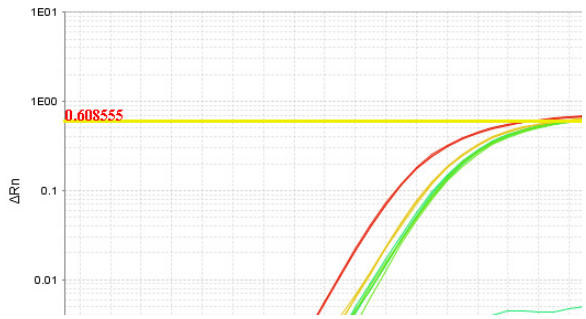
- **Outliers**
- **A typical amplification plot** – The QuantStudio™ 6 and 7 Flex Software automatically calculates baseline and threshold values based on the assumption that the data exhibit a *typical* amplification plot. A typical amplification plot has four distinct sections:
  - Plateau phase
  - Linear phase
  - Exponential (geometric phase)
  - Baseline

  A typical amplification plot should look like this:

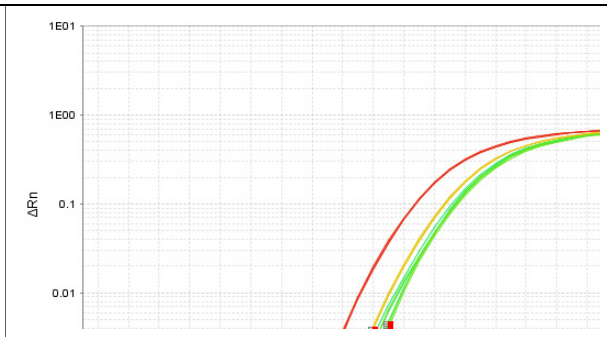
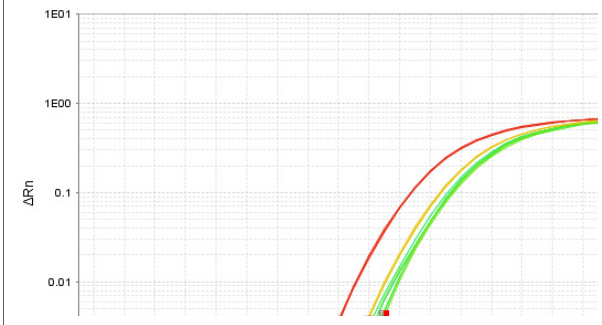
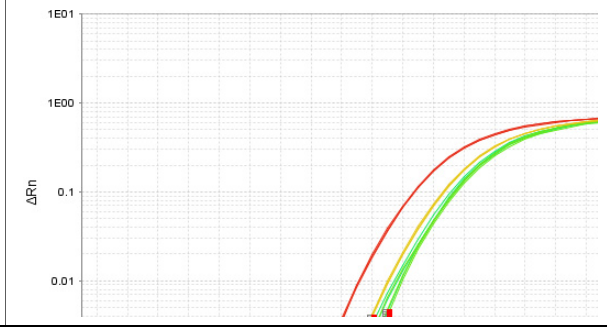


IMPORTANT! Experimental error (such as contamination or pipetting errors) can produce atypical amplification curves that can result in incorrect baseline and threshold value calculations by the QuantStudio™ 6 and 7 Flex Software. Therefore, Life Technologies recommends that you examine the Amplification Plot screen and review the assigned baseline and threshold values for each well after analysis completes.

- **Correct threshold values**

Threshold Set Correctly The threshold is set in the exponential phase of the amplification curve. Threshold settings above or below the optimum increase the standard deviation of the replicate groups.	
Threshold Set Too Low The threshold is set below the exponential phase of the amplification curve. The standard deviation is significantly higher than that for a plot where the threshold is set correctly. Drag the threshold bar up into the exponential phase of the curve.	
Threshold Set Too High The threshold is set above the exponential phase of the amplification curve. The standard deviation is significantly higher than that for a plot where the threshold is set correctly. Drag the threshold bar down into the exponential phase of the curve.	

- **Correct baseline values**

Baseline Set Correctly The amplification curve begins after the maximum baseline.	
Baseline Set Too Low The amplification curve begins too far to the right of the maximum baseline. Increase the End Cycle value.	
Baseline Set Too High The amplification curve begins before the maximum baseline. Decrease the End Cycle value.	

If your experiment does not meet the guidelines above, troubleshoot as follows:

- Omit wells (see "Improve CT precision by omitting wells" on page 120).

  Or
- Manually adjust the baseline and/or threshold (see "Adjust analysis settings" on page 117).

Confirm accurate dye signal using the Multicomponent Plot

The Multicomponent Plot screen displays the complete spectral contribution of each dye in a selected well over the duration of the PCR run.

Purpose

In the Comparative C_T example experiment, you review the Multicomponent Plot screen for:

- ROX™ dye (passive reference)
- FAM™ dye (reporter)
- Spikes, dips, and/or sudden changes
- Amplification in the negative control wells

View the Multicomponent Plot

1. From the Experiment Menu pane, select **Analysis** ▸ **Multicomponent Plot**.

   Note: If no data are displayed, click **Analyze**.

2. Display the unknown and standard wells one at a time in the Multicomponent Plot screen:

   a. Click the **Plate Layout** tab.

   b. Select one well in the plate layout; the well is shown in the Multicomponent Plot screen.

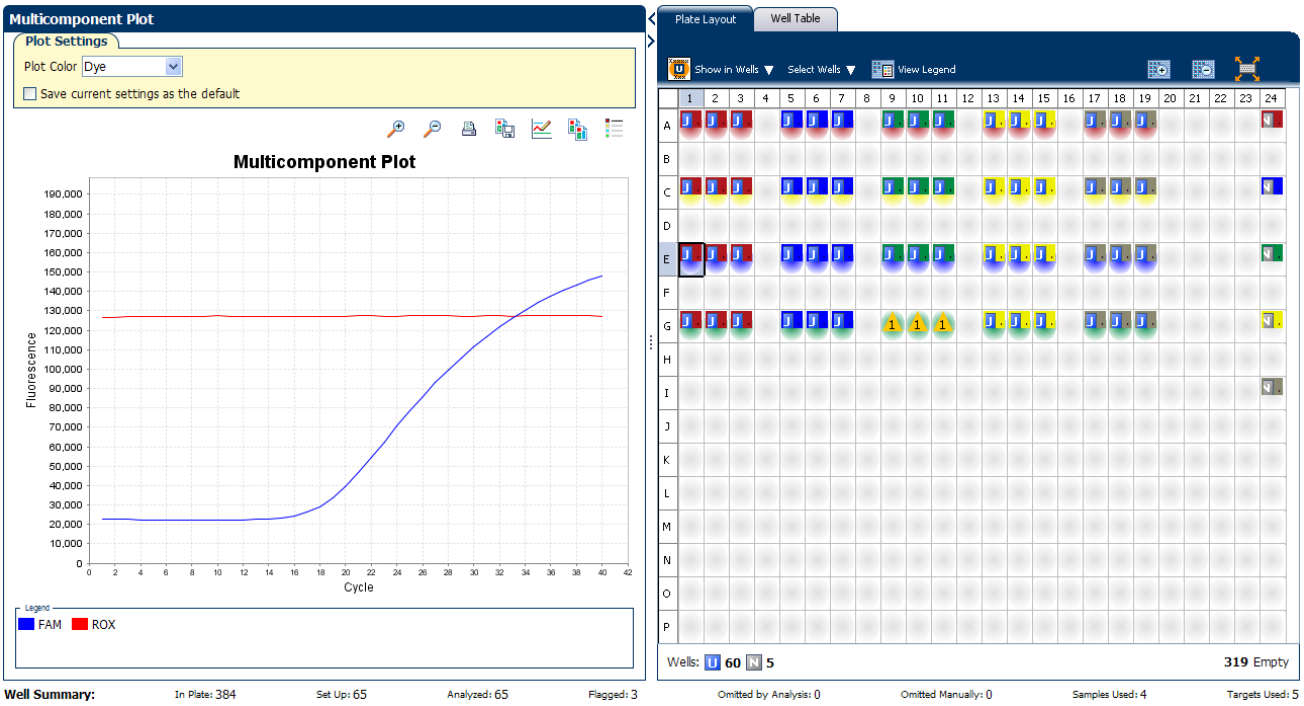
      Note: If you select multiple wells, the Multicomponent Plot screen displays the data for all selected wells simultaneously.

3. From the Plot Color drop-down menu, select **Dye**.

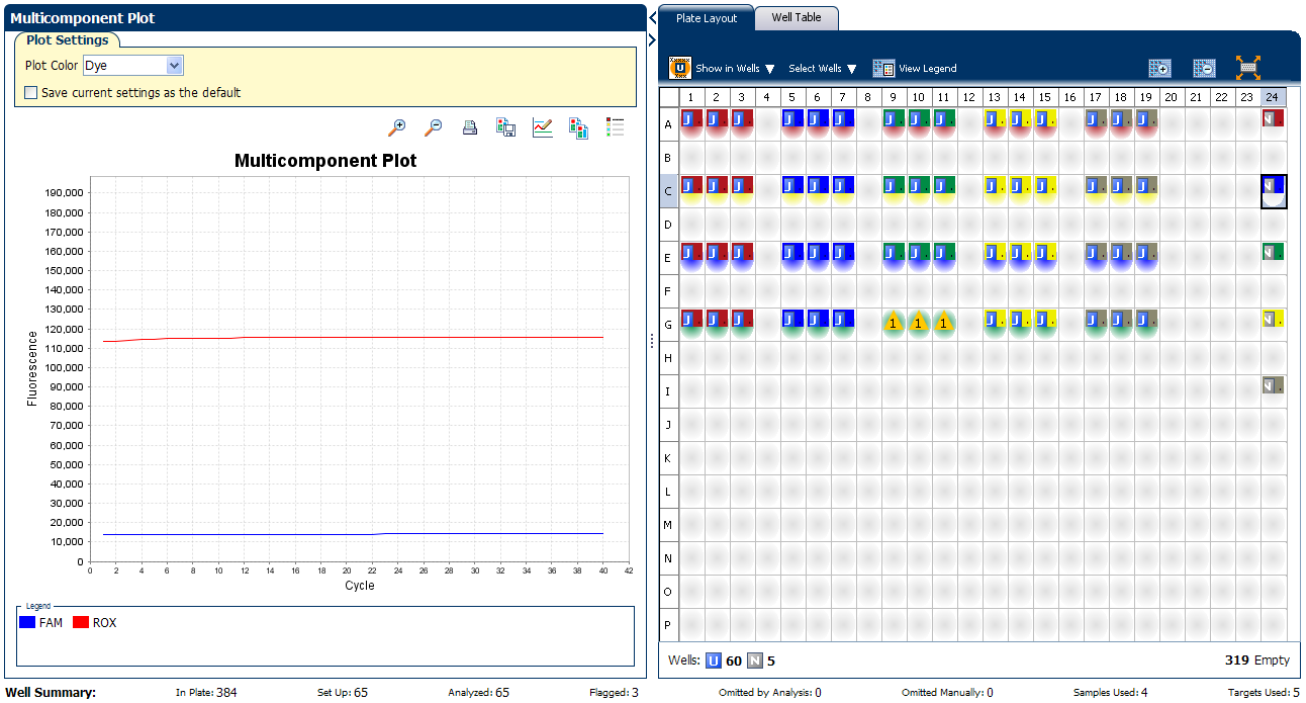
4. Click  **Show a legend for the plot** (default).

   Note: This is a toggle button. When the legend is displayed, the button changes to Hide the plot legend.

5. Check the FAM dye signals. In the example experiment, the FAM dye signal increases throughout the PCR process, indicating normal amplification.



6. Select the negative control wells one at time and check for amplification. In the example experiment, there is no amplification in the negative control wells.



Tips for confirming dye accuracy in your own experiment

When you analyze your own Comparative C_T experiment, look for:

- **Passive reference** – The passive reference dye fluorescence level should remain relatively constant throughout the PCR process.
- **Reporter dye** – The reporter dye fluorescence level should display a flat region corresponding to the baseline, followed by a rapid rise in fluorescence as the amplification proceeds.
- **Irregularities in the signal** – There should not be any spikes, dips, and/or sudden changes in the fluorescent signal.
- **Negative control wells** – There should not be any amplification in the negative control wells.

Determine signal accuracy using the Raw Data Plot

The Raw Data Plot screen displays the raw fluorescence signal (not normalized) for each optical filter for the selected wells during each cycle of the real-time PCR.

About the example experiment

In the Comparative C_T example experiment, you review the Raw Data Plot screen for a stable increase in signal (no abrupt changes or dips) from the appropriate filter.

View the Raw Data Plot

1. From the Experiment Menu pane, select **Analysis** ▸ **Raw Data Plot**.

   Note: If no data are displayed, click **Analyze**.

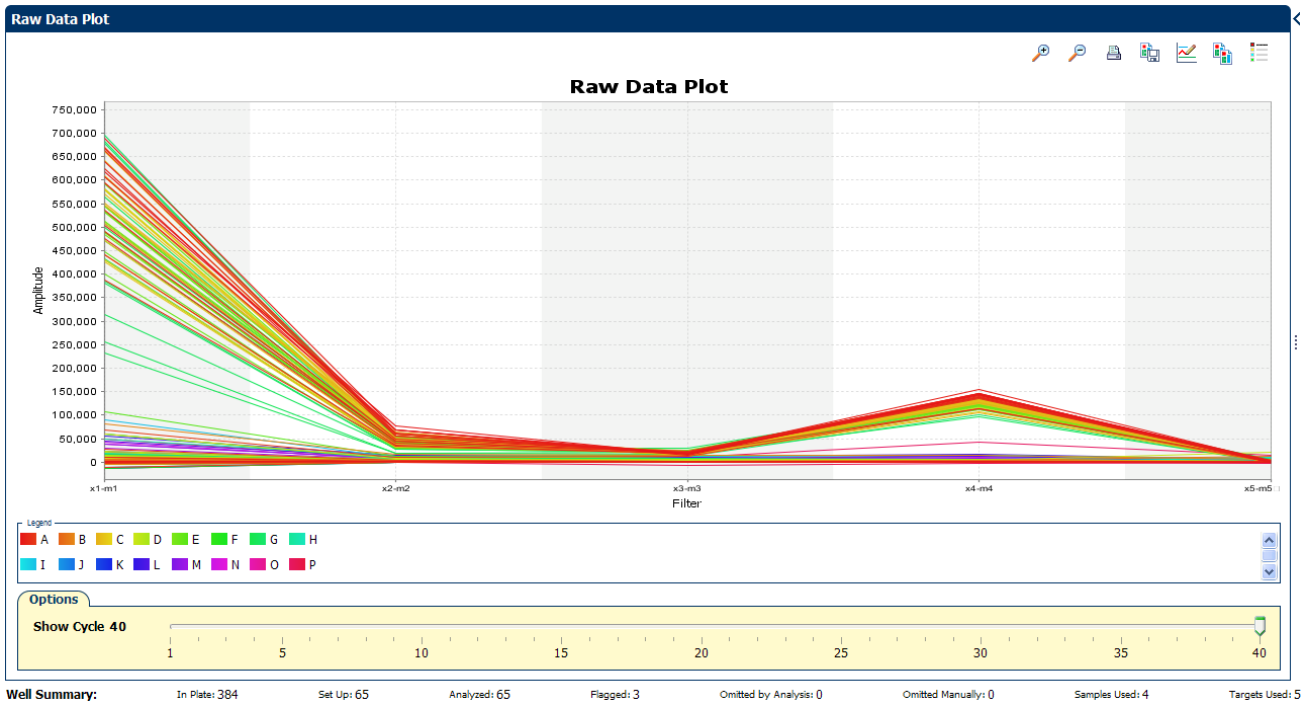
2. Display all 384 wells in the Raw Data Plot screen by clicking the upper left corner of the plate layout in the Plate Layout tab.

3. Click  **Show a legend for the plot** (default).

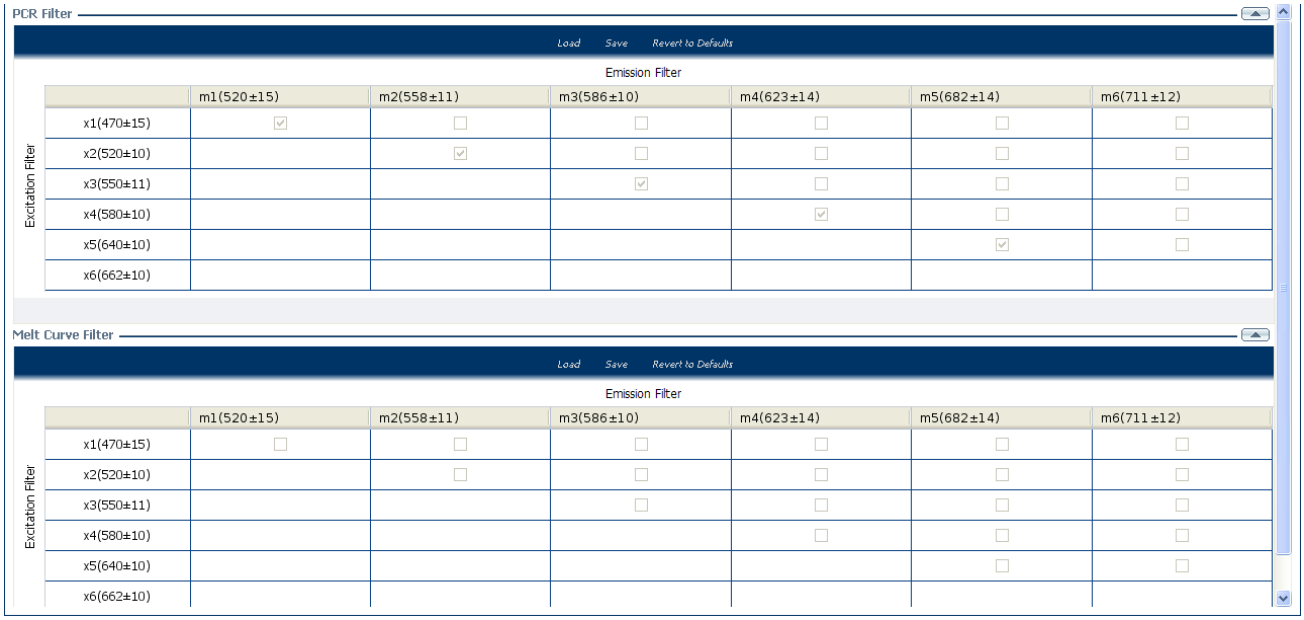
   Note: This is a toggle button. When the legend is displayed, the button changes to Hide the plot legend.

   Note: The legend displays the color code for each row of the reaction plate (see the legend in the Raw Data Plot shown below).

4. Click and drag the Show Cycle pointer from cycle 1 to cycle 40. In the example experiment, there is a stable increase in signal from filter 1, which corresponds to the FAM™ dye filter.



The filters used for the example experiment are:



Tips for determining signal accuracy in your own experiment

When you analyze your own Comparative C_T experiment, look for the following in each filter:

- Characteristic signal growth
- No abrupt changes or dips

View the endogenous control profile using the QC Plot

In the Comparative C_T experiment, the QC Plot screen displays the Endogenous Control Profile plot for all the targets in the experiment. The QC Plot serves as a tool to help you choose the best endogenous control for that experiment. The endogenous control profile plot is a visual display of the C_T level of the endogenous control across the sample. You can view up to four endogenous controls at a time. The sample is plotted on the X-axis, and C_T is plotted on the Y-axis. The expression is viewed as a color and shape combination in the plot. Endogenous controls are also known as reference genes.

Example experiment settings

In the example experiment, you can view the endogenous control profile of ACTB, GAPDH, GH1, and LPIN1 the QC Plot screen.

View the QC Plot

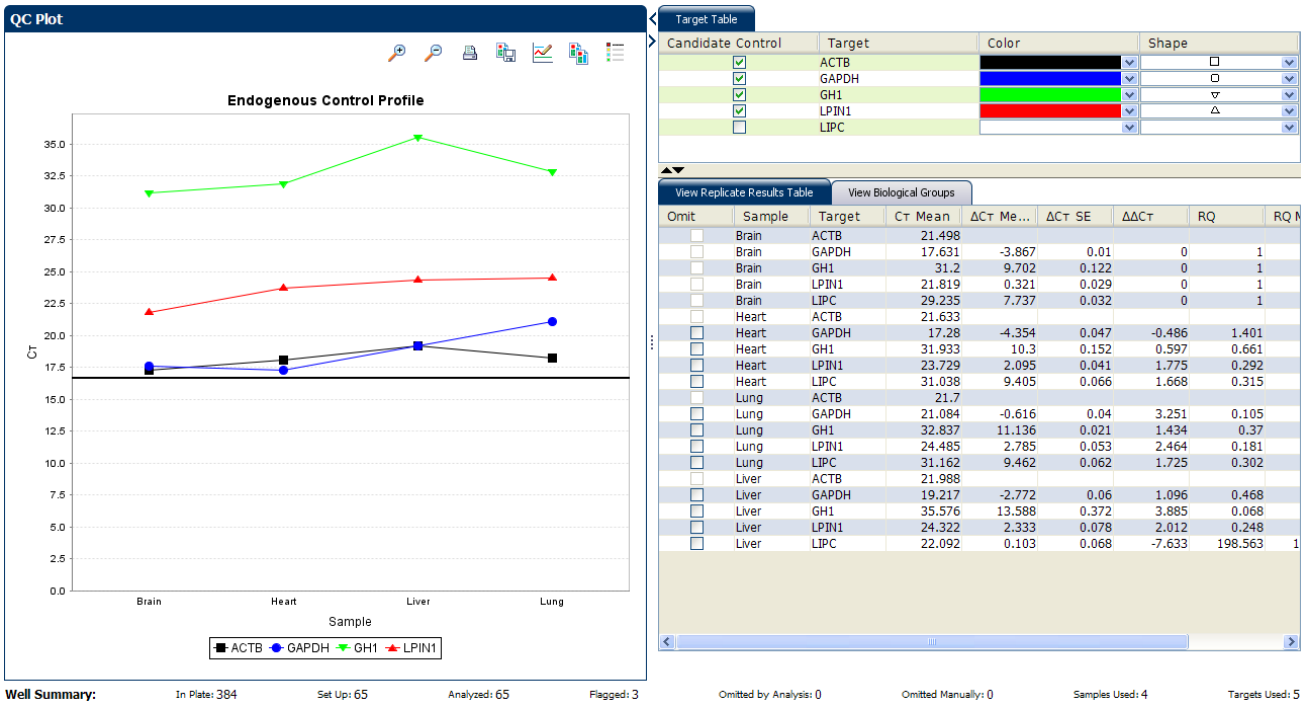
1. From the Experiment Menu pane, select **Analysis ▸ QC Plot**.

   Note: If no data are displayed, click **Analyze**.

2. In the QC Plot screen, click **Target Table**.
   a. In the Candidate Control column, select the check box of the target of the endogenous control profile to plot. In the example experiment, the endogenous controls selected are ACTB, GAPDH, GH1, and LPIN1.
   b. Select a color for each target, from the Color drop-down menu.
   c. Select a shape for each target, from the Shape drop-down menu.
3. Click the **View Replicate Results Table**.
4. Select the check box of the samples to plot. In the example experiment, all the four samples, Brain, Heart, Liver, and Lung are selected.
5. Click  **Show a legend for the plot** (default).

   Note: This is a toggle button. When the legend is displayed, the button changes to Hide the plot legend.

The following is an image of the QC Plot in the Comparative C_T example experiment:



Note: This example experiment does not define Biological Groups.

Review the flags in the QC Summary

The QC Summary screen displays a list of the QuantStudio™ 6 and 7 Flex Software flags, including the flag frequency and location for the open experiment.

Review the QC Summary screen in the Comparative C_T example experiment for any flags triggered by the experiment data. Wells G9, G10, and G11 have data that triggered the HIGHSD flag.

View the QC Summary

1. From the Experiment Menu pane, select **Analysis ▸ QC Summary**.

   Note: If no data are displayed, click **Analyze**.

2. Review the Flags Summary.

   Note: A 0 displayed in the Frequency column indicates that the flag does not appear in the experiment. If the frequency is >0, the flag appears somewhere in the experiment; the well position is listed in the Wells column.

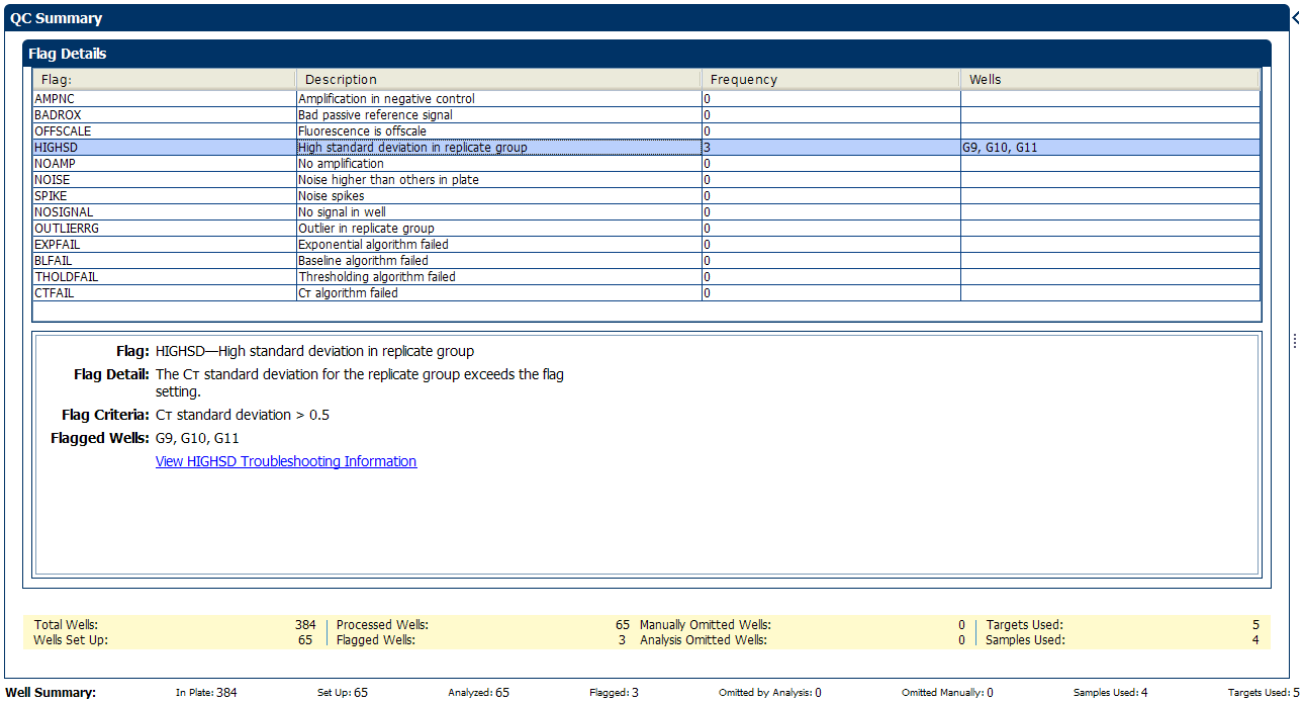
   In the example experiment, there are seven flagged wells.

3. In the Flag Details table, click each flag with a frequency >0 to display detailed information about the flag. In the example experiment. The HIGHSD flag appears three times, in the wells G9, G10, and G11, indicating high standard deviation in the replicate group.

   Note: The HIGHSD flag appears because the C_T values exceed the expected range due to low expression of the GH1 gene in the Liver sample.

4. *(Optional)* For those flags with frequency >0, click the troubleshooting link to view information on correcting the flag.

   The following is an image of the QC Summary for the example experiment:



Possible flags

The flags listed below may be triggered by the experiment data.

Flag	Description
Pre-processing flag	
OFFSCALE	Fluorescence is offscale
Primary analysis flags	
BADROX	Bad passive reference signal
NOAMP	No amplification
NOISE	Noise higher than others in plate
SPIKE	Noise spikes
NOSIGNAL	No signal in well
EXPFAIL	Exponential algorithm failed
BLFAIL	Baseline algorithm failed
THOLDFAIL	Thresholding algorithm failed

Flag	Description
CTFAIL	C_T algorithm failed
Secondary analysis flags	
OUTLIERRG	Outlier in replicate group
AMPNC	Amplification in negative control
HIGHSD	High standard deviation in replicate group

For more information

For more information on...	Refer to...	Publication number
Publishing data	Chapter 2 in Booklet 1, *Getting Started with QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Experiment*	4489822

Section 11.2 Adjust parameters for re-analysis of your own experiments

Adjust analysis settings

The Analysis Settings dialog box displays the analysis settings for the threshold cycle (C_T), flags, and advanced options.

If the default analysis settings in the QuantStudio™ 6 and 7 Flex Software are not suitable for your own experiment, you can change the settings in the Analysis Settings dialog box, then reanalyze your experiment.

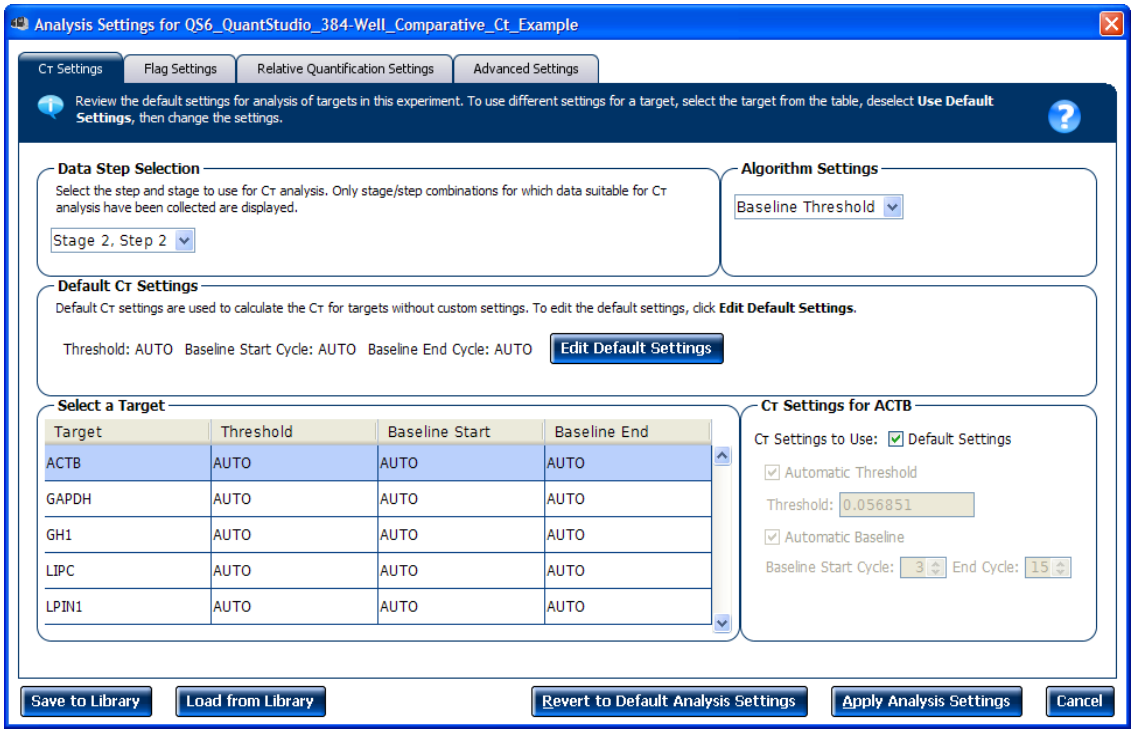
View the analysis settings

1. From the Experiment Menu pane, select **Analysis**.
2. Click **Analysis** ▸ **Analysis Settings** to open the Analysis Settings dialog box.

   In the example experiment, the default analysis settings are used for each tab:

   - C_T Settings
   - Flag Settings
   - Relative Quantification Settings
   - Advanced Settings

   The following is an image of the Analysis Settings dialog box for a Comparative C_T experiment:



3. View and, if necessary, change the analysis settings (see "Adjust analysis settings" below).

   Note: You can save the changes to the analysis settings to the Analysis Settings Library for later use. For more information, see About the Analysis Settings Library in Booklet 1, *Getting Started with QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Experiments*.

4. Click **Apply Analysis Settings** to apply the current analysis settings.

   Note: You can go back to the default analysis settings, by clicking **Revert to Default Analysis Settings**.

Adjust analysis settings

C_T Settings

- **Data Step Selection**

  Use this feature to select one stage/step combination for C_T analysis when there is more than one data collection point in the run method**.**

- **Algorithm Settings**

  Use the Baseline Threshold algorithm to determine the C_T values.

  The Baseline Threshold algorithm is an expression estimation algorithm that subtracts a baseline component and sets a fluorescent threshold in the exponential region for gene quantification.

- **Default C_T Settings**

  Use the default C_T settings feature to calculate C_T for the targets that do not have custom settings. To edit the default settings, click **Edit Default Settings**.

- **C_T Settings for Target**

  When you manually set the threshold and baseline, Life Technologies recommends:

Setting	Recommendation
Threshold	Enter a value for the threshold so that the threshold is: • Above the background. • Below the plateau and linear regions of the amplification curve. • Within the exponential phase of the amplification curve.
Baseline	Select the Start Cycle and End Cycle values so that the baseline ends before significant fluorescent signal is detected.

  Note: This setting is applicable only to the Baseline Threshold algorithm.

  Note: Selecting Automatic Threshold implies selection of automatic setting of the baseline. However, if Automatic Threshold is deselected, then you can choose between setting the baseline either automatically or manually.

Flag Settings

Use the Flag Settings tab to:

- Adjust the sensitivity so that more wells or fewer wells are flagged.
- Change the flags that are applied by the QuantStudio™ 6 and 7 Flex Software.

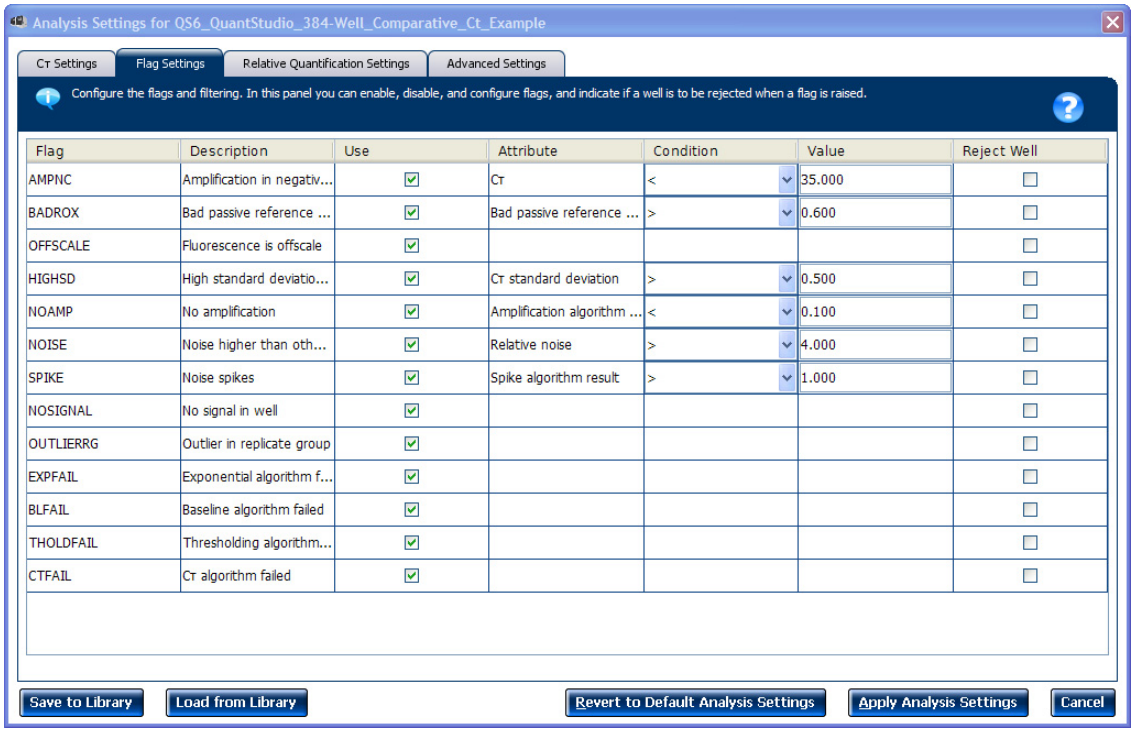
To adjust the flag settings

1. In the Use column, select the check boxes for flags to apply during analysis.
2. *(Optional)* If an attribute, condition, and value are listed for a flag, specify the setting for applying the flag.

   Note: If you choose to adjust the setting for applying a flag, make minor adjustments as you evaluate the appropriate setting.
3. In the Reject Well column, select the check boxes if you want the software to reject wells with the flag.

   Note: After you have rejected the flagged wells, analysis results depend on factors such as the experiment type and flag type. For example, rejecting wells flagged by HIGHSD in experiments using the Standard Deviation calculations may change the result of C_T SD. For some flags, analysis results calculated before the well is rejected are maintained.
4. Click **Apply Analysis Settings** in the Analysis Settings dialog box. If the run status is complete, the data are reanalyzed.

   The following is an image of the Flag Settings tab:



Relative Quantification Settings

Use the Relative Quantification Settings tab to:

- Change the type of analysis, singleplex or multiplex.
- Change the reference sample and/or endogenous control.

- Reject Outliers with ΔC_Tvalues less than or equal to the entered value.

  Note: The Outlier Rejection settings apply only to multiplex reactions.

- Select the algorithm to use to determine the relative quantification minimum and maximum values (error bars):
  - **Confidence Level** - Select to calculate the RQ minimum and maximum values based on the selected confidence level. Select the confidence level to use.
  - **Standard Deviations** - Select to calculate the RQ minimum and maximum values based on the selected number of standard deviations. Select the number of standard deviations to use.

Advanced Settings

Use the Advanced Settings tab to change baseline settings well-by-well.

To use custom baseline settings for a well-target combination:

1. Select one or more well-target combinations in the table.
2. Deselect the **Use C_T Settings Defined for Target** check box.
3. Define the custom baseline settings:
   - For automatic baseline calculations, select the **Automatic Baseline** check box.
   - To define the baseline, deselect the **Automatic Baseline** check box, then enter the baseline start cycle and baseline end cycle.

Improve C_T precision by omitting wells

Experimental error may cause some wells to be amplified insufficiently or not at all. These wells typically produce C_T values that differ significantly from the average for the associated replicate wells. If included in the calculations, these outliers can result in erroneous measurements; to ensure C_T precision, omit the outliers from the analysis.

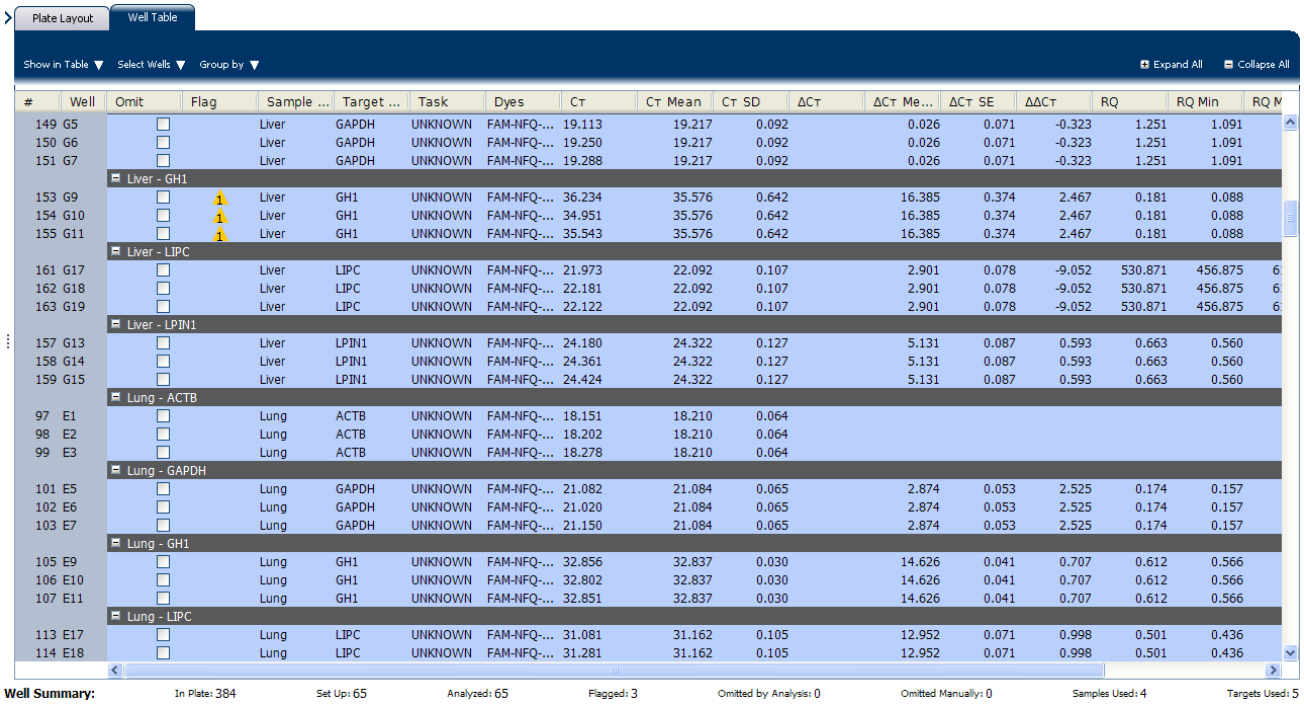
In the Comparative C_T example experiment, there are seven outliers. To remove these wells from analysis.

1. From the Experiment Menu pane, select **Analysis ▸ Amplification Plot**.

   Note: If no data are displayed, click **Analyze**.

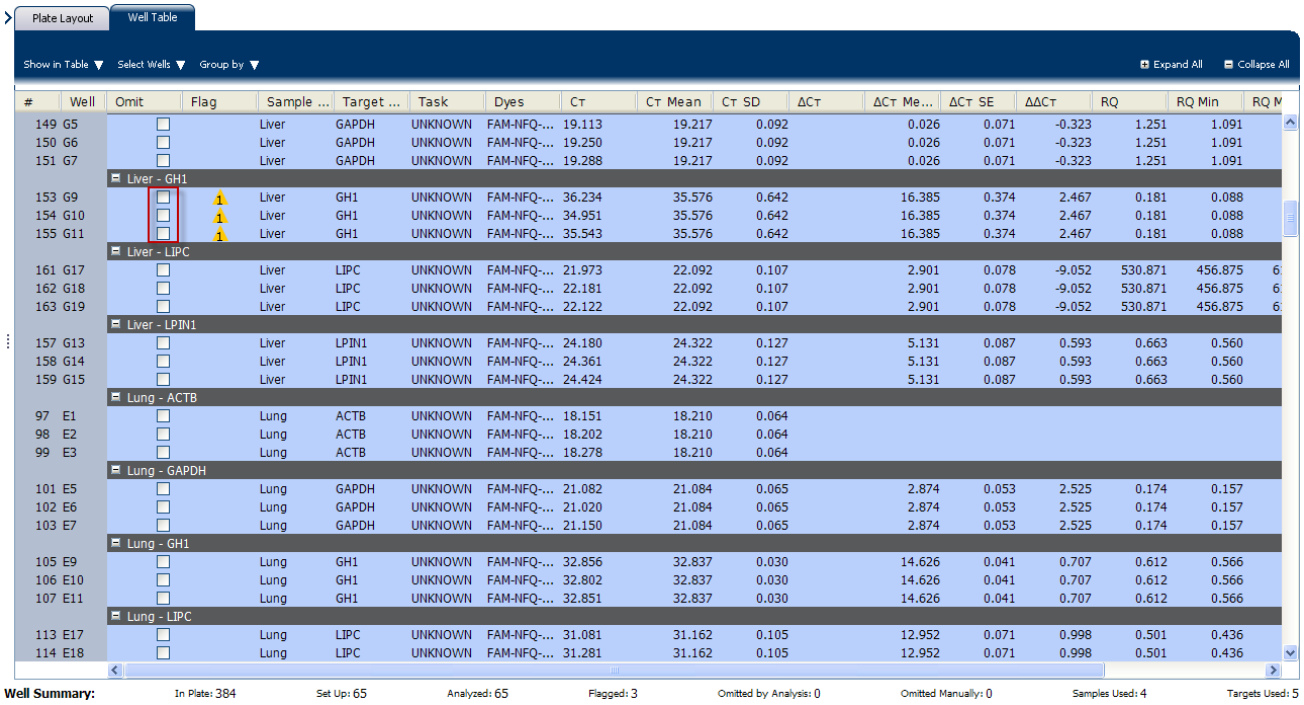
2. In the Amplification Plot screen, select **C_T vs. Well** from the Plot Type drop-down menu.
3. Select the **Well Table** tab.
4. In the Well Table, identify outliers:
   a. From the Group By drop-down menu, select **Replicate**.

b. Look for outliers in the replicate group (make sure they are flagged). In the example experiment, wells G9, G10, and G11 have outliers.



#	Well	Omit	Flag	Sample ...	Target ...	Task	Dyes	CT	CT Mean	CT SD	ΔCT	ΔCT Me...	ΔCT SE	ΔΔCT	RQ	RQ Min	RQ M
149	G5	☐		Liver	GAPDH	UNKNOWN	FAM-NFQ-...	19.113	19.217	0.092		0.026	0.071	-0.323	1.251	1.091	
150	G6	☐		Liver	GAPDH	UNKNOWN	FAM-NFQ-...	19.250	19.217	0.092		0.026	0.071	-0.323	1.251	1.091	
151	G7	☐		Liver	GAPDH	UNKNOWN	FAM-NFQ-...	19.288	19.217	0.092		0.026	0.071	-0.323	1.251	1.091	
		Liver - GH1															
153	G9	☐	1	Liver	GH1	UNKNOWN	FAM-NFQ-...	36.234	35.576	0.642		16.385	0.374	2.467	0.181	0.088	
154	G10	☐	1	Liver	GH1	UNKNOWN	FAM-NFQ-...	34.951	35.576	0.642		16.385	0.374	2.467	0.181	0.088	
155	G11	☐	1	Liver	GH1	UNKNOWN	FAM-NFQ-...	35.543	35.576	0.642		16.385	0.374	2.467	0.181	0.088	
		Liver - LIPC															
161	G17	☐		Liver	LIPC	UNKNOWN	FAM-NFQ-...	21.973	22.092	0.107		2.901	0.078	-9.052	530.871	456.875	
162	G18	☐		Liver	LIPC	UNKNOWN	FAM-NFQ-...	22.181	22.092	0.107		2.901	0.078	-9.052	530.871	456.875	
163	G19	☐		Liver	LIPC	UNKNOWN	FAM-NFQ-...	22.122	22.092	0.107		2.901	0.078	-9.052	530.871	456.875	
		Liver - LPIN1															
157	G13	☐		Liver	LPIN1	UNKNOWN	FAM-NFQ-...	24.180	24.322	0.127		5.131	0.087	0.593	0.663	0.560	
158	G14	☐		Liver	LPIN1	UNKNOWN	FAM-NFQ-...	24.361	24.322	0.127		5.131	0.087	0.593	0.663	0.560	
159	G15	☐		Liver	LPIN1	UNKNOWN	FAM-NFQ-...	24.424	24.322	0.127		5.131	0.087	0.593	0.663	0.560	
		Lung - ACTB															
97	E1	☐		Lung	ACTB	UNKNOWN	FAM-NFQ-...	18.151	18.210	0.064							
98	E2	☐		Lung	ACTB	UNKNOWN	FAM-NFQ-...	18.202	18.210	0.064							
99	E3	☐		Lung	ACTB	UNKNOWN	FAM-NFQ-...	18.278	18.210	0.064							
		Lung - GAPDH															
101	E5	☐		Lung	GAPDH	UNKNOWN	FAM-NFQ-...	21.082	21.084	0.065		2.874	0.053	2.525	0.174	0.157	
102	E6	☐		Lung	GAPDH	UNKNOWN	FAM-NFQ-...	21.020	21.084	0.065		2.874	0.053	2.525	0.174	0.157	
103	E7	☐		Lung	GAPDH	UNKNOWN	FAM-NFQ-...	21.150	21.084	0.065		2.874	0.053	2.525	0.174	0.157	
		Lung - GH1															
105	E9	☐		Lung	GH1	UNKNOWN	FAM-NFQ-...	32.856	32.837	0.030		14.626	0.041	0.707	0.612	0.566	
106	E10	☐		Lung	GH1	UNKNOWN	FAM-NFQ-...	32.802	32.837	0.030		14.626	0.041	0.707	0.612	0.566	
107	E11	☐		Lung	GH1	UNKNOWN	FAM-NFQ-...	32.851	32.837	0.030		14.626	0.041	0.707	0.612	0.566	
		Lung - LIPC															
113	E17	☐		Lung	LIPC	UNKNOWN	FAM-NFQ-...	31.081	31.162	0.105		12.952	0.071	0.998	0.501	0.436	
114	E18	☐		Lung	LIPC	UNKNOWN	FAM-NFQ-...	31.281	31.162	0.105		12.952	0.071	0.998	0.501	0.436	

c. Select the **Omit** check box next to outlying well(s).



#	Well	Omit	Flag	Sample ...	Target ...	Task	Dyes	CT	CT Mean	CT SD	ΔCT	ΔCT Me...	ΔCT SE	ΔΔCT	RQ	RQ Min	RQ M
149	G5	☐		Liver	GAPDH	UNKNOWN	FAM-NFQ-...	19.113	19.217	0.092		0.026	0.071	-0.323	1.251	1.091	
150	G6	☐		Liver	GAPDH	UNKNOWN	FAM-NFQ-...	19.250	19.217	0.092		0.026	0.071	-0.323	1.251	1.091	
151	G7	☐		Liver	GAPDH	UNKNOWN	FAM-NFQ-...	19.288	19.217	0.092		0.026	0.071	-0.323	1.251	1.091	
		Liver - GH1															
153	G9	☐	1	Liver	GH1	UNKNOWN	FAM-NFQ-...	36.234	35.576	0.642		16.385	0.374	2.467	0.181	0.088	
154	G10	☐	1	Liver	GH1	UNKNOWN	FAM-NFQ-...	34.951	35.576	0.642		16.385	0.374	2.467	0.181	0.088	
155	G11	☐	1	Liver	GH1	UNKNOWN	FAM-NFQ-...	35.543	35.576	0.642		16.385	0.374	2.467	0.181	0.088	
		Liver - LIPC															
161	G17	☐		Liver	LIPC	UNKNOWN	FAM-NFQ-...	21.973	22.092	0.107		2.901	0.078	-9.052	530.871	456.875	
162	G18	☐		Liver	LIPC	UNKNOWN	FAM-NFQ-...	22.181	22.092	0.107		2.901	0.078	-9.052	530.871	456.875	
163	G19	☐		Liver	LIPC	UNKNOWN	FAM-NFQ-...	22.122	22.092	0.107		2.901	0.078	-9.052	530.871	456.875	
		Liver - LPIN1															
157	G13	☐		Liver	LPIN1	UNKNOWN	FAM-NFQ-...	24.180	24.322	0.127		5.131	0.087	0.593	0.663	0.560	
158	G14	☐		Liver	LPIN1	UNKNOWN	FAM-NFQ-...	24.361	24.322	0.127		5.131	0.087	0.593	0.663	0.560	
159	G15	☐		Liver	LPIN1	UNKNOWN	FAM-NFQ-...	24.424	24.322	0.127		5.131	0.087	0.593	0.663	0.560	
		Lung - ACTB															
97	E1	☐		Lung	ACTB	UNKNOWN	FAM-NFQ-...	18.151	18.210	0.064							
98	E2	☐		Lung	ACTB	UNKNOWN	FAM-NFQ-...	18.202	18.210	0.064							
99	E3	☐		Lung	ACTB	UNKNOWN	FAM-NFQ-...	18.278	18.210	0.064							
		Lung - GAPDH															
101	E5	☐		Lung	GAPDH	UNKNOWN	FAM-NFQ-...	21.082	21.084	0.065		2.874	0.053	2.525	0.174	0.157	
102	E6	☐		Lung	GAPDH	UNKNOWN	FAM-NFQ-...	21.020	21.084	0.065		2.874	0.053	2.525	0.174	0.157	
103	E7	☐		Lung	GAPDH	UNKNOWN	FAM-NFQ-...	21.150	21.084	0.065		2.874	0.053	2.525	0.174	0.157	
		Lung - GH1															
105	E9	☐		Lung	GH1	UNKNOWN	FAM-NFQ-...	32.856	32.837	0.030		14.626	0.041	0.707	0.612	0.566	
106	E10	☐		Lung	GH1	UNKNOWN	FAM-NFQ-...	32.802	32.837	0.030		14.626	0.041	0.707	0.612	0.566	
107	E11	☐		Lung	GH1	UNKNOWN	FAM-NFQ-...	32.851	32.837	0.030		14.626	0.041	0.707	0.612	0.566	
		Lung - LIPC															
113	E17	☐		Lung	LIPC	UNKNOWN	FAM-NFQ-...	31.081	31.162	0.105		12.952	0.071	0.998	0.501	0.436	
114	E18	☐		Lung	LIPC	UNKNOWN	FAM-NFQ-...	31.281	31.162	0.105		12.952	0.071	0.998	0.501	0.436	

5. Click **Analyze** to reanalyze the experiment data with the outlying well(s) removed from the analysis.

   Note: You can also omit undesirable wells in an experiment from the Plate Layout screen. To omit a well from the Plate Layout screen, right-click the well and select **Omit**.

12 Export Analysis Results

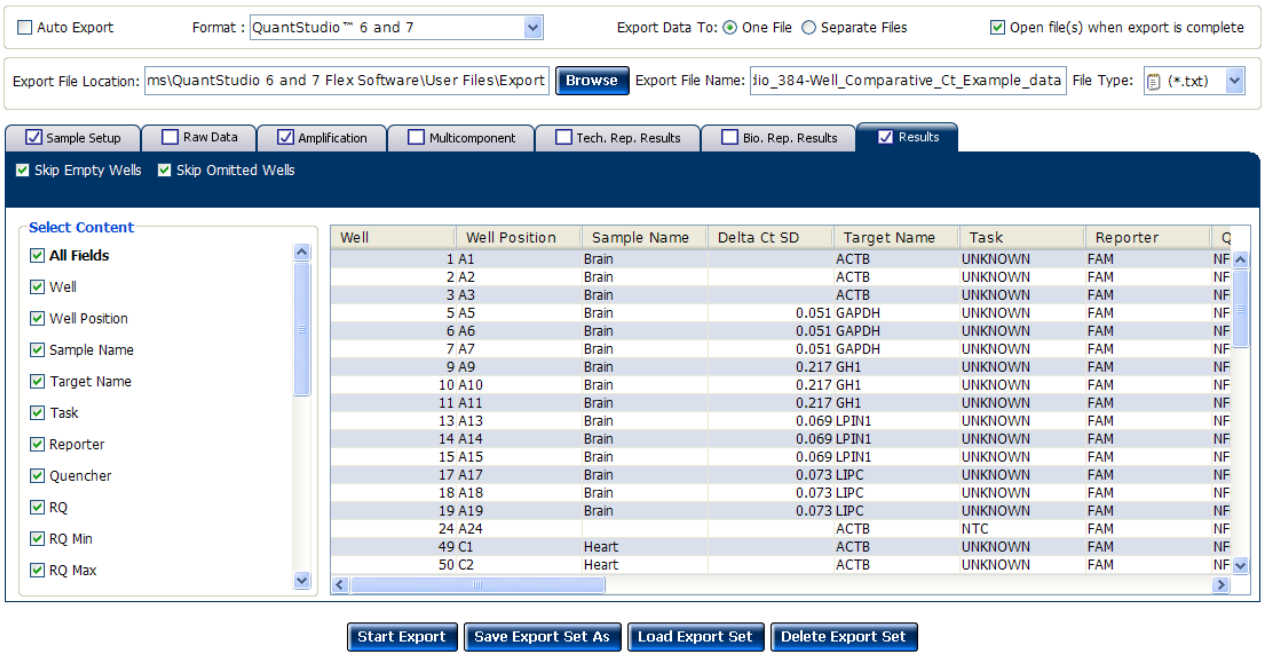
1. Open the Comparative C_T example experiment file that you analyzed in Chapter 11.
2. In the Experiment Menu, click  **Export.**

   Note: To export data automatically after analysis, select the **Auto Export** check box during experiment setup or before running the experiment. Auto export is unchecked for the example experiment.
3. Select **QuantStudio™ 6 and 7** format.

   Note: Select **7900** Format if you want to export the Clipped Data.
4. Complete the Export dialog box as shown below:

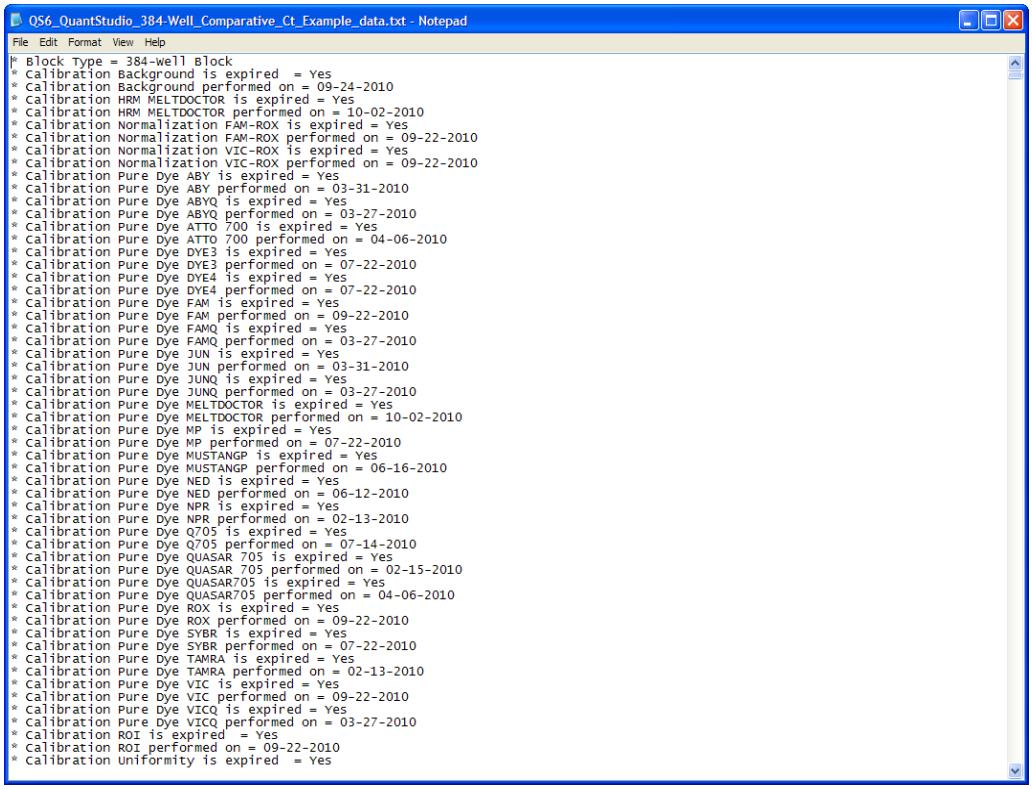
Field or Selection	Entry
Select Data to export/ Select Content	Results
Export Data To	One File
Export File Name	QS6_QuantStudio_384-Well_Comparative_Ct_Example_data
File Type	*.txt
Export File Location	C:\Applied Biosystems\QuantStudio 6 and 7 Flex Software\User Files\Export

Your Export screen should look like this:



Well	Well Position	Sample Name	Delta Ct SD	Target Name	Task	Reporter	Q
1	A1	Brain		ACTB	UNKNOWN	FAM	NF
2	A2	Brain		ACTB	UNKNOWN	FAM	NF
3	A3	Brain		ACTB	UNKNOWN	FAM	NF
5	A5	Brain	0.051	GAPDH	UNKNOWN	FAM	NF
6	A6	Brain	0.051	GAPDH	UNKNOWN	FAM	NF
7	A7	Brain	0.051	GAPDH	UNKNOWN	FAM	NF
9	A9	Brain	0.217	GH1	UNKNOWN	FAM	NF
10	A10	Brain	0.217	GH1	UNKNOWN	FAM	NF
11	A11	Brain	0.217	GH1	UNKNOWN	FAM	NF
13	A13	Brain	0.069	LPIN1	UNKNOWN	FAM	NF
14	A14	Brain	0.069	LPIN1	UNKNOWN	FAM	NF
15	A15	Brain	0.069	LPIN1	UNKNOWN	FAM	NF
17	A17	Brain	0.073	LIPC	UNKNOWN	FAM	NF
18	A18	Brain	0.073	LIPC	UNKNOWN	FAM	NF
19	A19	Brain	0.073	LIPC	UNKNOWN	FAM	NF
24	A24			ACTB	NTC	FAM	NF
49	C1	Heart		ACTB	UNKNOWN	FAM	NF
50	C2	Heart		ACTB	UNKNOWN	FAM	NF

Your exported file when opened in Notepad should look like this:



13 Design and Analyze a Gene Expression Study

The QuantStudio™ 6 and 7 Flex Real-Time PCR Development Software can combine the analysis of experiments that use the Relative Standard Curve or the Comparative C_T ($\Delta\Delta C_T$) quantification methods into a Gene Expression study. A study provides a wider range for analyzing and comparing target behavior across multiple experiments.

Note: You can import different types of quantification experiments into a single Gene Expression study, but make sure the run method and experiment type are identical for all the experiments in that study. Also make sure that the experiments have been run on the same type of qPCR instrument.

Note: You can design an example study using the example experiments provided with the QuantStudio™ 6 and 7 Flex Real-Time PCR Development Software. This chapter explains how to design and analyze multiple Comparative C_T experiments as a study. When you design a Gene Expression study, make sure that each experiment in that Gene Expression study has a unique name. Absence of a unique name, leads to failure of the run.

This chapter covers:

About Gene Expression studies

In a Gene Expression study, you can...	You cannot...
• Specify the endogenous control(s) and reference sample for the study. • Set individual efficiency values for each target. • Select the control type when applicable. • Set baseline and threshold values and confidence levels, or set the number of standard deviations for Comparative $_{CT}$ Min./Max. • Omit wells individually or together through their association with replicate groups (technical or biological).	• Create, add, or modify samples. • Create, add, or modify targets. • Change assay tasks. You can perform these operations in individual experiments.

About the example study

In the Comparative C_T example study:

- Two reaction plates (experiments) are used.
- Experiments that you add to the study are two Comparative C_T experiments that have already been analyzed.
- The cDNA was prepared from total RNA that was isolated from the following 4 samples:
  - Heart
  - Liver
  - Brain
  - Lung
- Five targets (assays) are used:
  - LIPC: Hs00165106_m1
  - GAPDH: Hs99999905_m1
  - ACTB: Hs99999903_m1
  - LP1N1: Hs00299515_m1
  - GH1: Hs00236859_m1
- The reference sample is Brain.
- The endogenous control is ACTB.
- Each experiment in the study was designed for singleplex PCR, where the target and endogenous control assays are run in separate wells.
- For each experiment in the study, reactions were set up for 2-step RT-PCR:
  - The cDNA was reverse-transcribed from total RNA samples using Invitrogen VILO Kit.
  - The reactions were prepared using TaqMan® Fast Universal PCR Master Mix (2X).

Design a study

Create a new study

To create a new study in the QuantStudio™ 6 and 7 Flex Real-Time PCR Development Software, from the Home screen, click **New Gene Expression Study**.

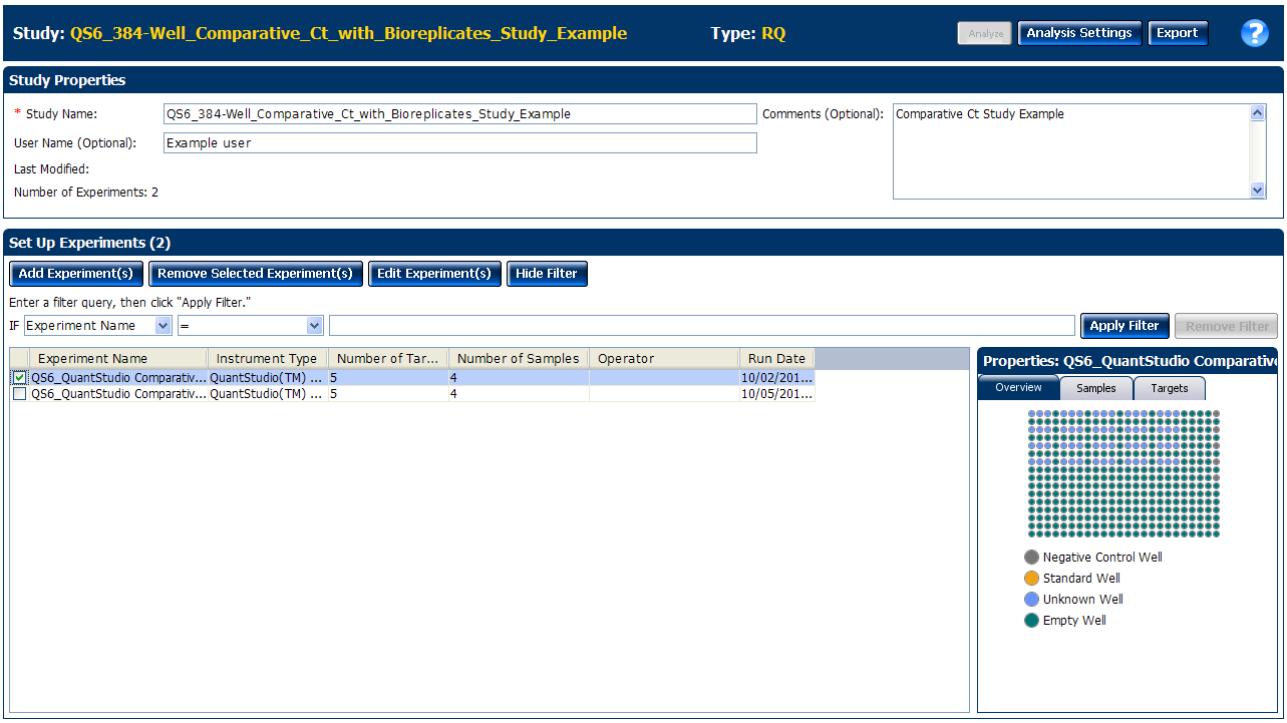
Set up the study in the Study Properties screen

1. In the Study Menu pane, select **Setup ▸ Study Properties**
2. In the Study Properties pane, click the **Study Name** field, then enter QS6_384-Well_Comparative_Ct_with_Bioreplicates_Study_Example.
3. Click the **Comments** field, then enter **Example Comparative C_T Study for the Comparative Ct Experiments**.
4. In the Setup Experiments pane, click **Add Experiment(s)**.
5. In the Open dialog box, browse to the **QS6_384-Well_Comparative_Ct_Example_1.eds and QS6_384-Well_Comparative_Ct_Example_2.eds** files at:

   C:\Program Files\Applied Biosystems\QuantStudio 6 and 7 Flex Software\examples\QS6Flex

   The QuantStudio™ 6 and 7 Flex Real-Time PCR Development Software displays the details of the experiment in the Properties pane.

   Your Study Properties screen should look like this:



Filter the experiments in the study

To narrow your search for an experiment, define and apply a filter:

1. In the left-most drop-down menu, select an experiment attribute to query.
2. In the center drop-down menu, select an operator for the query.
3. In the right-most field, enter the condition to look for, then click **Apply Filter**.

After you apply a filter, click **Hide Filter/Show Filter** to hide or show the filter tool, or click **Remove Filter** to remove the filter.

Define Replicates

In the Define Replicates screen, create biological replicate groups and use them to associate samples for the analysis. Biological replicates allow you to assess the representative nature of your results as they relate to the population being studied. Including biological replicates can give insight into any natural variation that is in the population.

Example study settings

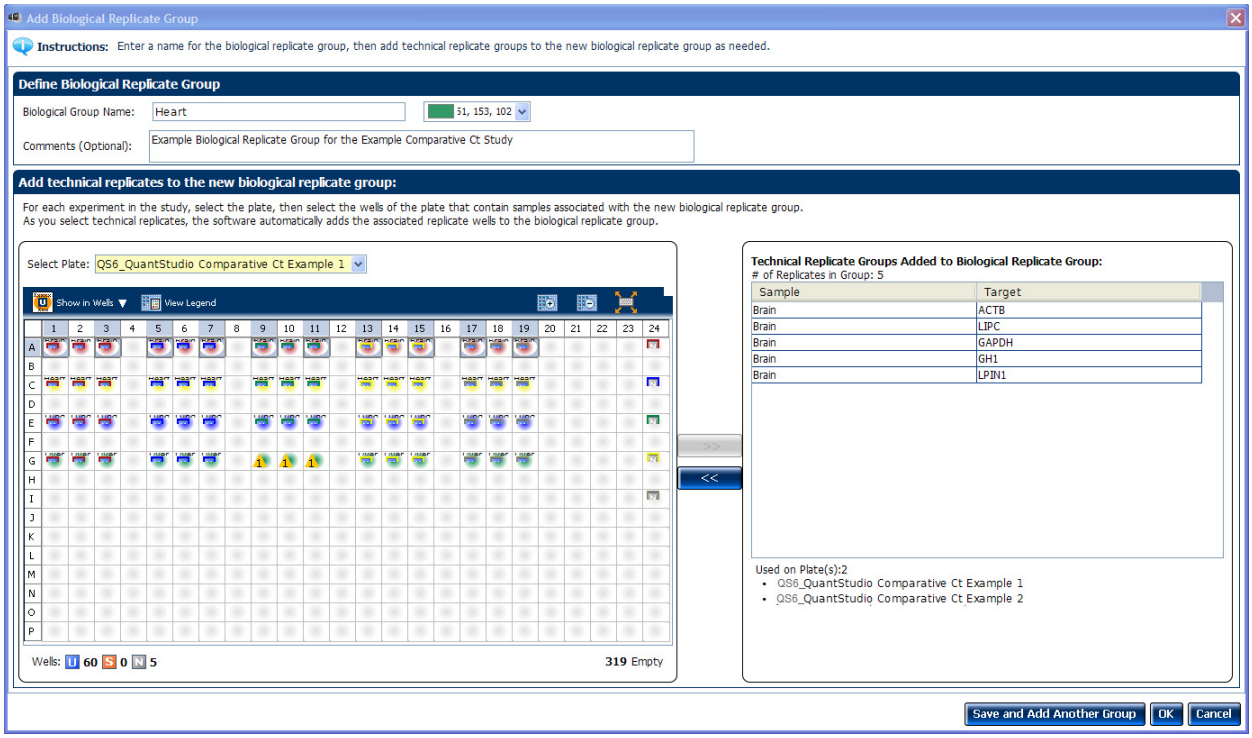
The Gene Expression example study contains four biological replicate groups. These are lung, heart, brain, and liver. Each biological groups consists eight replicates.

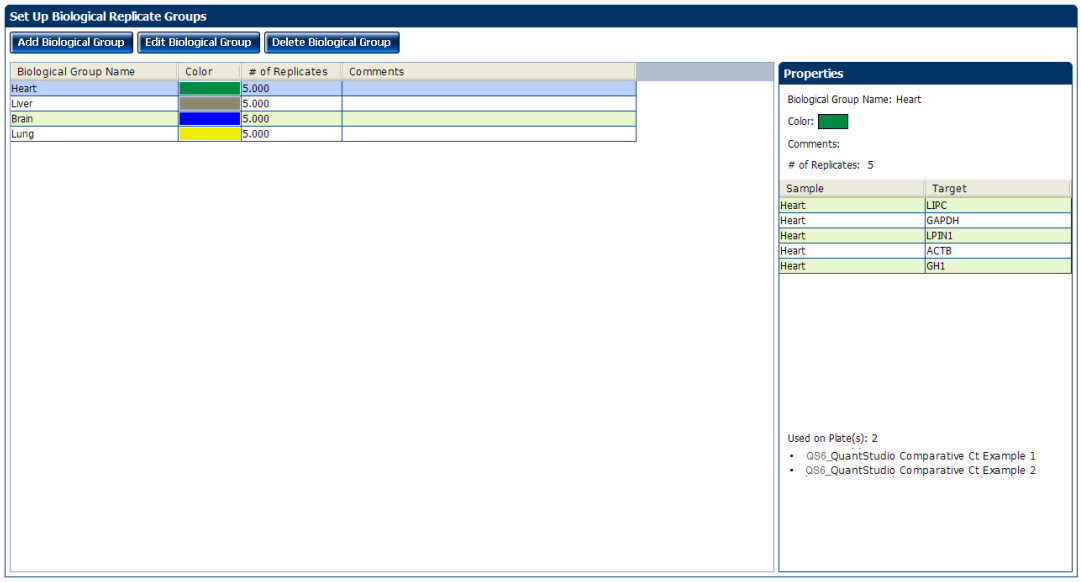
To define replicates

1. In the Study Menu pane, select **Setup ▸ Define Replicates**.
2. Click **Add Biological Group** to open the Add Biological Replicate Group dialog box.
3. Define the biological replicate group:
    a. Click the **Biological Group Name** field, then enter **Heart.**
    b. In the Color field, select the color.
    c. Click the **Comments** field, then enter **Example Biological Replicate Group for the Example Comparative Ct Study**.
4. Add technical replicates:
    a. From the Select Plate drop-down menu, select **QuantStudio Comparative Ct Example 1.eds**.
    b. In the plate layout, select wells **A1, A3, A5, A7, A9, A11, A13, A15, A17, and A19,** then click 

to add the technical replicate wells that are associated with the selected well to the biological group.
    c. Click **OK**.
    d. Perform steps a through c to add the remaining technical replicate wells associated with the selected well to the biological group.

5. On the Set Up Biological Replicates screen, select the **Heart** biological group that you just added to the study. The QuantStudio™ 6 and 7 Flex Real-Time PCR Development Software displays the details of the biological group in the Properties pane.





6. Click **Analyze**, then close the study:
   a. Select **File** ▸ **Close**.
   b. At the prompt, click **Yes** to save the changes.

c. In the Save Study as dialog box, click **Save** to accept the default file name and location. The example study is saved and closed, and you are returned to the Home screen. You can also save the study you create at C:\Program Files\Applied Biosystems\QuantStudio 6 and 7 Flex Software\examples\QS6Flex.

Edit a Biological Replicate Group

1. Open the study of interest.
2. In the Study Menu pane, select **Setup** ▸ **Define Replicates**.
3. Click **Edit Biological Group** to open the Edit Biological Replicate Group dialog box.
4. Edit the group information:
   a. From the Biological Group Name drop-down menu, select the group to edit.
   b. To change the group name, click **Edit**, enter a new name, then click **OK**.
   c. To change the color, select a color from the drop-down menu.
   d. (Optional) Enter comments.
5. From the Select Plate drop-down menu, select the experiment of interest.
6. Add samples to the biological group:
   a. In the Plate Layout, select the well(s) of the plate that contain samples to add to the biological group.
   b. Click  to add the samples that are associated with the selected wells to the biological group.
7. Remove samples from the biological group:
   a. In the Technical Replicate Groups Added to Biological Replicate Group pane, select a sample. You can select only one sample at a time.
   b. Click  to remove the sample from the biological group.
8. Repeat steps 5 through 7 for the other experiments in the study.
9. Click **OK** to save the changes and return to the study; click **Cancel** to exit the dialog box without saving the changes.

Remove Biological Replicate Groups

1. Open the study of interest.
2. In the Study Menu pane, select **Setup** ▸ **Define Replicates**.
3. In the Define Replicates screen, select the group to remove, then click **Delete Biological Group**.
4. Click **Yes** to remove the group from the study; click **No** to keep the group in the study.

Tips for designing your own study

- Enter a study name that is descriptive and easy to remember. You can enter up to 100 characters in the Study Name field. The study name is used as the default file name.

  You can only use the alpha-numeric, period (.), hyphen (-), underscore (_), and spaces ( ) characters in the Experiment Name field.
- *(Optional)* Enter comments to describe the study. You can enter up to 1000 characters in the Comments field.
- Use the default user name, or enter a new user name, to identify the owner of the study. You can enter up to 100 characters in the User Name field.

  Note: If security is enabled, the User Name field is automatically populated with the name you selected at log in.
- You can add an unlimited number of experiments (reaction plates) to the study. Click **Add Experiment (s)** or **Remove Selected Experiment(s)** to add or remove experiments to or from a study. Note the following:
  - Each experiment in the study must:
    - Have one or more common endogenous control(s). The endogenous control(s) must be present on each reaction plate within the study.

      Note: The endogenous control gene for a given sample must be run on the same plate as the target gene for a given sample in order for a study to be created.
    - Have identical thermal cycling parameters (the same number of steps and cycles). The 6 and 7 Flex Real-Time PCR Development Software cannot combine in the same study experiments that use Fast and Standard thermal cycling conditions.
    - Have the same passive reference.
    - Have the same experiment type.
    - Have been run on the same instrument type.
  - As the default, the QuantStudio™ 6 and 7 Flex Real-Time PCR Development Software assigns the endogenous control and reference sample for a study based on the analysis settings of the first experiment that you added to the study.
  - If experiments that contain biological replicate groups are added to a study, the QuantStudio™ 6 and 7 Flex Real-Time PCR Development Software automatically merges the matching biological groups.
  - The QuantStudio™ 6 and 7 Flex Real-Time PCR Development Software automatically analyzes a study after you add more than one experiment to it. Consequently, to ensure that the software uses the correct settings, Life Technologies recommends that you review the analysis settings of your study after adding multiple experiments.
- When adding experiments to the study, **Ctrl-click** multiple experiments in the Open dialog box to add them to the study.
- Select an experiment that has been added to the study to view its properties in the Properties pane.
- Filter the experiments added to the study to simplify the list for easier review. See "Filter the experiments in the study" on page 127.

- Enter a biological replicate group name that is descriptive and easy to remember. You can enter up to 100 characters in the Biological Group Name field. You can only use the alpha-numeric, full-stop (.), hyphen (-), underscore (_), and space ( ) characters in the Experiment Name field.
- (Optional) Enter comments to describe the biological replicate group. You can enter up to 1000 characters in the Comments field.
- You can add an unlimited number of technical replicates to a biological group.

IMPORTANT! A sample cannot belong to more than one biological group.

  - Click-drag over the desired wells, or **Ctrl-click** or **Shift-click** in the plate layout to select multiple wells.
  - Click the upper-left corner of the plate layout to select all wells.
- You can change the name of a biological replicate group, change its color identification and description, and add or remove technical replicates. See "Edit a Biological Replicate Group" on page 130.
- You can remove an existing biological replicate group. See "Remove Biological Replicate Groups" on page 130.

IMPORTANT! After you remove a biological replicate group from a study, you cannot restore it.

Analyze the example study

This section explains how to use the QuantStudio™ 6 and 7 Flex Real-Time PCR Development Software to analyze the Gene Expression example study. It also describes Life Technologies recommended best practices as you perform the analysis.

Note: The Comparative C_T Study Example.edm file illustrated in this chapter demonstrates the use of biological replicate groups. You can also create a study without the use of biological replicate groups.

Access the example study

Access the example study that you created on your computer.

C:\Program Files\Applied Biosystems\QuantStudio 6 and 7 Flex Software\examples\QS6_384 Well Comparative Ct with Bioreplicates Study Example.edm

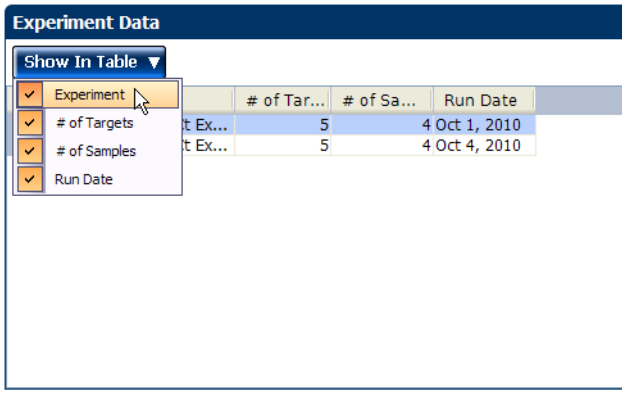
View the experiment data and well results

The Experiment Data and Well Results Data panes appear in the Amplification Plot, Multicomponent Plot, and Multiple Plots View screens.

The Gene Expression screen displays the Replicate Results Data and the Well Results Data panes.

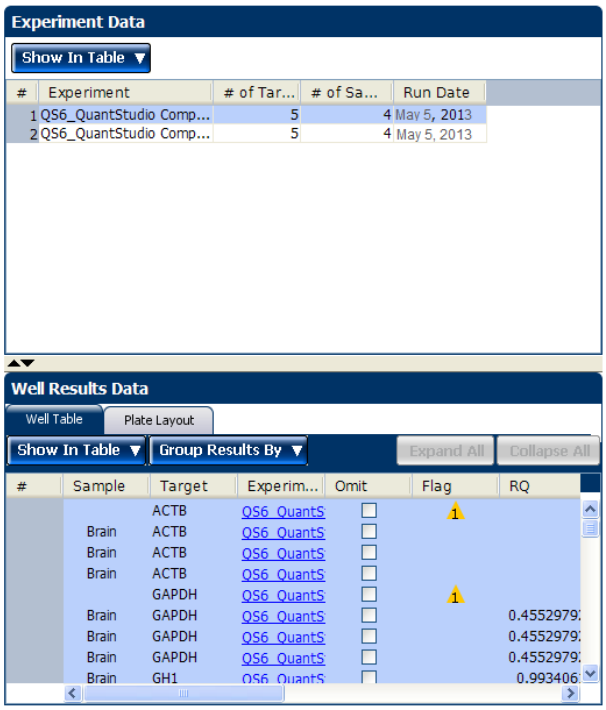
Only the Well Results Data pane appears in the QC Plots View screen.

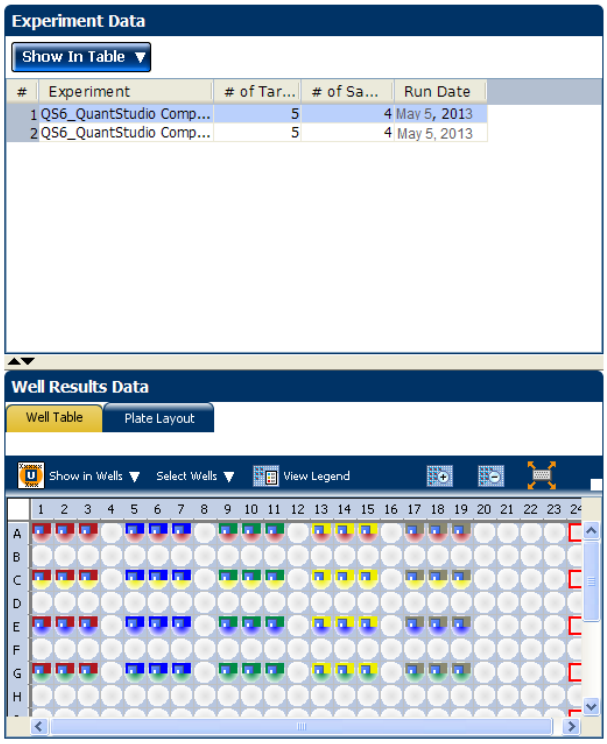
- To display or hide columns in the Experiment Data pane– from the Show In Table drop-down menu, select or deselect one or more options: **Experiments**, **# of Targets**, **# of Samples**, **Run Date**.



- To display a subset of the study data in the plots– Select one or more rows in the Experiment Data pane or the Well Results Data pane, then select the **Hide unselected data from plot** check box appearing in the plot pane to display data only from the selected rows.

The Experiments Data pane lists each reaction plate (experiment) that is added to a study. The data that are displayed in the Well Results Data pane depend on which tab you select in the Experiment Data pane:

Tab	Description	Illustration
Well Table tab	When you select one or more experiments in the Experiment Data pane, the well table displays the wells that make up the selected experiment(s).	

Tab	Description	Illustration
Plate Layout tab	When you select one experiment in the Experiment Data pane, the plate layout displays the plate layout for the selected experiment. If you select more than one experiment in the Experiment Data pane, only the plate layout for the first experiment is displayed in the Plate Layout tab.	

View the analysis settings

The Analysis Settings dialog box displays the analysis settings for the threshold cycle (C_T), flags, and relative quantification. If the default analysis settings in the QuantStudio™ 6 and 7 Flex Real-Time PCR Development Software are not suitable for your study, you can change the settings in the Analysis Settings dialog box, then reanalyze your study.

Note: In the Comparative C_T example study, the default analysis settings are used without changes.

To adjust the analysis settings

1. From the Study Menu pane, select **Analysis**.
2. Click **Analysis Settings** to open the Analysis Settings dialog box.
3. Select the **Relative Quantification Settings** tab, then view the default reference sample and endogenous control. In the example study, the default reference sample is brain and the default endogenous control is 18S.
4. Select the **C_T Settings** tab, then the **Flag Settings** tab. In the example study, the default analysis settings are used in each tab.

5. View and, if necessary, change the analysis settings. For more information on the changes to analysis settings, refer to "Tips for analyzing your own study" on page 135.

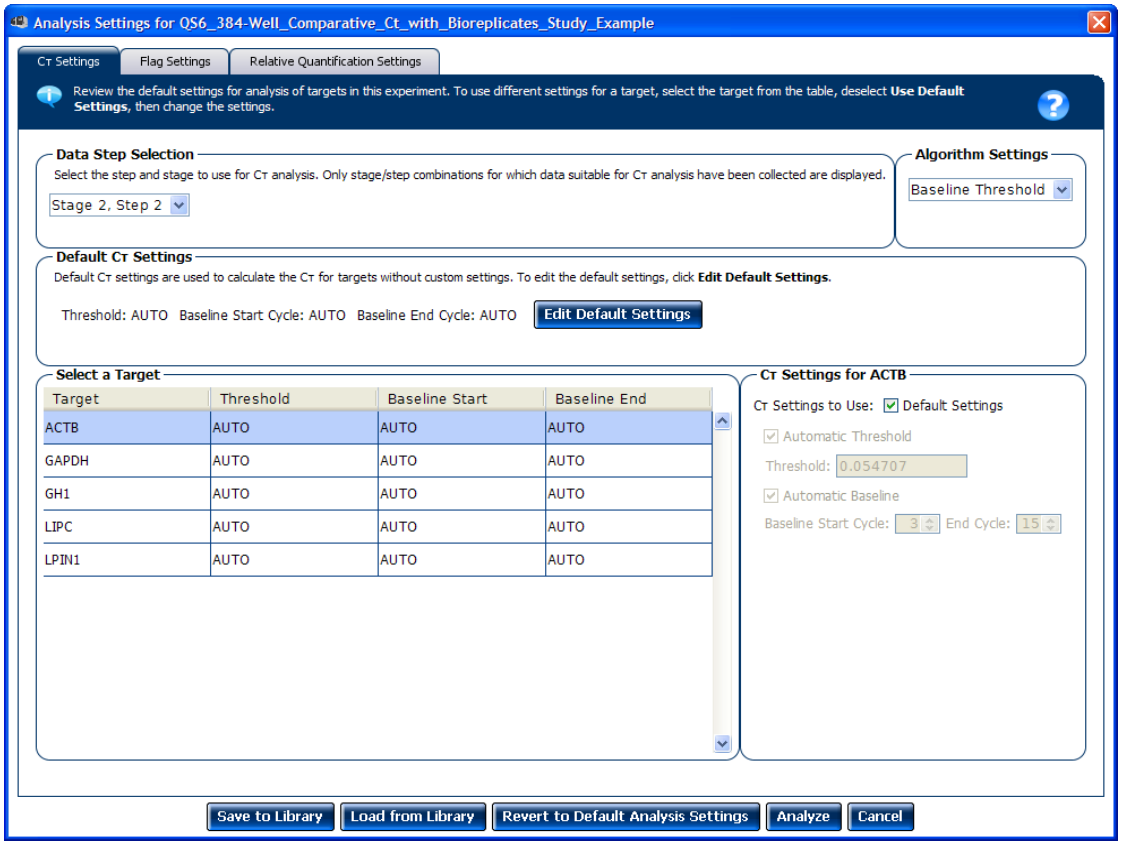
   Note: You can save the changes to the analysis settings to the Analysis Settings Library for later use. For more information, see About the Analysis Settings Library in Booklet 1, *Getting Started with QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Experiments*.

6. Click **Apply Analysis Settings** to apply the current analysis settings.

   Note: You can go back to the default analysis settings by clicking **Revert to Default Analysis Settings**.

7. Click **Analyze**.

   The following is an image of the Analysis Settings dialog box for a Comparative C_T Study:



Tips for analyzing your own study

Unless you have already determined the optimal settings for your study, use the default analysis settings in the QuantStudio™ 6 and 7 Flex Real-Time PCR Development Software. If the default settings are not suitable for your study, change the settings as described below.

C_T Settings

- **Data Step Selection**

  Use this feature to select multiple locations of analysis, in case several are chosen.

- **Algorithm Settings**

Use the Baseline Threshold algorithm to determine the C_T values.

The Baseline Threshold algorithm is an expression estimation algorithm that subtracts a baseline component and sets a fluorescent threshold in the exponential region for gene quantification.

- **Default C_T Settings**

  Use the default C_T settings feature to calculate C_T for the targets that do not have custom settings. To edit the default settings, click **Edit Default Settings**.

- **C_T Settings for Target**

  When you manually set the threshold and baseline, Life Technologies recommends:

Setting	Recommendation
Threshold	Enter a value for the threshold so that the threshold is: • Above the background. • Below the plateau and linear regions of the amplification curve. • Within the exponential phase of the amplification curve.
Baseline	Select the Start Cycle and End Cycle values so that the baseline ends before significant fluorescent signal is detected.

Note: This setting is applicable only to the Baseline-Threshold algorithm.

Note: Selecting Automatic Threshold implies selection of automatic setting of the baseline. However, if Automatic Threshold is deselected, then you can choose between setting the baseline either automatically or manually.

Flag Settings

Use the Flag settings tab to:

- Adjust the sensitivity so that more wells or fewer wells are flagged.
- Change the flags that are applied by the QuantStudio™ 6 and 7 Flex Real-Time PCR Development Software.

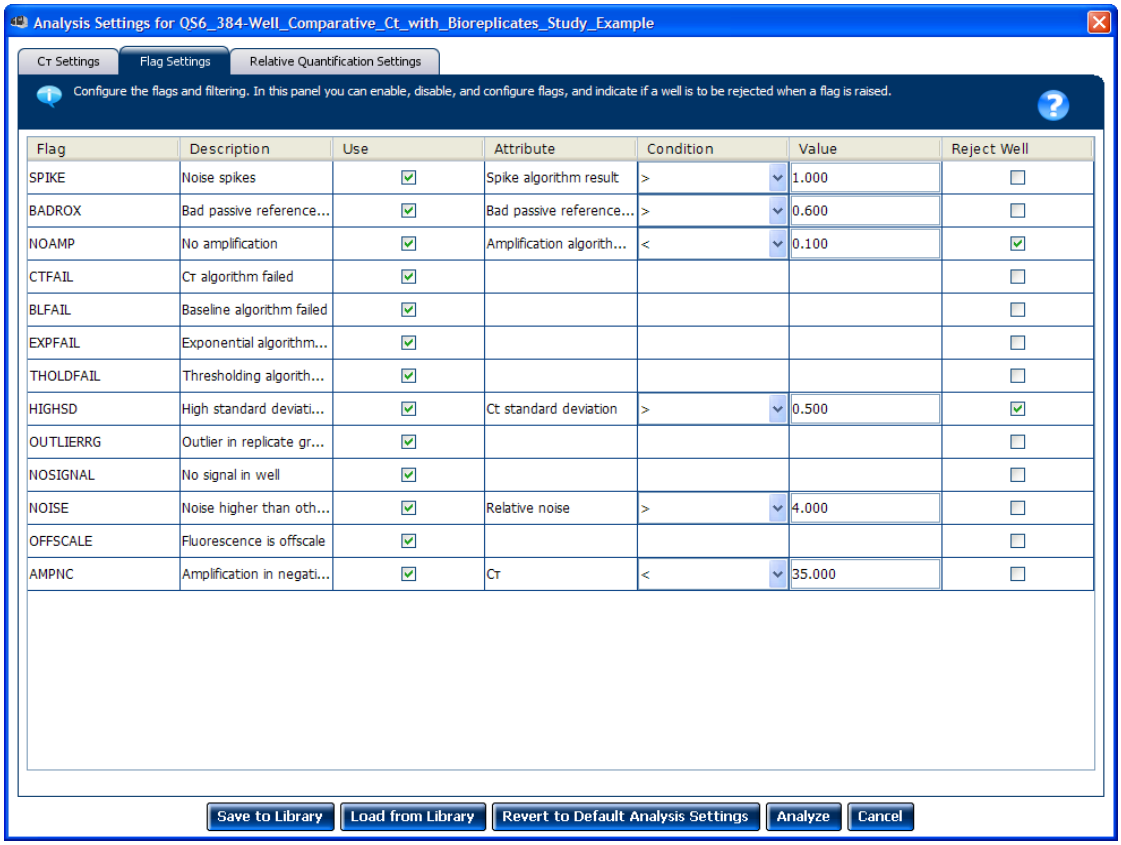
To adjust the flag settings

1. In the Use column, select the check boxes for flags to apply during analysis.
2. *(Optional)* If an attribute, condition, and value are listed for a flag, specify the setting for applying the flag.

   Note: If you choose to adjust the setting for applying a flag, make minor adjustments as you evaluate the appropriate setting.
3. In the Reject Well column, select the check boxes if you want the software to reject wells with the flag.

   Note: After you have rejected the flagged wells, analysis results depend on factors such as the experiment type and flag type. For example, rejecting wells flagged by HIGHSD in experiments using the Standard Deviation calculations may change the result of C_T SD. For some flags, analysis results calculated before the well is rejected are maintained.
4. Click **Apply Analysis Settings** in the Analysis Settings dialog box. If the run status is complete, the data are reanalyzed.

The following is an image of the Flag Settings tab:



Relative Quantification Settings

Use the Relative Quantification Settings tab to:

- Change the reference sample and/or endogenous control. As the default, the QuantStudio™ 6 and 7 Flex Real-Time PCR Development Software assigns the endogenous control and reference sample for a study based on the analysis settings of the first experiment added to it. You can add multiple endogenous controls to a study.
- Correct the amplification efficiency. You can enter a percentage value between 1% and 150% for each target. When you set an assay to have a value that differs from 100% efficiency, the software uses the relative standard curve algorithm.
- (For multiplex reactions) Specify the ΔC_T value at which to reject replicates (outlier rejection).
- Select the algorithm to use to determine the relative quantification minimum and maximum values (error bars):
  - **Confidence Level** - Select to calculate the RQ minimum and maximum values based on the selected confidence level. Select the confidence level to use.
  - **Standard Deviations** - Select to calculate the RQ minimum and maximum values based on the selected number of standard deviations. Select the number of standard deviations to use.

Assess amplification results using the Amplification Plot

In the comparative C_T example study, you review each target in the Amplification Plot screen for correct baseline and threshold values.

1. From the Study Menu pane, select **Analysis ▸ Amplification Plot**.

   Note: If no data are displayed, click **Analyze**.

2. In the Experiment Data pane, select all of the experiments (click and drag to select all rows in the table).

3. In the Amplification Plot pane, set the parameters for the plot:

   a. From the Plot Type drop-down menu, select **ΔRn vs Cycle**.

   b. From the Plot Color drop-down menu, select **Well**.

   c. Click  **Show a legend for the plot**.

   Note: This is a toggle button. When the legend is displayed, the button changes to Hide the plot legend.

   d. From the Target drop-down menu, select **ACTB** to highlight all ACTB wells in the study.

4. View the baseline values:

   a. From the Graph Type drop-down menu, select **Linear**.

   b. Select the **Show Baseline** check box to show the start cycle and end cycle.

   c. Verify that the baseline is set correctly: The end cycle should be set a few cycles before the cycle number where significant fluorescence is detected. In the example study, the baseline is set correctly.

   Note: The data shown in the screenshot below is of the example experiment. Results vary depending on the experiment setup.

5. View the threshold values:

a. From the Graph Type drop-down menu, select **Log**.

b. Deselect the **Show Baseline** check box, then select the **Show Threshold** check box to show the threshold.

c. Verify that the threshold is set correctly. In the example study, the threshold is in the exponential phase.

6. Repeat steps 3 through 5 for the remaining targets.

Tips for assessing amplification in your own study

Ensure that your study meets the following requirements:

- **A typical amplification plot** – See the amplification plot examples in the Chapter 5 and Chapter 11.
- **Correct baseline and threshold values** – See the threshold examples and the baseline examples in Chapter 5 and Chapter 11.

If your study does not meet these requirements, you can:

- Manually adjust the baseline and/or threshold. See "View the analysis settings" on page 134.
- Omit individual wells from the analysis. See "Omit wells from the analysis" on page 164.

Assess the gene expression profile using the Gene Expression Plot

The Gene Expression Plot screen displays the results of the relative quantification calculations in the gene expression profile. Three plots are available:

- **RQ vs Target** – Groups the relative quantification (RQ) values by target. Each sample is plotted for each target. You can view the plot as the following graph types: linear, log10, Ln, log2.
- **RQ vs Sample** – (displayed only when the Technical Replicates tab is selected in the Replicate Results Data pane) – Groups the relative quantification (RQ) values by sample. Each target is plotted for each sample. You can view the plot as the following graph types: linear, log10, Ln, log2.
- **RQ vs BioGroup** – (displayed only when the Biological Replicates tab is selected in the Replicate Results Data pane) – Groups the relative quantification (RQ) values by biological replicate group. Each target is plotted for each sample. You can view the plot as the following graph types: linear, log10, Ln, log2.

About the example study

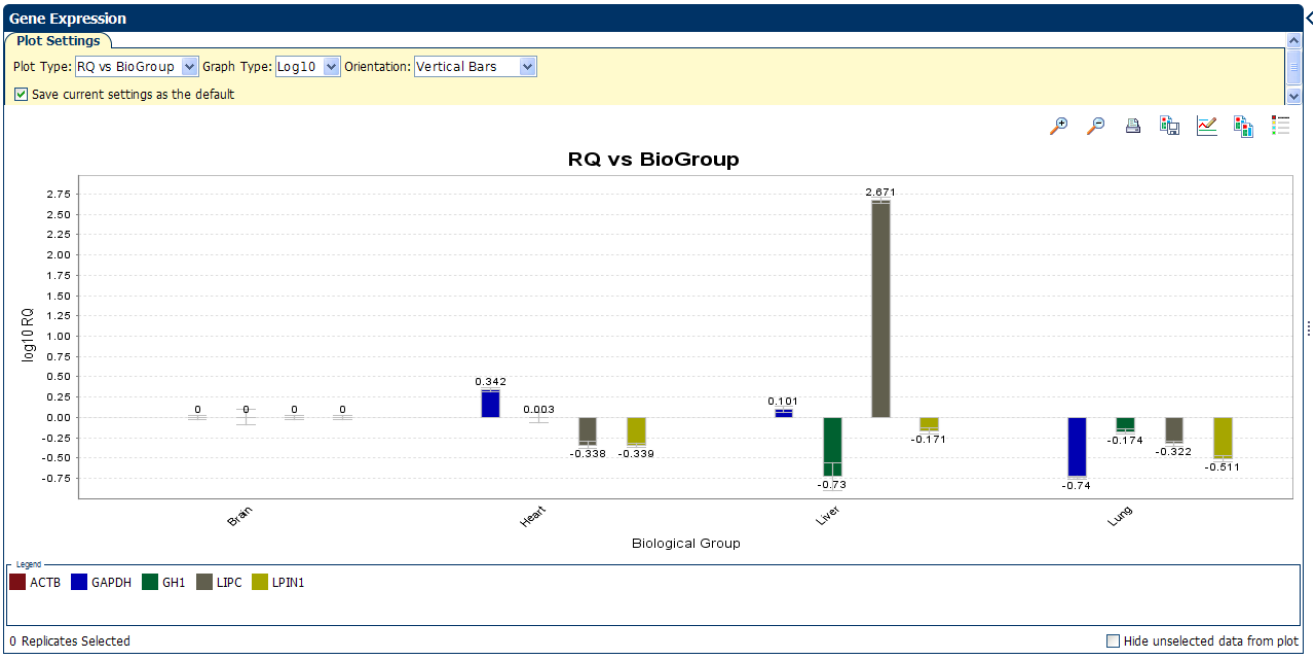
In the Comparative C_T example study, you review each target in the Gene Expression Plot screen for the expression level (or fold change) of the target sample relative to the reference biological replicate group.

View the Gene Expression Plot

1. From the Study Menu pane, select **Analysis ▸ Gene Expression**.
2. In the Gene Expression Plot pane, set the parameters for the plot:
   a. In the Plot Type drop-down menu, select **RQ vs BioGroup**.
   b. In the Graph Type drop-down menu, select **Log10**.
   c. In the Orientation drop-down menu, select **Vertical Bars**.
   d. Click  **Show a legend for the plot**.

      Note: This is a toggle button. When the legend is displayed, the button changes to Hide the plot legend.

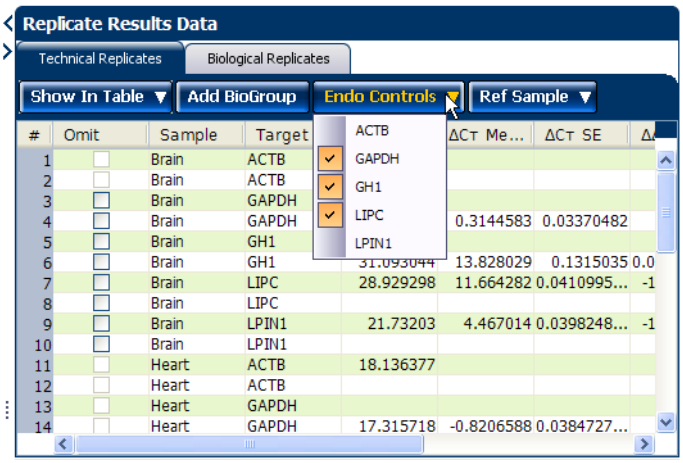
In the example study, the expression levels of multiple targets in the biological replicate group are displayed relative to the expression levels of the same targets in the reference biological replicate group (universal). Because the reference sample is compared to itself, the relative expression level is 1. When the result is displayed in the Log10 graph type, the expression level of the reference sample appears as 0 in the graph (log10 of 1 = 0).

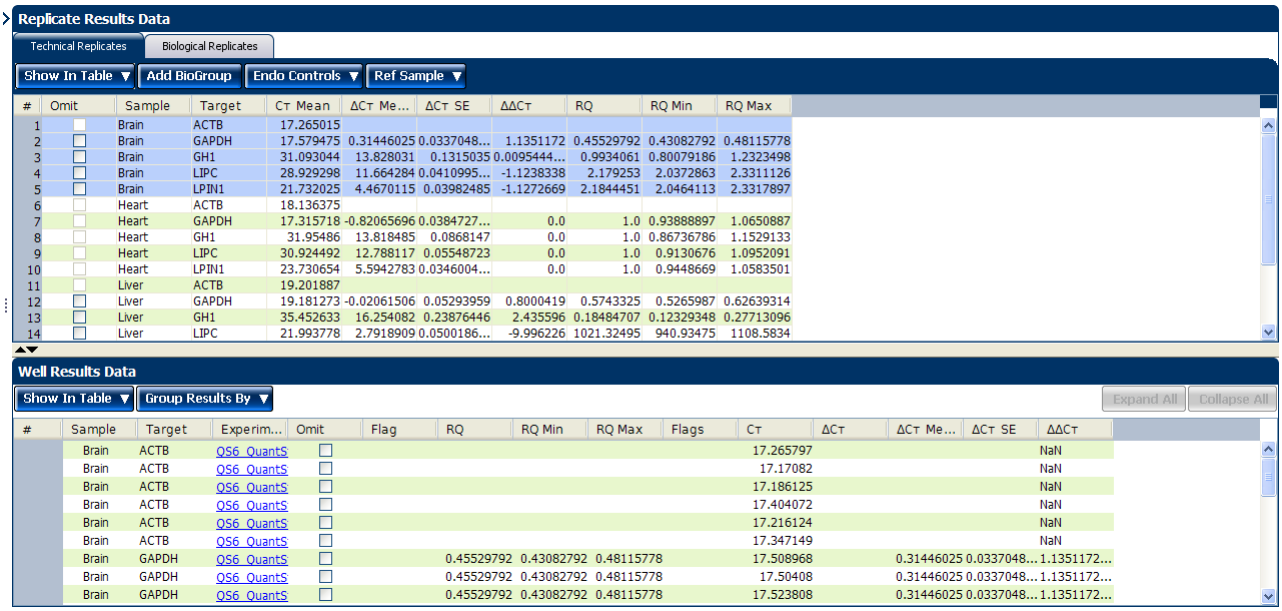


3. Select multiple endogenous controls:
    a. In the Replicate Results Data pane, select the **Technical Replicates** tab.

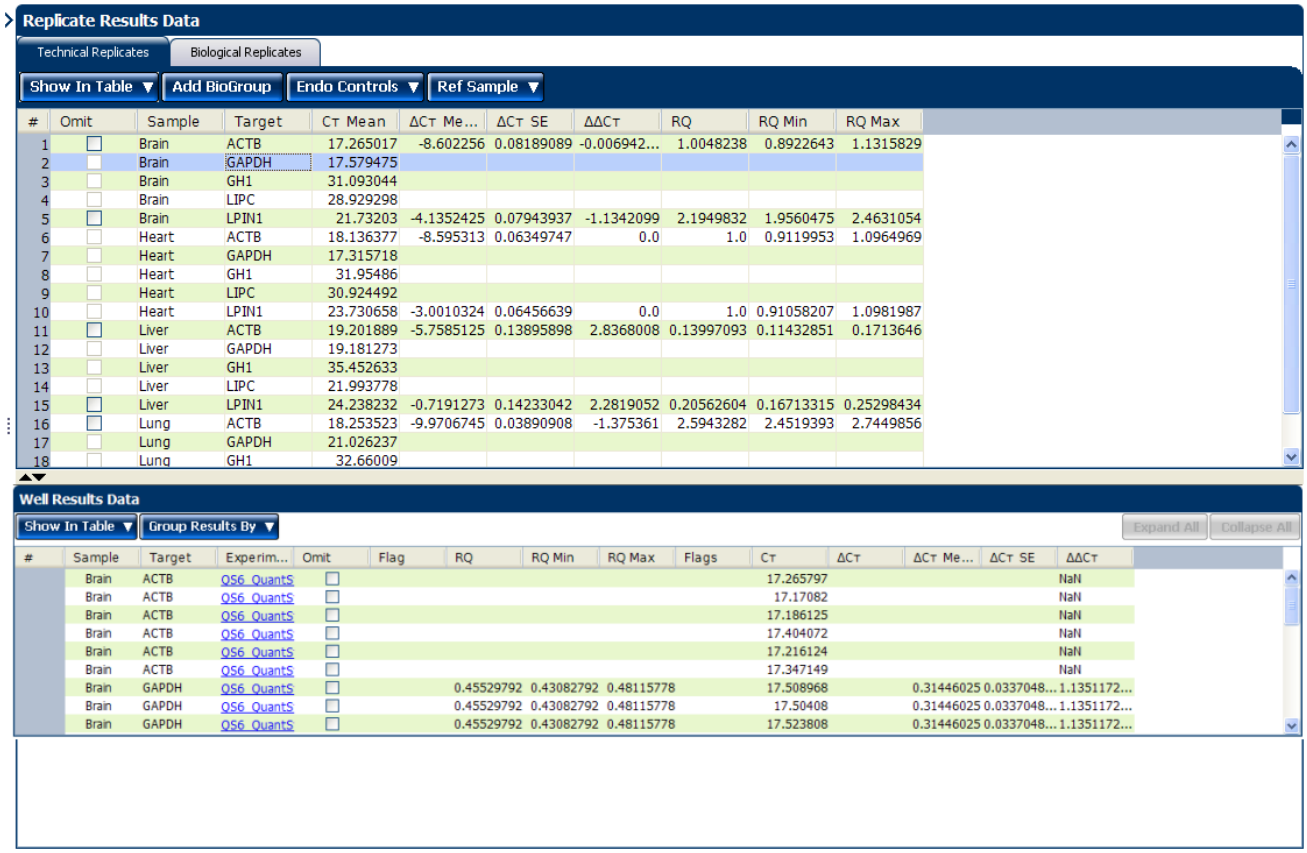
b. From the Endo Controls drop-down menu, select **GAPDH**, **GH1**, and **LIPC**, then deselect **ACTB**.

Note: You can also select the endogenous controls in the Analysis Settings dialog box. See "View the analysis settings" on page 134.





4. Click **Analyze**. In the example study shown below, all samples for the endogenous controls **GAPDH**, **ACTB**, and **LIPC** have no values (with the exception of the C_T Mean value), and the RQ values for the remaining samples change. For example, in the example study, the RQ value for the brain sample changes from 1.0 (with 18S as the endogenous control) to none with the new endogenous controls.



#	Omit	Sample	Target	CT Mean	ΔCT Me...	ΔCT SE	ΔΔCT	RQ	RQ Min	RQ Max
1	☐	Brain	ACTB	17.265017	-8.602256	0.08189089	-0.006942...	1.0048238	0.8922643	1.1315829
2		Brain	GAPDH	17.579475						
3		Brain	GH1	31.093044						
4		Brain	LIPC	28.929298						
5	☐	Brain	LPIN1	21.73203	-4.1352425	0.07943937	-1.1342099	2.1949832	1.9560475	2.4631054
6		Heart	ACTB	18.136377	-8.595313	0.06349747	0.0	1.0	0.9119953	1.0964969
7		Heart	GAPDH	17.315718						
8		Heart	GH1	31.95486						
9		Heart	LIPC	30.924492						
10		Heart	LPIN1	23.730658	-3.0010324	0.06456639	0.0	1.0	0.91058207	1.0981987
11	☐	Liver	ACTB	19.201889	-5.7585125	0.13895898	2.8368008	0.13997093	0.11432851	0.1713646
12		Liver	GAPDH	19.181273						
13		Liver	GH1	35.452633						
14		Liver	LIPC	21.993778						
15	☐	Liver	LPIN1	24.238232	-0.7191273	0.14233042	2.2819052	0.20562604	0.16713315	0.25298434
16	☐	Lung	ACTB	18.253523	-9.9706745	0.03890908	-1.375361	2.5943282	2.4519393	2.7449856
17		Lung	GAPDH	21.026237						
18		Lung	GH1	32.66009						

#	Sample	Target	Experim...	Omit	Flag	RQ	RQ Min	RQ Max	Flags	CT	ΔCT	ΔCT Me...	ΔCT SE	ΔΔCT
	Brain	ACTB	QS6_QuantS	☐						17.265797				NaN
	Brain	ACTB	QS6_QuantS	☐						17.17082				NaN
	Brain	ACTB	QS6_QuantS	☐						17.186125				NaN
	Brain	ACTB	QS6_QuantS	☐						17.404072				NaN
	Brain	ACTB	QS6_QuantS	☐						17.216124				NaN
	Brain	ACTB	QS6_QuantS	☐						17.347149				NaN
	Brain	GAPDH	QS6_QuantS	☐		0.45529792	0.43082792	0.48115778		17.508968		0.31446025	0.0337048...	1.1351172...
	Brain	GAPDH	QS6_QuantS	☐		0.45529792	0.43082792	0.48115778		17.50408		0.31446025	0.0337048...	1.1351172...
	Brain	GAPDH	QS6_QuantS	☐		0.45529792	0.43082792	0.48115778		17.523808		0.31446025	0.0337048...	1.1351172...

Tips for assessing the gene profile in your own study

Look for:

- Differences in gene expression (as a fold change) relative to the reference sample.
- Standard deviation in the replicate groups (C_T SD values).

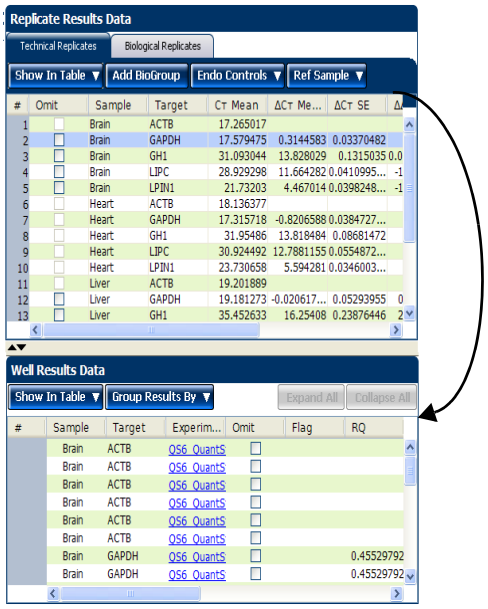
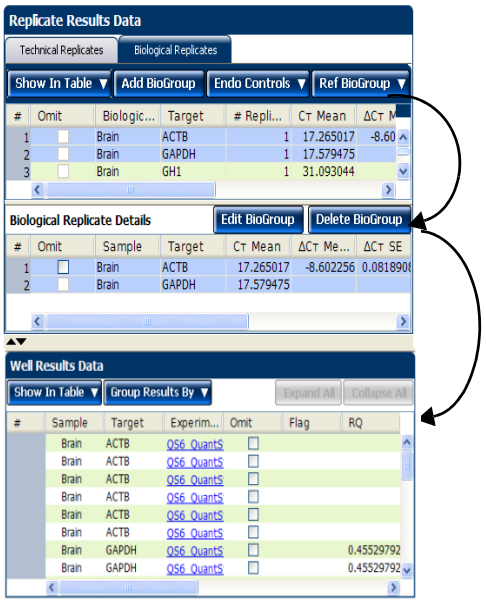
If needed, you can omit outliers. See "Omit replicates from the analysis" on page 147.

Note: To display a subset of the study data in the Gene Expression Plot pane, select one or more rows in the Technical Replicates tab or the Biological Replicates tab, then select **Hide unselected data from plot** to display data only from the selected rows.

View the replicate results data and the well results data

The Replicate Results Data pane lists each reaction plate (experiment) that is added to a study. The results of the study are arranged by technical or biological replicate association.

The data that are displayed in the Well Results Data pane depend on which tab you select in the Replicate Results Data pane:

Technical Replicates Tab	Biological Replicates Tab
This tab arranges the results of the relative quantification analysis by technical replicate group. The ViiA™ 7 Software displays the results for each sample/target combination as a row in the table. You can view the members of a technical replicate group by selecting the appropriate row in the table. When a row is selected, the Well Results Data pane displays the wells that make up the selected technical replicate group. 	This tab arranges the results of the relative quantification analysis by biological replicate group. The QuantStudio™ 6 and 7 Flex Software displays the results for each biological group as a row in the table (each row displays a biological sample with its target). You can view the members of a biological replicate group by selecting the appropriate row from the table. When a row is selected: • The Biological Replicate Details table displays the technical replicate groups that make up the selected biological replicate group. • The Well Results Data pane displays the individual members of the technical replicate groups that make up the selected biological replicate group. 

Column descriptions

The table below provides definitions for the column headings that appear in the tables in the Technical Replicates and Biological Replicates tabs.

Note: To show or hide columns in a table, select or deselect the column name from the Show In Table drop-down menu.

Column	Description
ΔC_T	The calculated ΔC_T value for the replicate group associated with the test sample. **Note:** The ΔC_T value is calculated only for multiplex experiments and is calculated at the well level (that is, the individual technical replicate level) by subtracting the target C_T value from the endogenous control $_{CT}$ value.
ΔC_T Mean	The arithmetic average of the technical replicate C_T values for the sample replicate group. **Note:** The ΔC_T Mean value is calculated at the reaction plate level and represents the mean difference between the target $_{CT}$ values and the endogenous control C_T values for all the technical replicates for that sample that are present on the plate.
ΔC_T SE	The Standard Error of the mean associated with the reported Mean ΔC_T value. **Note:** The ΔC_T SE value is calculated differently for multiplex and singleplex experiments. For multiplex experiments, the calculation is at the well level. For singleplex experiments, the calculation combines the plate-level $_{CT}$ value variation between the target and the endogenous control. **Note:** If you select the Standard Deviation option in the RQ Min/Max calculations on the Relative Quantification Settings in the Analysis Settings dialog box, ΔC_T SD, that is the Standard Deviation values are calculated by the QuantStudio™ 6 and 7 Flex Software.
$\Delta\Delta C_T$	The calculated $\Delta\Delta_{CT}$ value for the replicate group associated with the reference sample.
# Replicates	The number of biological replicate groups in the study.
Biological Group	The name of the biological replicate group.
C_T	Threshold cycle; the PCR cycle number at which the fluorescence meets the threshold in the amplification plot.
C_T Mean	The arithmetic average of the technical replicate $_{CT}$ values.
Experiment	The name of the experiment file (for example, heart.eds).
Flag	The number of QC flags that the well generated as listed in the ▲ symbol.
Omit (Replicate Results Data pane)	Indicates the omission status of the members of the associated technical or biological replicate group(s): • A check mark (✓) indicates that all replicates have been removed from the analysis. • A hyphen (–) indicates that one or more replicates have been removed from the analysis.
Omit (Well Results Data pane)	Indicates the omission status of the well. A check mark (✓) indicates that the well has been removed from the analysis.
RQ	The calculated relative level of gene expression for the replicate group that is associated with the test sample.

Column	Description
RQ Max	The maximum relative level of gene expression in the test samples calculated using the confidence level set in the Analysis Settings dialog box. **Note:** The maximum includes the variability associated with the endogenous control and targets in only the test samples.
RQ Min	The minimum relative level of gene expression in the test samples calculated using the confidence level set in the Analysis Settings dialog box. **Note:** The minimum includes the variability associated with the endogenous control and targets in only the test samples.
Sample	The sample associated with the data displayed in the row.
Target	The target assay associated with the data displayed in the row.
Well	The location of the well in the reaction plate.

About the example study

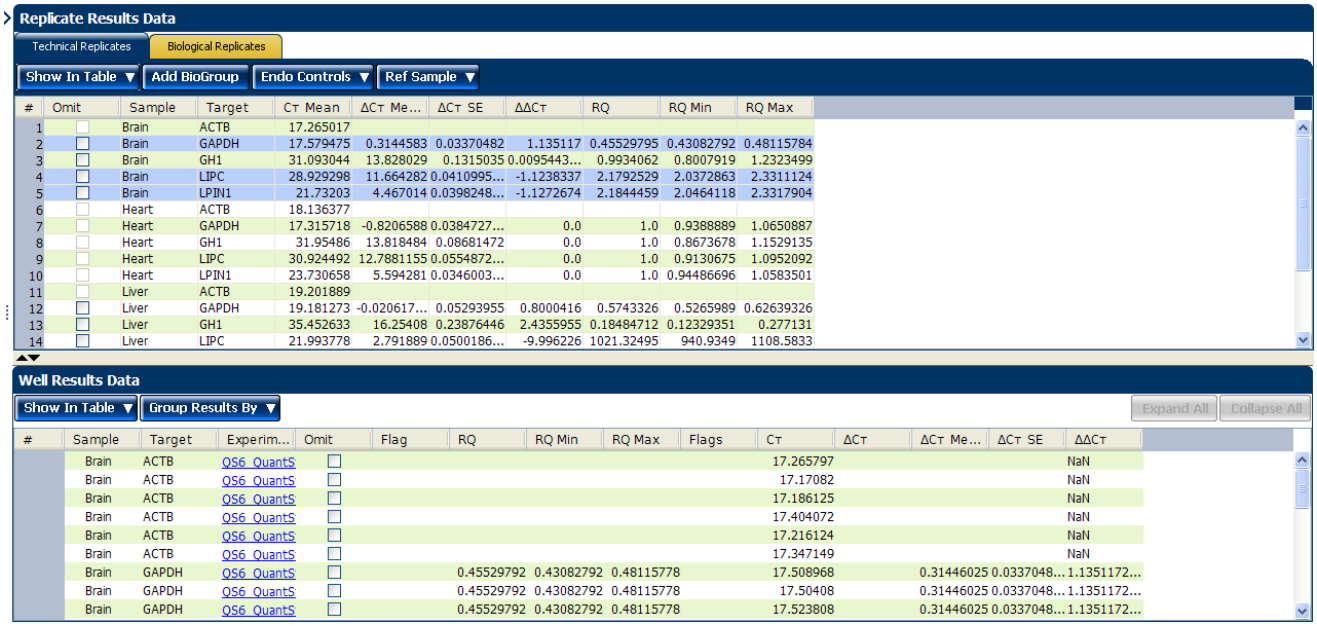
In the Comparative C_T example study, you review the Replicate Results Data pane and the Well Results Data pane to evaluate the C_T precision of the replicate groups and view related RQ information.

View the results data

1. From the Study Menu pane, select **Analysis ▸ Gene Expression**.
2. Click ⊴ at the top left of the Replicate Results Data pane.
3. View the technical replicates:
   a. Click the **Technical Replicates** tab. The table displays the results by technical replicate group.
   b. In the Technical Replicates table, select the following groups:
      - Brain/GAPDH (row2)
      - Brain/LIPC (row 4)
      - Brain/LPIN1 (row 5)

      The Well Results Data pane displays all wells that make up the selected groups.
4. View the values in the Well Results Data pane:
   a. From the Group Results By drop-down menu, select **Target**.

b. View the C_T, ΔC_T Mean, and ΔC_T SE values to evaluate the C_T precision of the replicate groups. In the example study, the low ΔC_T SE values indicate these replicates have good C_T precision.



Tips for viewing replicate results in your own study

- Select the **Technical Replicates** tab or the **Biological Replicates** tab to organize and view the sample data according to the associated technical replicate group or biological replicate group.
- View all wells for a technical or biological replicate group by selecting the appropriate row in the table. When a row is selected, the Well Results Data pane displays the wells that make up the group. **Ctrl-click** to select multiple rows.
- Change the endogenous control by clicking **Endo Control**, then selecting a new target.
- Change the reference sample by clicking **Ref Sample**, then selecting a new sample.
- Add biological replicate groups by clicking **Add BioGroup**.
- Omit biological or technical replicates from the analysis. See "Omit replicates from the analysis" on page 147.

  Note: The QuantStudio 384 Well Comparative Ct with Bioreplicates Study Example.edm file demonstrates the use of biological replicate groups. You can also create example study that does not use biological replicate groups.

Omit replicates from the analysis

To omit a technical or biological replicate from the analysis:

1. From the Study Menu pane, select **Analysis ▸ Gene Expression**.
2. Select the **Technical Replicates** or **Biological Replicates** tab according to the type of replicate that you want to omit.
3. In the replicate table, scroll to the biological or technical replicate of interest, then select the corresponding check box in the Omit column.

4. Click **Analyze** when you finish omitting wells.

IMPORTANT! You cannot omit *all* technical replicates that belong to a reference sample or a reference biological group, or that serve as the endogenous control for a study.

Note: You can also omit the biological replicates in the Biological Replicate Details table at the bottom of the Biological Replicates tab.

Confirm accurate dye signal using the Multicomponent Plot

The Multicomponent Plot screen displays the complete spectral contribution of each dye over the duration of the PCR run in a selected well of any experiment that is added to the study.

About the example study

In the comparative C_T example study, you review the Multicomponent Plot screen for:

- ROX™ dye (passive reference)
- FAM™ dye (reporter)
- Spikes, dips, and/or sudden changes
- Amplification in the negative control wells

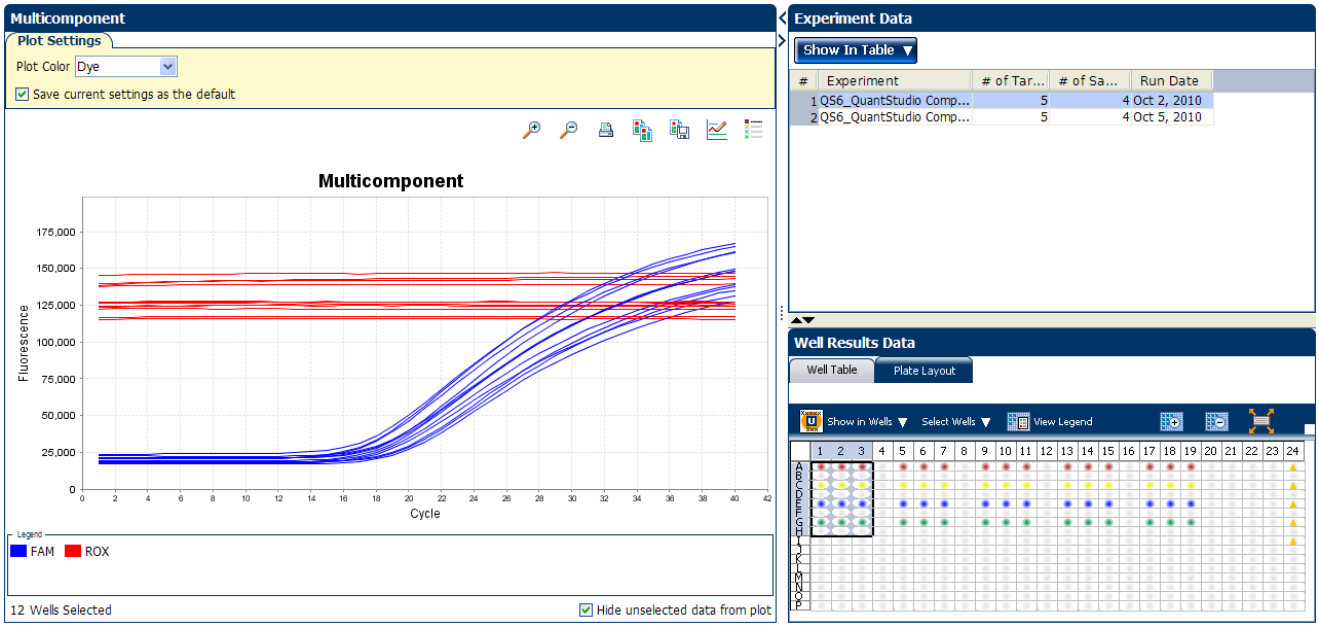
View the Multicomponent Plot

1. From the Study Menu pane, select **Analysis ▸ Multicomponent Plot**.
2. In the Experiment Data pane, select the **QuantStudio Comparative Ct Example 1.eds** experiment.
3. Display the unknown wells one at a time in the Multicomponent Plot pane:
   a. Click the **Plate Layout** tab.
   b. Select one well in the plate layout; the well is shown in the Multicomponent Plot pane.

   Note: If you select multiple wells, the Multicomponent Plot pane displays the data for all selected wells simultaneously.
4. In the Multicomponent Plot pane, set the parameters for the plot:
   a. From the Plot Color drop-down menu, select **Dye**.
   b. Click  **Show a legend for the plot**.

   Note: This is a toggle button. When the legend is displayed, the button changes to Hide the plot legend.
5. Check the FAM dye signal. In the example study, the FAM dye signal increases throughout the PCR process, which indicates normal amplification.
6. Check the ROX dye signal. In the example study, the ROX dye signal remains constant throughout the PCR process, which indicates typical data.

7. Repeat steps 2 through 6 for the remaining experiments in the study.



Tips for confirming dye-signal accuracy in your own studies

- **Passive reference** – The passive reference dye fluorescence level should remain relatively constant throughout the PCR process.
- **Reporter dye** – The reporter dye fluorescence level should display a flat region corresponding to the baseline, followed by a rapid rise in fluorescence as the amplification proceeds.
- **Irregularities in the fluorescence** – There should not be any spikes, dips, and/or sudden changes in the fluorescence.
- **Negative control wells** – There should not be any amplification in the negative control wells.

If your study does not meet the guidelines above, you can omit individual wells from the analysis. See “Omit wells from the analysis” on page 164.

View the QC Plots

The QC Plots screen displays the endogenous control and replicate analysis results for each reaction plate (experiment) added to the study. The following plots are derived from the experiments added to a study:

- Endogenous Control Profile
- Box Plot
- Technical Replicates Correlation
- Biological Replicates Correlation

View the Endogenous Control Profile plot

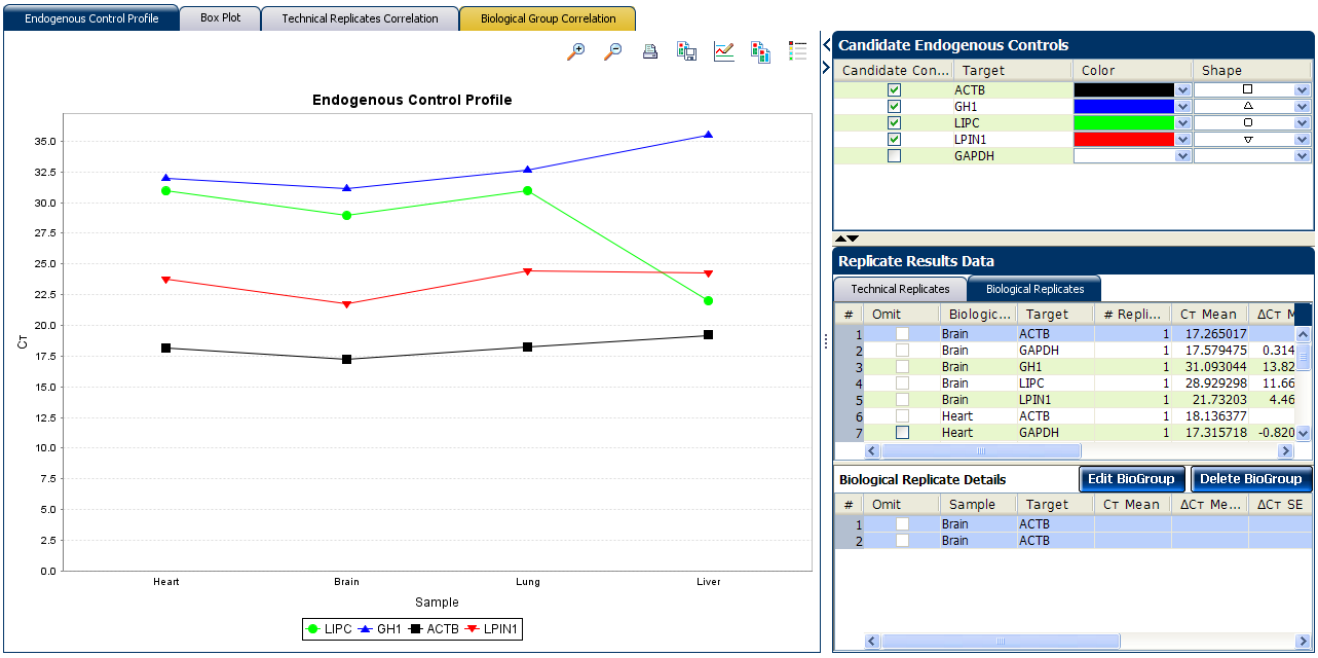
The QC Plots screen displays the Endogenous Control Profile plot for the endogenous controls used in the experiments added to a gene expression study. The endogenous control profile plot displays how much of the endogenous control is expressed in a sample. The sample is plotted on the X-axis, and the C_T is plotted on the Y-axis. The expression is viewed as a color and shape combination in the plot.

In the example study, you view four potential endogenous controls expressed in four samples. The potential endogenous controls are:

- ACTB
- GH1
- LIPC
- LPIN1

The samples are brain, heart, lung, and liver.

1. From the Study Menu pane, select **Analysis** ▸ **QC Plots**.
2. In the QC Plots pane, click **Endogenous Control Profile**.
3. In the **Candidate Endogenous Controls** pane, select the check boxes of those Targets whose profile you want to view in the plot pane. In the example study, the **Target** ACTB is chosen to be the endogenous control because it is expressed at similar levels in three out of four of the given samples.
4. In the Replicate Results Data pane, view results by replicate group.
   a. Click the **Technical Replicates** tab. The table displays the results by technical replicate group.
   b. Click the **Biological Replicates** tab. The table displays the results by biological replicate group.



View the box plots

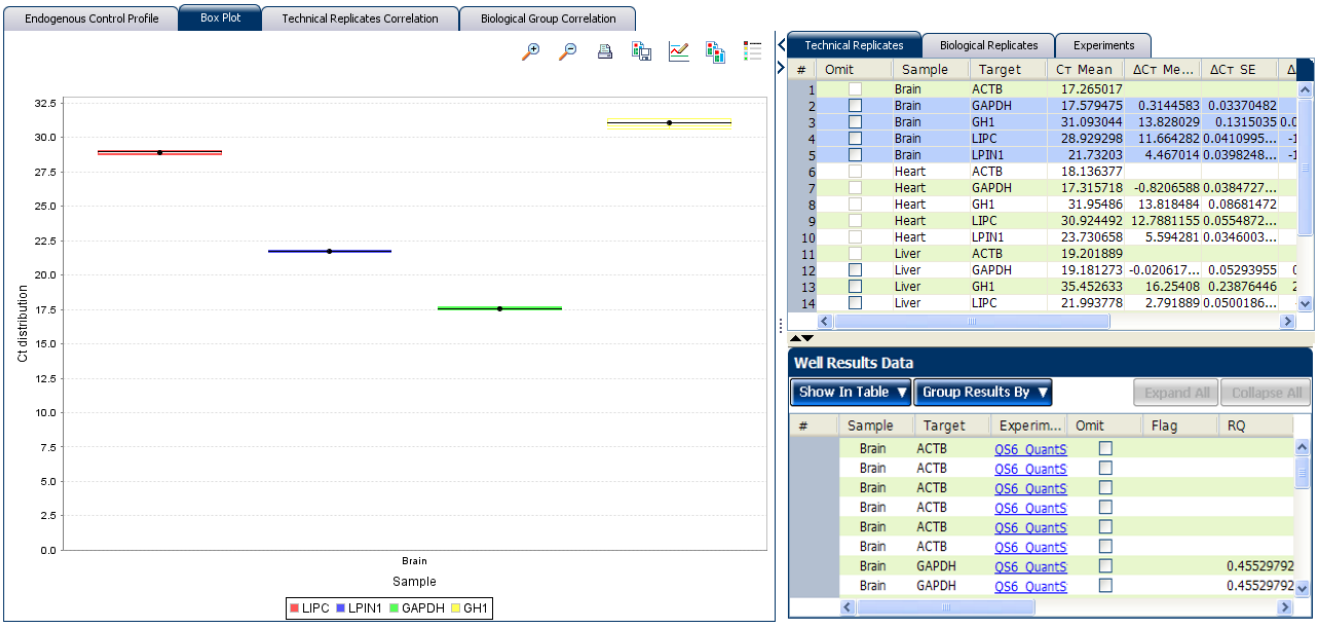
The Box Plot displays the C_T distribution of a particular **Target** in samples. You can see the individual C_T values/ raw data with this plot.

In the example study, you view the box plots of five targets in four different samples:

- LPIN1
- GAPDH
- LIPC
- GH1
- ACTB

The samples are Heart, Liver, Lung, and Brain.

1. In the QC Plots pane, click **Box Plots** to access the Replicate Results Data pane.
2. In the Replicate Results Data pane, click the **Technical Replicates** tab. The table displays the results by technical replicate group.
3. Click the **Biological Replicates** tab. The table displays the results by biological replicate group.
4. Click the Experiments tab to select the experiment whose Box plot to view.
5. View the values in the Well Results Data pane. View the C_T, ΔC_T Mean, and ΔC_T SE values to evaluate the C_T precision of the replicate groups. In the example study, the low ΔC_T SE values indicate these replicates have good C_T precision.



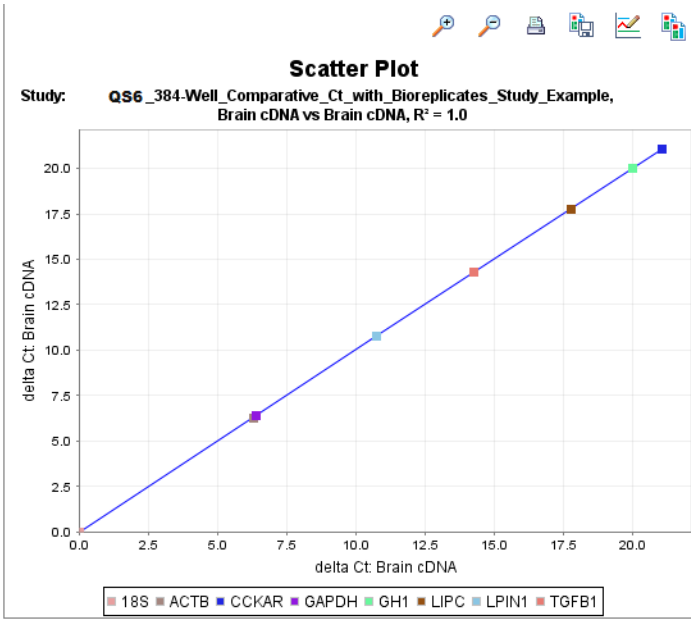
View the Technical Replicates Correlation plot

The Technical Replicates Correlation plot displays the correlation between the target genes in one or more samples.

The Technical Correlation Group plot is made of two components, the scatter plot and the heat map.

Scatter Plot

The scatter plot shows the distribution of ΔC_T of targets for different samples.

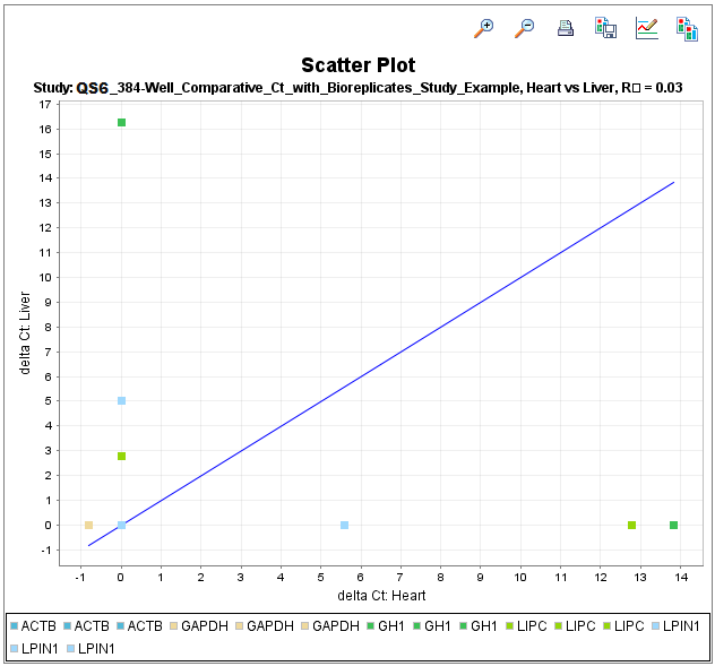


If a correlation (represented by R^2 or the coefficient of determination) exists among the targets in the samples, the targets appear on or along the line of reference. If the correlation is weak or not present, the targets appear scattered in the plot, away from the line of reference.

The line of reference is fixed in the QuantStudio™ 6 and 7 Flex Real-Time PCR Development Software.

If:

- $R^2 = / \approx 1$, then the correlation is strong
- $R^2 < 1$, then the correlation is weak
- $R^2 = 0$, then there is no correlation



Heat maps

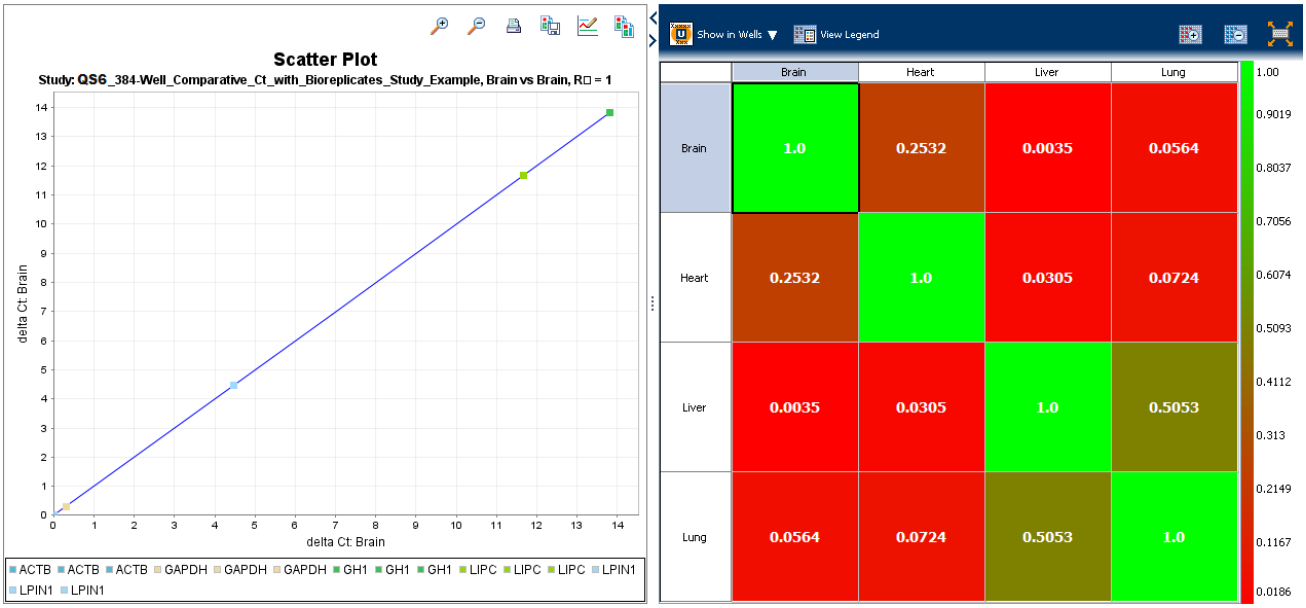
The heat map shows the variation of coefficient determination for different scatter plots. Each cell of the heat map represents a different scatter plot, and therefore a different value for R^2. The cells inclined to red represent a lower R^2 value; the cells inclined to green represent a higher R^2 value.

In the example study, you view the scatter plots and heat maps of five targets in four different samples:

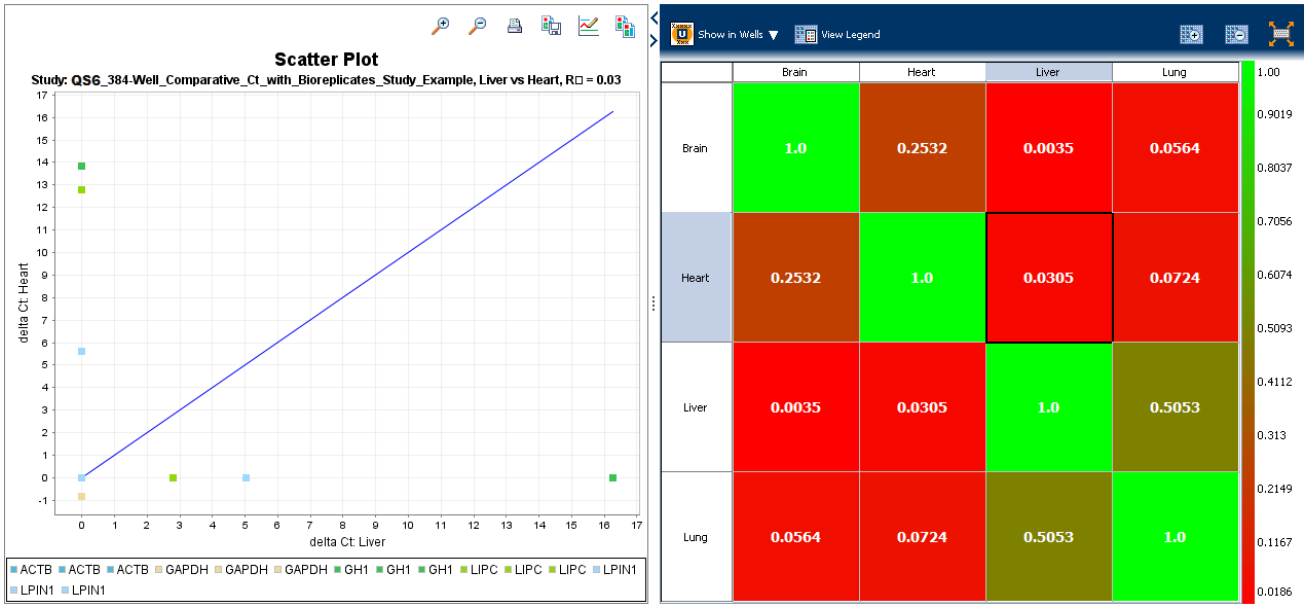
- LPIN1
- GAPDH
- LIPC
- GH1
- ACTB

The samples are Brain, Heart, Kidney and Lung.

1. In the QC Plots pane, click **Technical Replicates Correlation** to access the Heat Map.
2. In the Heat Map, click the cell with a correlation value, R^2 =/≈ 1. The corresponding scatter plot pane displays the scatter plot of the targets in that technical replicate group along the line of reference.



3. Click the cell with a correlation value, R^2 =/≈ 0. The corresponding scatter plot pane displays the scatter plot of the targets in that technical replicate group away from the line of reference.



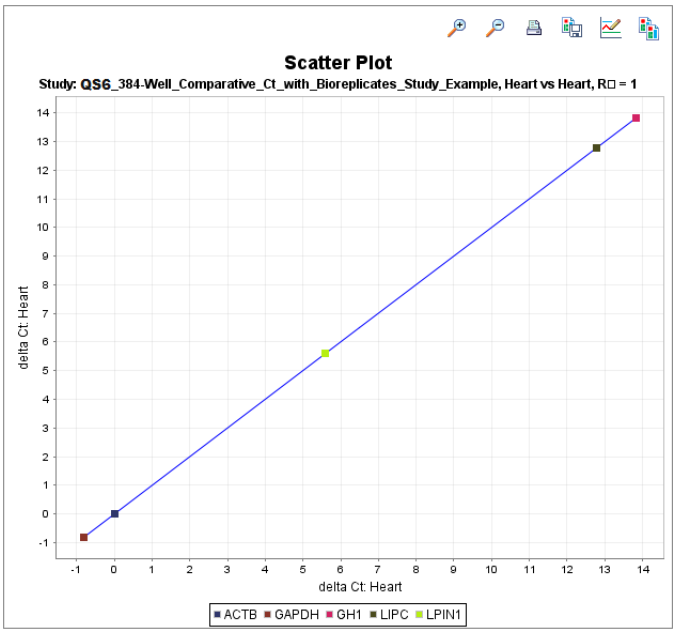
View the Biological Group Correlation plot

The Biological Group Correlation plot displays the correlation between the target genes in one or more biological group samples. Biological groups provide a broader set of samples, with the same targets.

Note: If the experiments in your study do not use biological replicate groups, see "Define Replicates" on page 128 to create a new biological group.

The Biological Group Correlation plot is also made of two components, the scatter plot and the heat map.

Scatter Plot

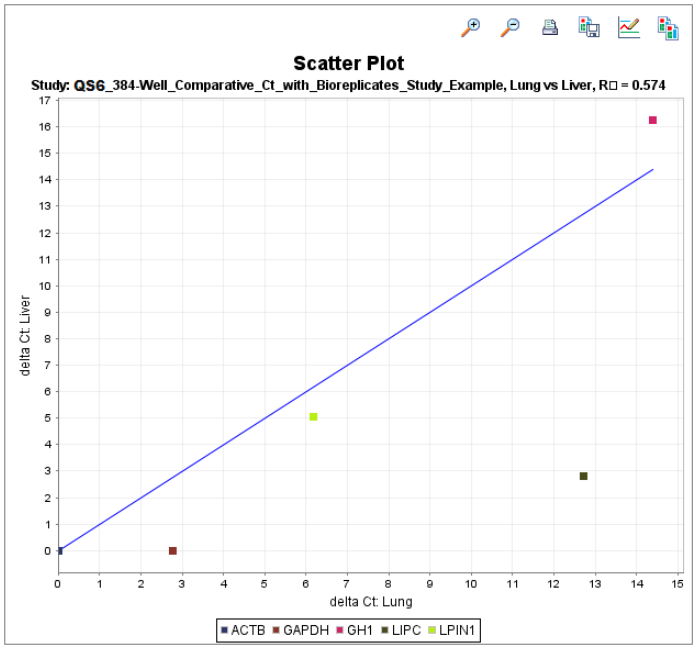


The scatter plot shows the distribution of ΔC_T of targets for different biological groups. If a correlation (represented by R^2 or the coefficient of determination) exists among the targets in the biological groups, the targets appear on or along the line of reference. A weak correlation or no correlation is represented by the targets being scattered in the plot and away from the line of reference.

Note: The line of reference is fixed in the QuantStudio™ 6 and 7 Flex Real-Time PCR Development Software.

If:

- $R^2 = / \approx 1$, then the correlation is strong
- $R^2 < 1$, then the correlation is weak
- $R^2 = 0$, then there is no correlation



Heat maps

The heat map shows the variation of coefficient determination for different scatter plots. Each cell of the heat map represents a different scatter plot, and therefore a different value for R^2. The dull green cells represent a lower R^2 value; the bright green cells represent a higher R^2 value.

In the example study, you view the scatter plots and heat maps of eight targets across four biological groups:

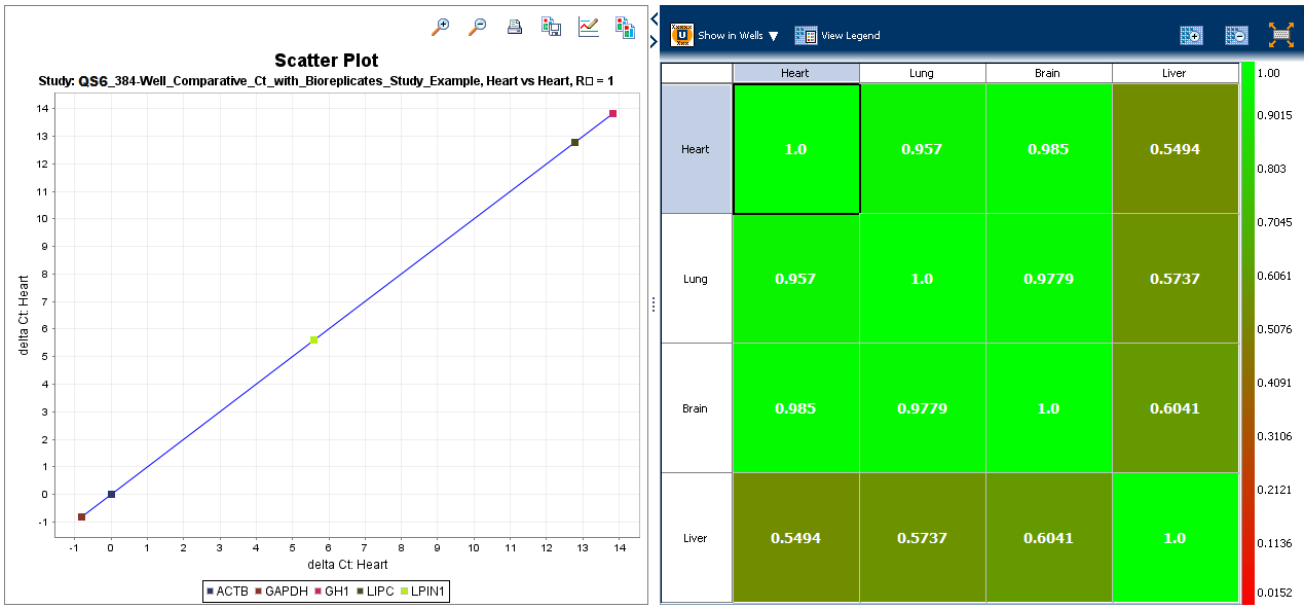
- LPIN1
- GAPDH
- LIPC
- GH1
- ACTB

Technical replicate samples of Brain cDNA belong to the biological group Brain, those of Lung cDNA belong to biological group Lung, technical replicate samples of Liver cDNA belong to the biological group Liver, and those of Heart cDNA belong to the biological group Heart.

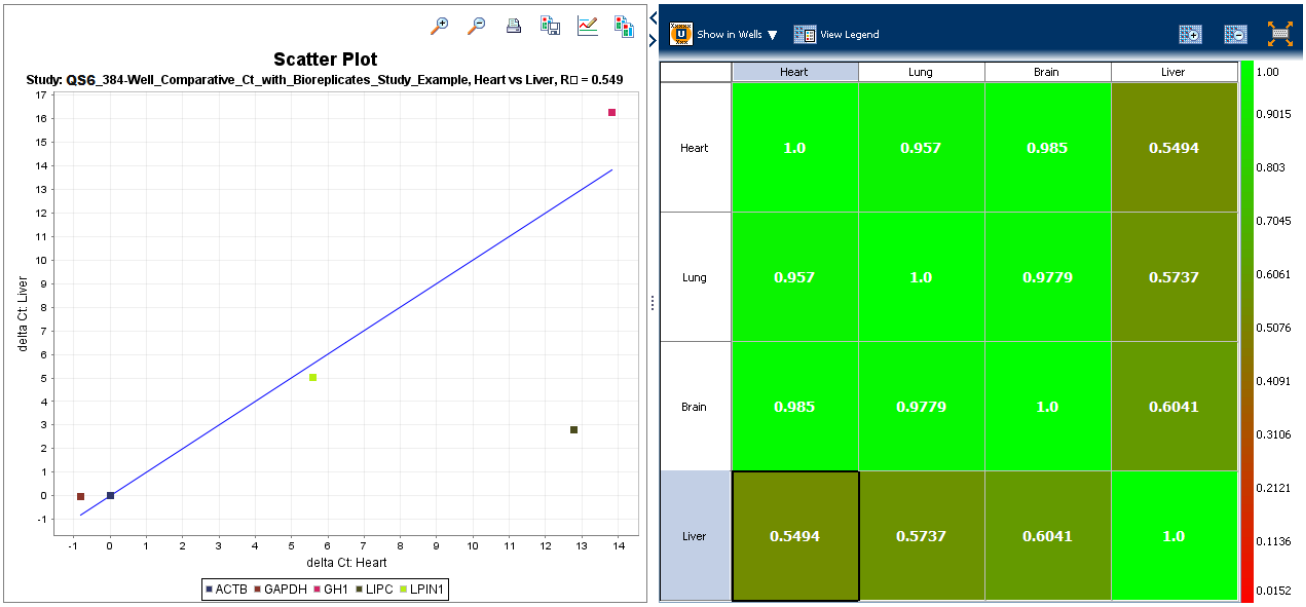
To view the Biological Group Correlation plot:

1. In the QC Plots pane, click **Biological Group Correlation** to access the heat map.

2. In the Heat Map, click the cell with a correlation value, R^2 =/≈ 1. The corresponding scatter plot pane displays the scatter plot of the targets in that biological replicate group along the line of reference.



3. Click the cell with a correlation value, R^2 =/≈ 0. The corresponding scatter plot pane displays the scatter plot of the targets in that biological replicate group away from the line of reference.



Tips for viewing your own scatter plots and heat maps

When you analyze your study, look for scatter plots and heat maps that display a correlation value that is =/≈ 1 for samples that come from the same source or tissue (technical or biological replicates). If the replicates do not correlate well, that could be a sign that there is a problem with a sample.

If your study does not meet the guidelines above, you can omit individual wells from the analysis. See "Omit wells from the analysis" on page 164.

View the QC Summary

The QC Summary screen displays a list of the QuantStudio™ 6 and 7 Flex Real-Time PCR Development Software flags, and it includes the flag frequency and location for any experiment that is added to a study.

About the example study

In the Comparative C_T example study, you review the QC Summary screen for any flags generated by the study data. In the example study, several wells produced data that generated flags.

View the QC Summary

1. From the Study Menu pane, select **Analysis ▸ QC Summary**.

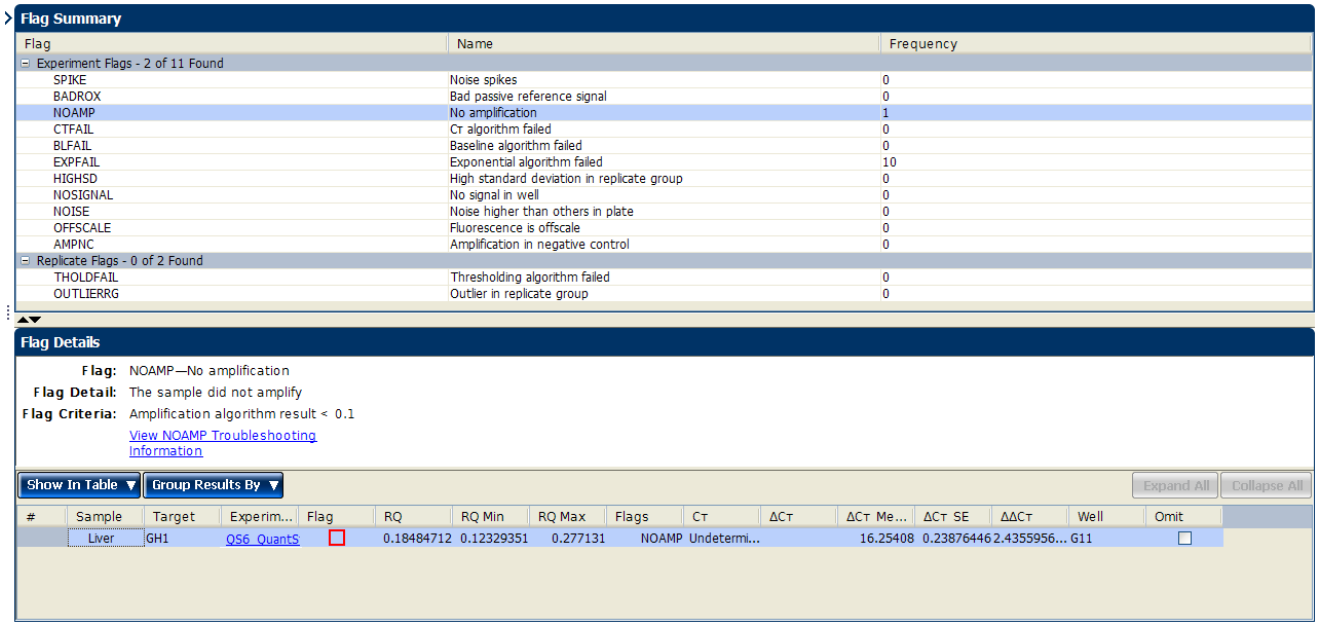
   Note: If no data are displayed, click **Analyze**.

2. In the Flags Summary table, look in the Frequency column to determine which flags appear in the study. In the example study. The EXPFAIL flag appears 10 times and the NOAMP flag appears once.

   Note: A 0 displayed in the Frequency column indicates that the flag does not appear in the study.

3. For each flag that appears in the study, click the flag row to display details about the flag in the Flag Details table. In the example study, the NOAMP flag indicates no amplification and the EXPFAIL flag indicates that the exponential algorithm failed.

4. Consider removing the NOAMP well from the analysis. See "Omit wells from the analysis" on page 164.



Possible flags

For Comparative C_T studies, the flags listed below may be generated by the study data.

If a flag does not appear in the study, its frequency is 0. If the frequency is >0, the flag appears somewhere in the study, and the associated well position is listed in the Wells column.

Flag	Description
Pre-processing flag	
OFFSCALE	Fluorescence is offscale
Primary analysis flags	
BADROX	Bad passive reference signal
NOAMP	No amplification
NOISE	Noise higher than others in plate
SPIKE	Noise spikes
NOSIGNAL	No signal in well
EXPFAIL	Exponential algorithm failed
BLFAIL	Baseline algorithm failed
THOLDFAIL	Thresholding algorithm failed
CTFAIL	C_T algorithm failed
Secondary analysis flags	
OUTLIERRG	Outlier in replicate group
AMPNC	Amplification in negative control
HIGHSD	High standard deviation in replicate group

Tips for using flags to evaluate your study

- In the Flag Summary table, click each flag that has a frequency >0 to display details about the flag in the Flag Details table. If needed, click the troubleshooting link in the Flag Details table to view information on correcting the flag.

  Note: In the Flag Details table, the numbers on each flag symbol indicate the number of flags generated for that well. For example, indicates that two flags have been generated for that well.

- You can change the flag settings. For more information, see "Flag Settings" on page 136:
  - Adjust the sensitivity so that more wells or fewer wells are flagged.
  - Change the flags that are applied by the QuantStudio™ 6 and 7 Flex Real-Time PCR Development Software.
- You can omit individual wells from the analysis. See "Omit wells from the analysis" on page 164.

Compare analysis settings

Use the Compare Settings screen to perform a side-by-side comparison of analysis settings for a comparative $_{CT}$ study. You can change one or more of the analysis settings, then compare the new results with the previous results. For example, you can compare the effects of:

- Using multiple endogenous controls versus a single endogenous control.
- Changing the amplification efficiency of a specific target versus keeping it at 100%.

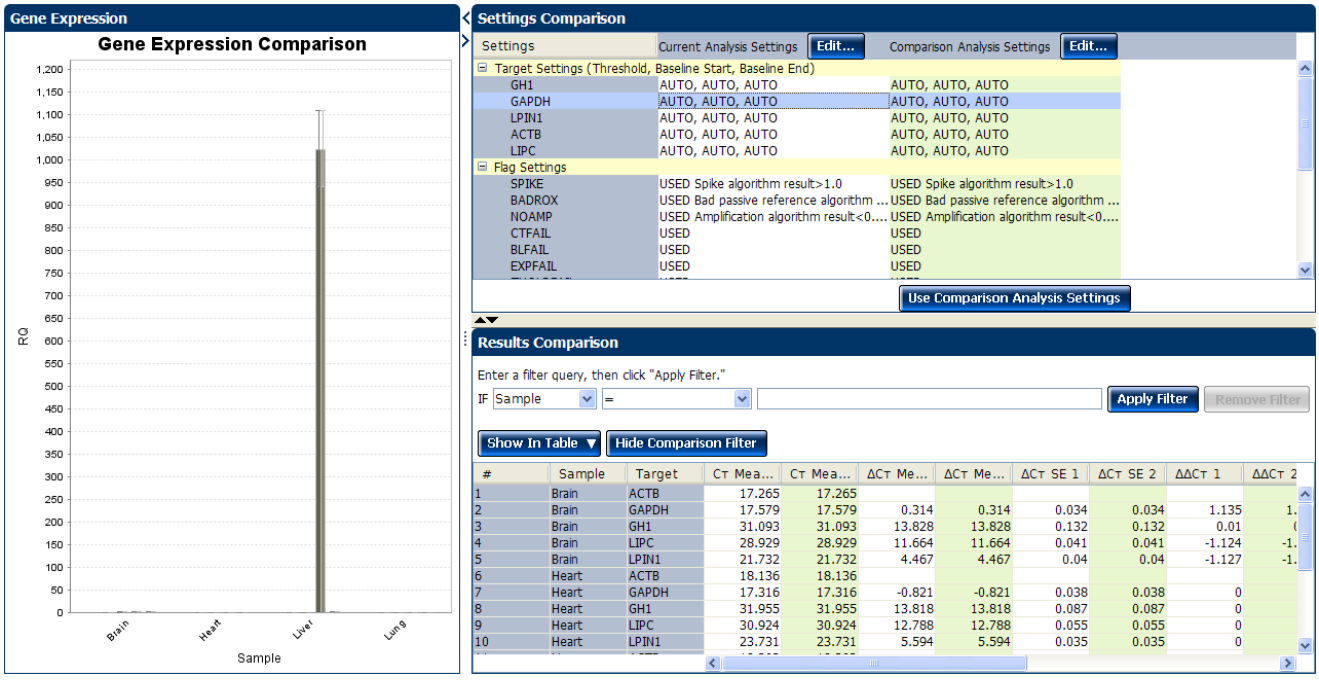
About the example study

In the comparative C_T example study, you change the endogenous control to LPIN1, then compare results.

Modify comparison criteria

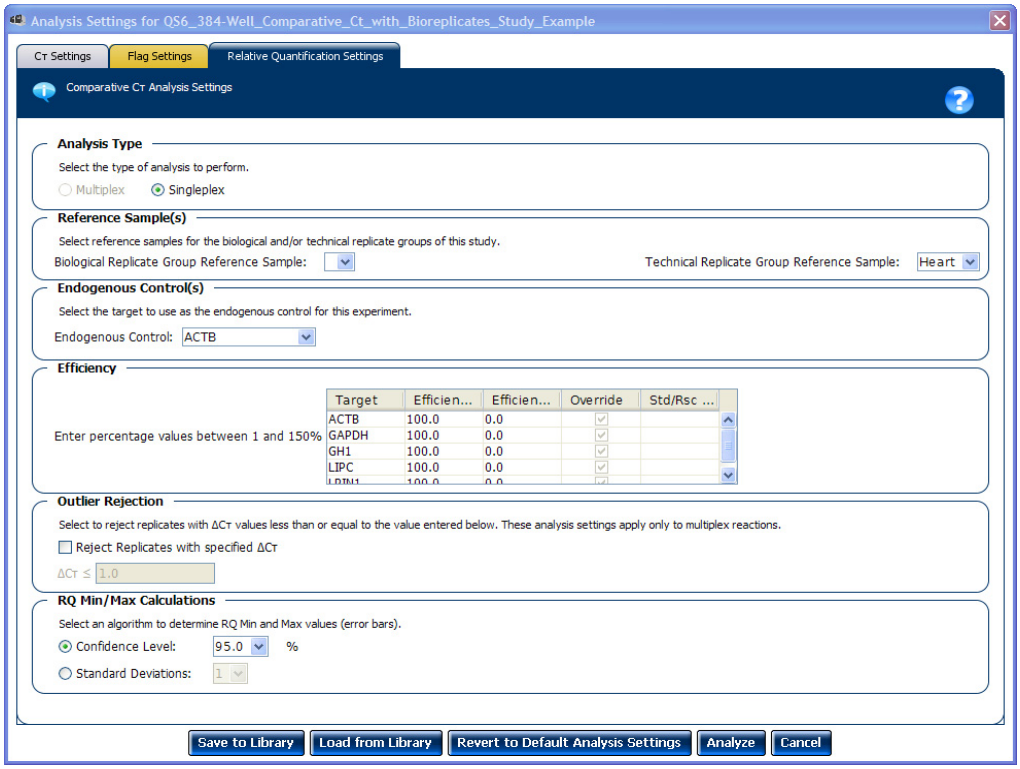
1. From the Study Menu pane, select **Analysis ▸ Compare Settings**. When the Compare Settings screen is initially displayed (before you make any changes):
   - In the Settings Comparison pane, the green column is titled "Current Analysis Settings," the white column is titled "Comparison Analysis Settings," and the Use Comparison Analysis Settings button is under the green column.
   - In the Results Comparison pane, values in the white columns and values in the green columns are the same.
   - The Gene Expression Comparison plot is the same plot that is displayed in the Gene Expression screen (**Analysis ▸ Gene Expression**).

2. In the Settings Comparison pane, click **Edit** in the green column to open the Comparison Analysis Settings dialog box.



3. In the Comparison Analysis Settings dialog box, change the endogenous control:
   a. Select the **Relative Quantification Settings** tab.
   b. In the Endogenous Control(s) pane, select **LIPN1** from the Endogenous Control drop-down menu.

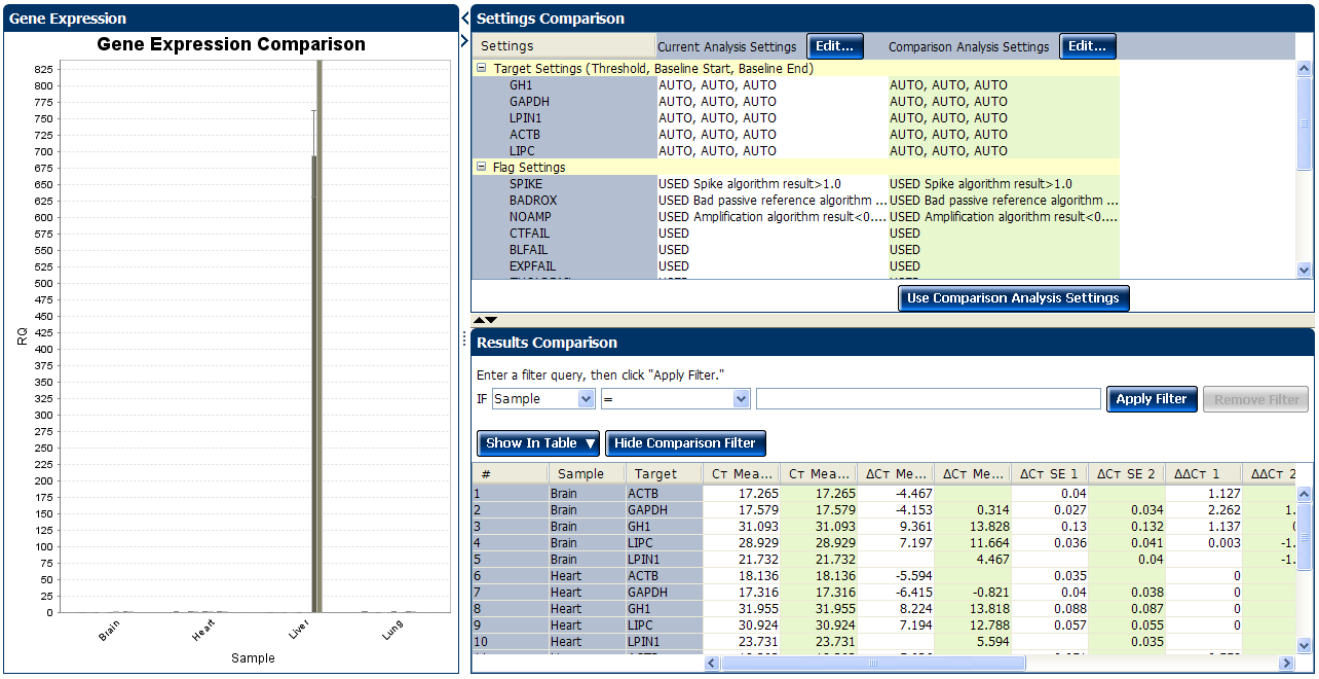
c. Click **Analyze** to analyze the data and close the dialog box.



4. In the Settings Comparison pane, click **Use Comparison Analysis Settings** under the green column, then compare the results:
   - In the Settings Comparison pane, the white column is titled "Comparison Analysis Settings," the green column is titled "Current Analysis Settings," and the Use Comparison Analysis Settings button is under the white column.
   - In the Results Comparison pane, values in the white columns are based on the default analysis settings, and values in the green columns are based on the modified analysis settings. In the example experiment, changing the endogenous control to 18S affects the RQ values. To view the RQ values, scroll to the left. If desired, you can click and drag the RQ column headings so that they appear first in the table.
   - The Gene Expression Comparison plot displays the default analysis settings (the values from the white columns).

     Note: The Gene Expression Comparison plot has limited functions. For example, you cannot change to log scale and you cannot view by target.

Note: The default analysis settings are the settings automatically made by the software when the study is initially analyzed.



5. From the Study Menu pane, select **Analysis ▸ Gene Expression** to view the gene expression plot using the modified analysis settings.

   Note: In the Gene Expression screen you can view the modified data in log scale, by target, and so on. See "Assess the gene expression profile using the Gene Expression Plot" on page 140.

6. *(Optional)* View the data in the other analysis screens. All other analysis screens for the study display the data using the modified analysis settings.

7. Close the study.

   - Save your changes before closing the study.

     Or

   - Close the study without saving your changes. If you do not save your changes, the software reverts to the default analysis settings the next time you open the study.

Tips for managing your own comparison

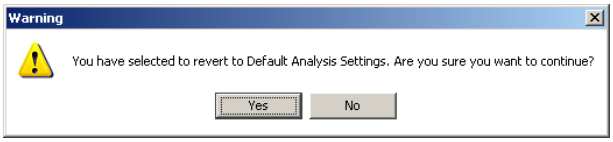
- Edit the comparison analysis settings as desired. For information on editing the settings, see "View the analysis settings" on page 134.
- After making your first round of changes to the analysis settings, you can continue making changes using one of the following methods:
  - (Recommended) Revert to the saved analysis settings, then make new changes. To do this: In the Settings Comparison pane, click **Use Comparative Analysis Settings** (now under the white column) to revert to the saved analysis settings, then repeat steps 2 through 6 above. This method ensures that you do not lose the saved analysis settings.

    Note: If you have made changes, but have not saved them, the software reverts to the default analysis settings when you click **Use Comparative Analysis Settings**. The default analysis settings are the settings automatically made by the software when the study is initially analyzed.
  - Continually compare new settings with previous settings. To do this: In the Settings Comparison pane, alternate clicking **Edit** in the white and green columns, then repeat steps 3 through 6 above. This method does not allow you to return to your saved settings; subsequent comparisons are made with the previous analysis settings, building upon any changes that you have already made.

Revert to the default analysis settings

IMPORTANT! The default analysis settings are defined by the software. If you make changes to the analysis settings and save the study, the saved changes are lost when you revert to the default analysis settings.

1. In the Settings Comparison pane, click **Edit** next to the settings you want to revert to the default: *Current Analysis Settings* or *Comparison Analysis Settings*.
2. In the Analysis Settings dialog box, revert to defaults and reanalyze the data:
   a. Click **Revert to Default Analysis Settings**.
   b. At the prompt, click **Yes**.



   c. Click **Analyze** to analyze the data and close the dialog box.
3. In the Settings Comparison pane, click **Use Comparison Analysis Settings**. In the Results Comparison pane, values for the settings you selected to edit in step 1 ("Current Analysis Settings" or "Comparison Analysis Settings") are generated according to the default analysis settings.

Omit wells from the analysis

You can use the Well Table to omit individual wells from the analysis. To omit a well:

1. From the Study Menu pane, select one of the following analysis screens:
   - **Analysis ▸ Amplification Plot**
   - **Analysis ▸ Multicomponent Plot**
   - **Analysis ▸ Multiple Plots View**
2. In the Experiment Data pane, select the experiment that contains the well to omit.

3. In the Well Results Data pane, click the **Well Table** tab, then select the check box in the Omit column for the well to omit.

   Note: You cannot omit all technical replicates that belong to a reference sample or a reference biological group, or that serve as the endogenous control.

Export the study

Note: If you are using RealTime StatMiner® Software to analyze the exported data, make sure you have assigned a sample to all the wells containing data in the individual experiments. If a sample is not assigned, the RealTime StatMiner® Software reports an error during import.

You can export the data within a study from the Analysis screen. To export a study:

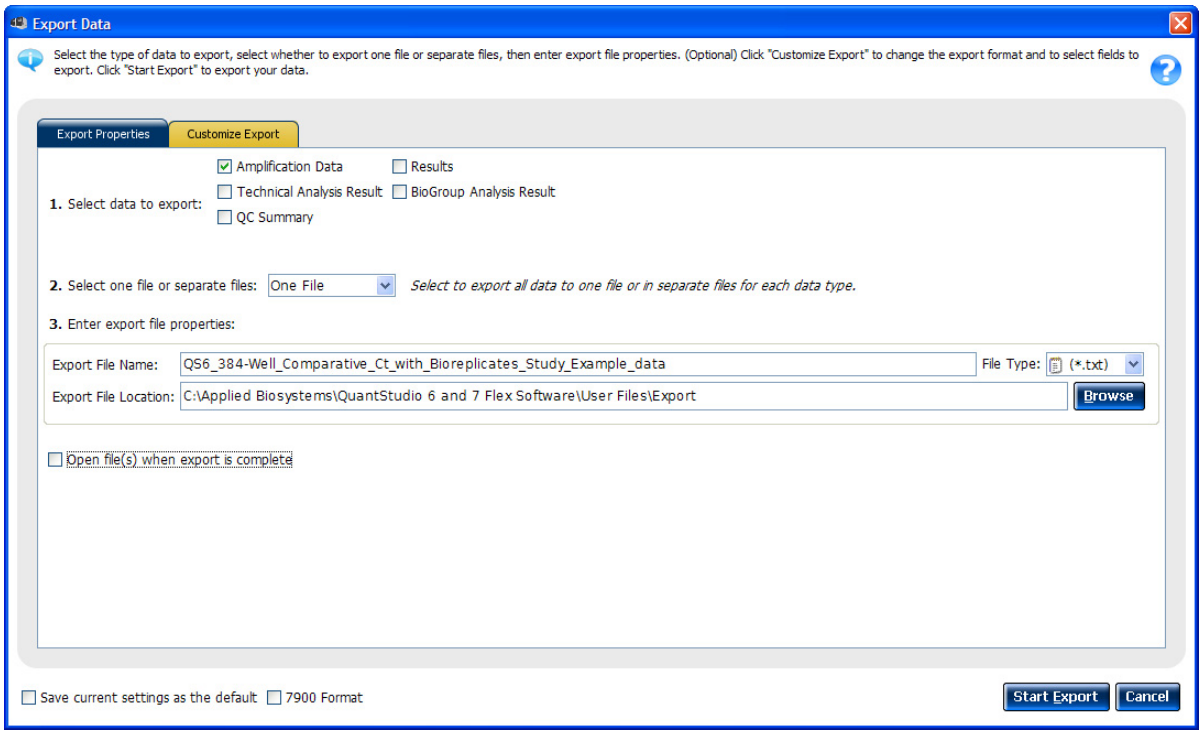
1. On the Analysis screen, click **Export** to access the Export Properties tab.
2. Define export properties.
   a. Select the data to export. Options are:
      - Amplification Data
      - Results
      - Technical Analysis Result
      - BioGroup Analysis Result
      - QC Summary
   b. Select **One File** or **Separate Files** from the drop-down menu to export all data to one file or in separate files for each data type respectively.
   c. Enter the export file properties and file name.
   d. Select the file type from the **File Type** drop-down menu. You can choose from ***.txt**, ***.xls**, and ***.xlsx**.
   e. Enter the Export File Location. The default location is C:\Applied Biosystems\QuantStudio 6 and 7 Flex Software\User Files\Export.
   f. Select the Open file(s) when export is complete check box to automatically open the file when export is complete.

   For the example study, enter:

Field or Selection	Entry
Select Data to export	Amplification Data
Select one file or separate files	One File
Export File Name	QS6_384-Well_Comparative_Ct_with_Bioreplicates_Study_Example_data
File Type	*.txt
Export File Location	C:\Applied Biosystems\QuantStudio 6 and 7 Flex Software\User Files\Export

Field or Selection	Entry
Open file(s) when export is complete	Unchecked
Save current settings as the default	Unchecked
7900 Format	Unchecked

The following is an image of the Export Data screen:



3. To change the export format, complete the tasks on the Customize Export tab.

   Note: To complete the tasks on the Customize Export tab you must have at least one type of data to export.

   a. Select the data from the Export Properties tab. The type of data that you selected in the Export Properties tab appears in the Customize field in the Customize Export tab.

   b. Select the data type content.

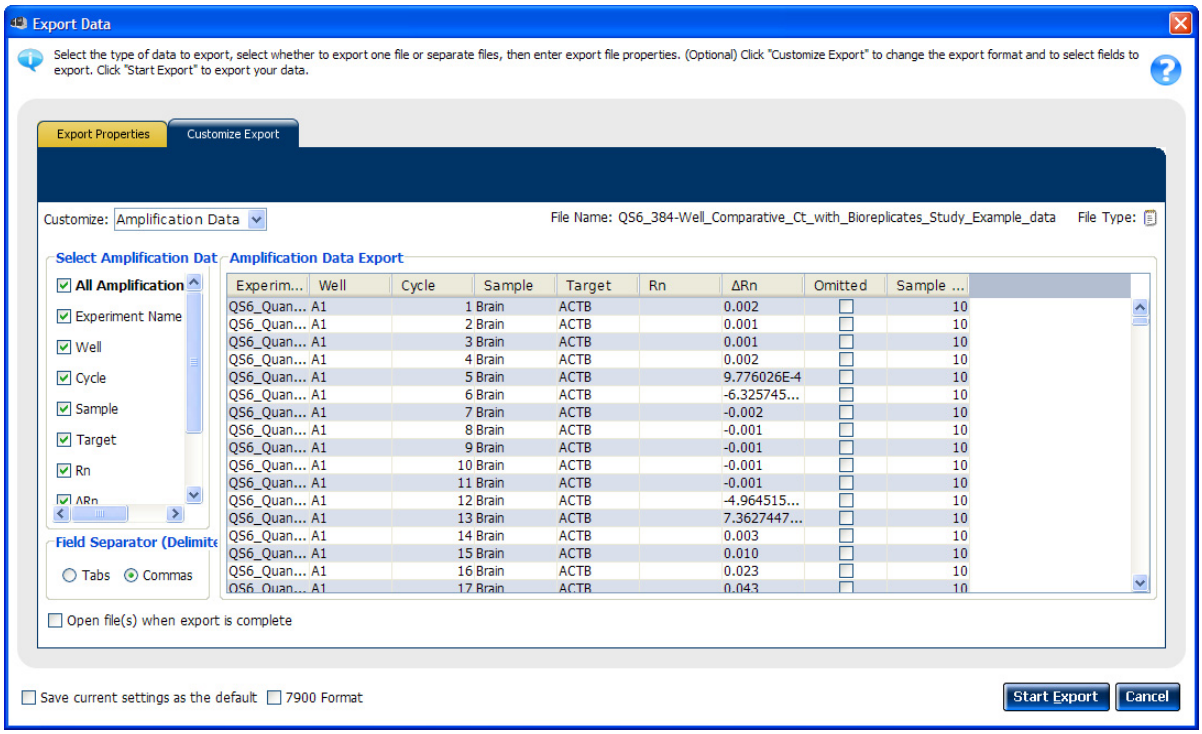
   c. Select the Tabs or Commas radio button to select the Field Separator (Delimiter).

   d. Select the Open file(s) when export is complete check box to automatically open the file when export is complete.

   For the example study, enter:

Field or Selection	Entry
Customize	Amplification Data
Field Separator (Delimited)	Commas
Open file(s) when export is complete	Unchecked
Save current settings as the default	Unchecked
7900 Format	Unchecked

The following is an image of the Export Data screen for customized export of Amplification Data:



4. Select the **Save current settings as the default** check box to save the export settings that you have modified. Alternatively, select the **7900 Format** check box to save the export settings in the 7900 format.

5. Click **Start Export**.

For more information

For more information on	Refer to	Publication number
Calculating Relative Quantification Values	*User Bulletin #2: Relative Quantitation of Gene Expression*.	4303859
Consumables	Chapter 1 in Booklet 1, *Getting Started with QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Experiments* Appendix A in Booklet 7, *QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Experiments - Appendixes*	4489822
Publishing data	Chapter 2 in Booklet 1, *Getting Started with QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Experiments.*	4489822

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USER GUIDE

applied biosystems®
by life technologies™

QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Getting Started Guide for Genotyping Experiments

Booklet 4

Publication Number 4489822
Revision A

life technologies™

Contents

1 About Genotyping Experiments

This chapter covers:

IMPORTANT! First-time users of the QuantStudio™ 6 and 7 Flex Real-Time PCR System Software please read Booklet 1, *Getting Started with QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Experiments* and Booklet 7, *QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Experiments - Appendixes* of this binder thoroughly. The booklet provides information and general instructions that are applicable to all the experiments described in this binder

Note: For more information about any of the topics discussed in this guide, access the Help from within QuantStudio™ 6 and 7 Flex Real-Time PCR System Software by pressing F1, clicking in the toolbar, or selecting **Help ▸ QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Help**.

About data collection

Genotyping experiments are performed to detect single nucleotide polymorphism (SNP) variants of a target nucleic acid sequence in samples. The PCR reactions contain primers designed to amplify the sequence containing the SNP and reagents to detect two different alleles.

You can collect the results of a genotyping experiment in two different ways: At the end of the experiment, or continuously during the experiment. Data collection at the end of the experiment is called end-point data collection. Data collection during the experiment run is considered real-time PCR. The real-time data helps further data analysis.

In end-point data collection, the normalized intensity of the reporter dye, or Rn, is the data collected. Some end-point experiments also include pre-PCR (data collected before the amplification process) data collection. The software calculates the delta Rn (ΔRn) value per the following formula:

ΔRn = Rn (post-PCR read) – Rn (pre-PCR read), where Rn = normalized readings.

About TaqMan® SNP Genotyping assays

A Genotyping assay detects variants of a single nucleic acid sequence, without quantifying the target. The presence of two probes in each reaction allows Genotyping of the two possible variants at the single nucleotide polymorphism (SNP) site in a target sequence.

Each TaqMan® SNP Genotyping Assay consists of a single, ready-to-use tube containing:

- Two sequence-specific primers for amplifying the polymorphism of interest
- Two allele-specific TaqMan® MGB probes for detecting the alleles for the specific polymorphism of interest

About TaqMan® MGB probes

Each allele-specific TaqMan® MGB probe has:

- A reporter dye at its 5′ end:
  - VIC® dye is linked to the 5′ end of the Allele 1 probe.
  - FAM™ dye is linked to the 5′ end of the Allele 2 probe.

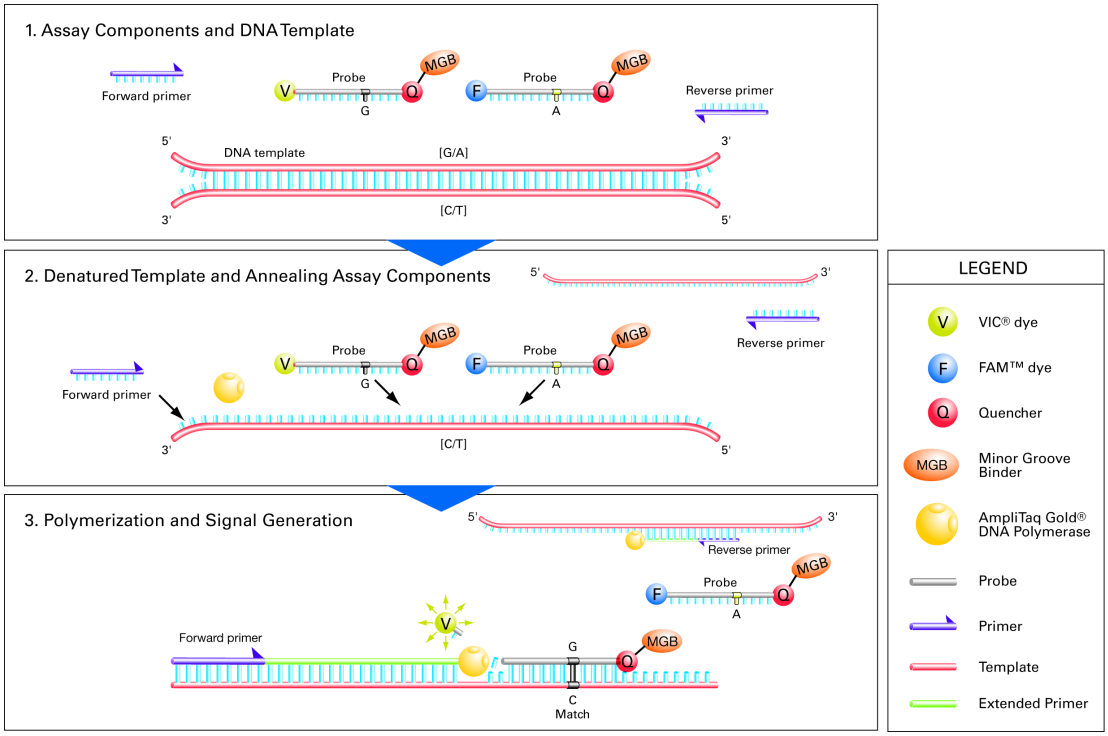
  The Allele 1 VIC® dye-labeled probe corresponds to the first nucleotide inside the square brackets of the context sequence in the assay information file (AIF) shipped with each order. The Allele 2 FAM™ dye-labeled probe corresponds to the second nucleotide inside the square brackets of the context sequence in the AIF. For the context sequence ATCGATT[G/T]ATCC, the VIC® dye-labeled probe binds to the G allele, and the FAM™ dye-labeled probe to the T allele.
- A minor groove binder (MGB), which increases the melting temperature (T_m) for a given probe length and allows the design of shorter probes. The use of shorter probes results in greater differences in T_m values between matched and mismatched probes, and more robust genotyping.
- A non-fluorescent quencher (NFQ) at its 3′ end, which allows for detection of the reporter dye fluorescence with greater sensitivity than with a fluorescent quencher.

5′ nuclease assay

The figure below is a schematic depiction of the 5′ nuclease assay. During PCR:

- Each TaqMan® MGB probe anneals specifically to its complementary sequence between the forward and reverse primer sites.
- When the oligonucleotide probe is intact, the proximity of the quencher dye to the reporter dye quenches the reporter signal.
- AmpliTaq Gold® DNA polymerase extends the primers bound to the genomic DNA template.

- AmpliTaq Gold® DNA polymerase (with its 5′ nuclease activity) cleaves probes that are hybridized to the target sequence.
- Cleavage of the probes hybridized to the target sequence separates the quencher dye from the reporter dye, resulting in increased fluorescence by the reporter. The fluorescence signal generated by PCR amplification indicates which alleles are present in the sample.



Minimizing non-specific fluorescence

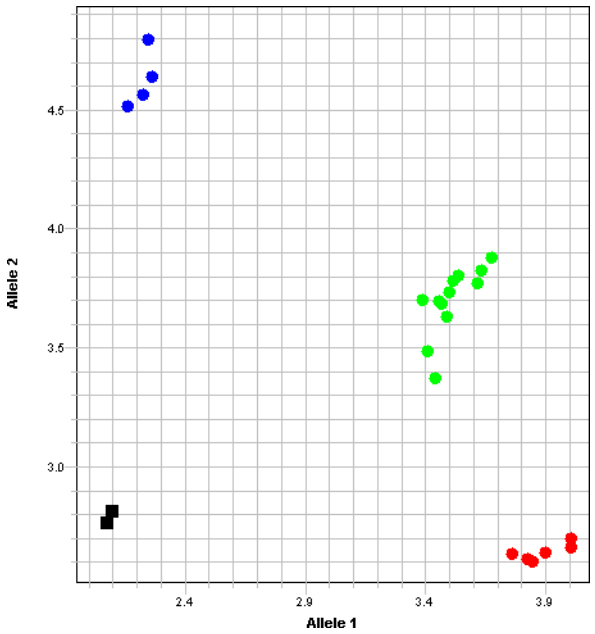
In TaqMan® assays, fluorescence from nonspecifically bound probes is reduced, because nucleotide mismatches between a probe and a sequence reduce the chances that the probe will be cleaved. The probe's short length means that a one-base-pair mismatch will a have a larger negative effect on the binding. The mismatched probe will not bind tightly to the allele; the AmpliTaq Gold® DNA polymerase will likely displace the probe without cleaving the dye.

Reading and analyzing the plates

The QuantStudio™ 6 and 7 Flex Real-Time PCR System Software genotypes the DNA samples from the reaction plate simultaneously. First, the software normalizes the fluorescence of the reporter dyes to the fluorescence of the passive reference dye in each well. Next, the software plots the normalized intensities (Rn) of the reporter dyes

in each sample well on an Allelic Discrimination Plot, which contrasts the reporter dye intensities of the allele-specific probes. Finally, the QuantStudio™ 6 and 7 Flex Real-Time PCR System Software algorithmically clusters the sample data, and assigns a genotype call to the samples of each cluster according to its position on the plot.

Note: The QuantStudio™ 6 and 7 Flex Real-Time PCR System Software clustering algorithm does not call genotypes when only one genotype is present in an experiment.



The clustering of datapoints can vary along the horizontal axis (Allele 1), vertical axis (Allele 2), or diagonal (Allele 1/Allele 2). This variation results from differences in the extent of reporter dye fluorescent intensity after PCR amplification. The table below shows the correlation between fluorescence signals and sequences in a sample.

A substantial increase in...	Indicates...
VIC® dye-labeled probe fluorescence only	Homozygosity for Allele 1
FAM™ dye-labeled probe fluorescence only	Homozygosity for Allele 2
Both VIC® and FAM™ dye-labeled probes fluorescence	Allele 1-Allele 2 heterozygosity

About the example experiment

To illustrate how to perform Genotyping experiments, this guide leads you through the process of designing and performing an example experiment. The example experiment represents a typical setup that you can use to quickly familiarize yourself with a QuantStudio™ 6 and 7 Flex Real-Time PCR System Software.

The objective of the example Genotyping experiment is to investigate SNP rs8039, where possible genotypes are AA, AC, and CC. In the example, two unknown genomic DNA (gDNA) samples were genotyped using TaqMan® Drug Metabolism Genotyping Assay ID C__11711420_30. The reactions were set up so that the PCR

primers and probes that target both alleles of SNP rs8039 were present in the same well. The PCR was performed using the TaqMan® Genotyping Master Mix and run according to the protocol that is described in the *Performing a TaqMan® Drug Metabolism Genotyping Assay.*

2 Design the Experiment

This chapter explains how to design the example experiment from the Experiment Setup menu.

This chapter covers:

Note: To automatically export the analyzed data to a specified location, select the Auto Export task from the Export screen, before running the experiment. For more information on Auto Export, refer to Chapter 2 in Booklet 1, *Getting Started with QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Experiments*.

Define the experiment properties

Click **Experiment Setup** ▸ **Experiment Properties** to create a new experiment in the QuantStudio™ 6 and 7 Flex Real-Time PCR System Software. Enter:

Field	Entry
Experiment Name	QuantStudio_384-Well_SNP_Genotyping_Example
Barcode	Leave field empty
User Name	Example User
Comments	Genotyping example
Instrument type	QuantStudio™ 6 Flex System
Block	384-Well Block
Experiment Type	Genotyping
Reagents	TaqMan® Reagents
Ramp speed	Standard
Reagent information	NA

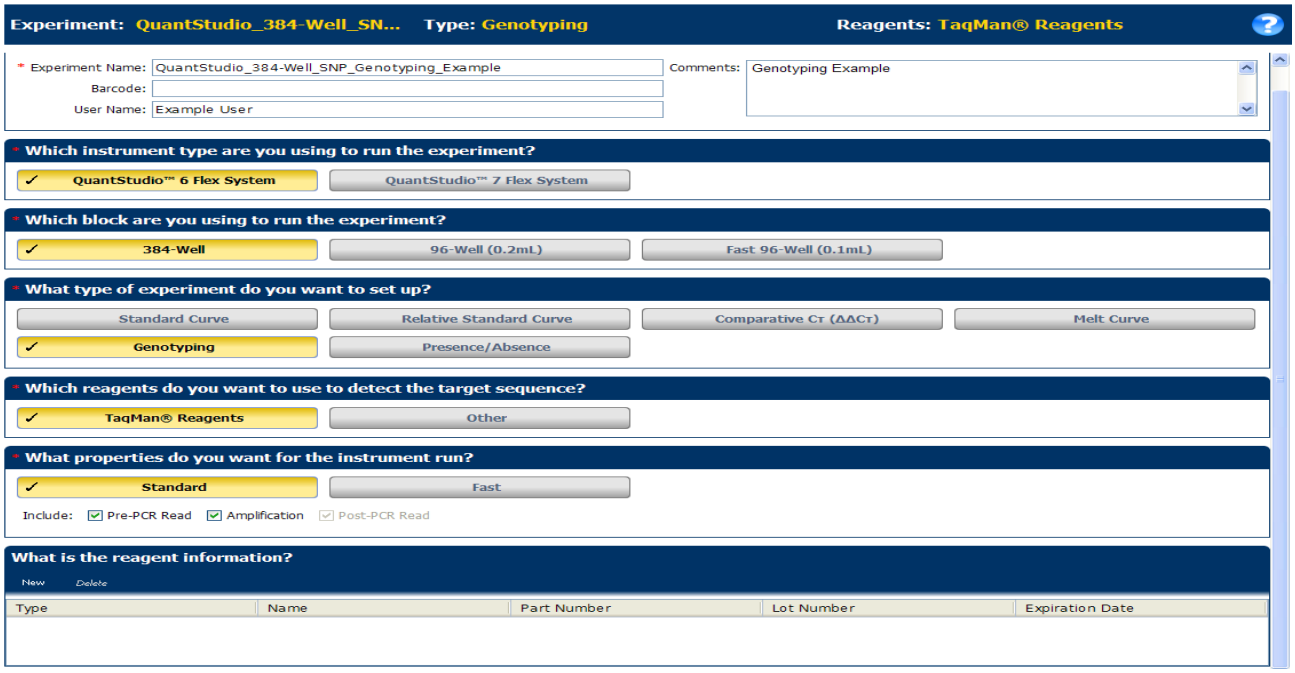
Select all three data-collection check boxes: Pre-PCR, Amplification, and Post-PCR collection methods:

Pre-PCR Read	Checked
Amplification	Checked

Post-PCR Read	Checked

Save the experiment.

Your Experiment Properties screen should look like this:



Define SNPs and samples

Click **Define** to access the Define screen. Enter:

1. SNP Assays

SNP assay name	NCBI SNP reference	Context sequence	Allele 1	Reporter	Quencher	Allele 2	Reporter	Quencher	Color
SNP Assay 1			Allele1	VIC	NFQ-MGB	Allele2	FAM	NFQ-MGB	
SNP Assay 2			Allele1	VIC	NFQ-MGB	Allele2	FAM	NFQ-MGB	

Note: The NCBI SNP reference and Context sequence fields are optional fields and are used for reference. They are not required to run an experiment.

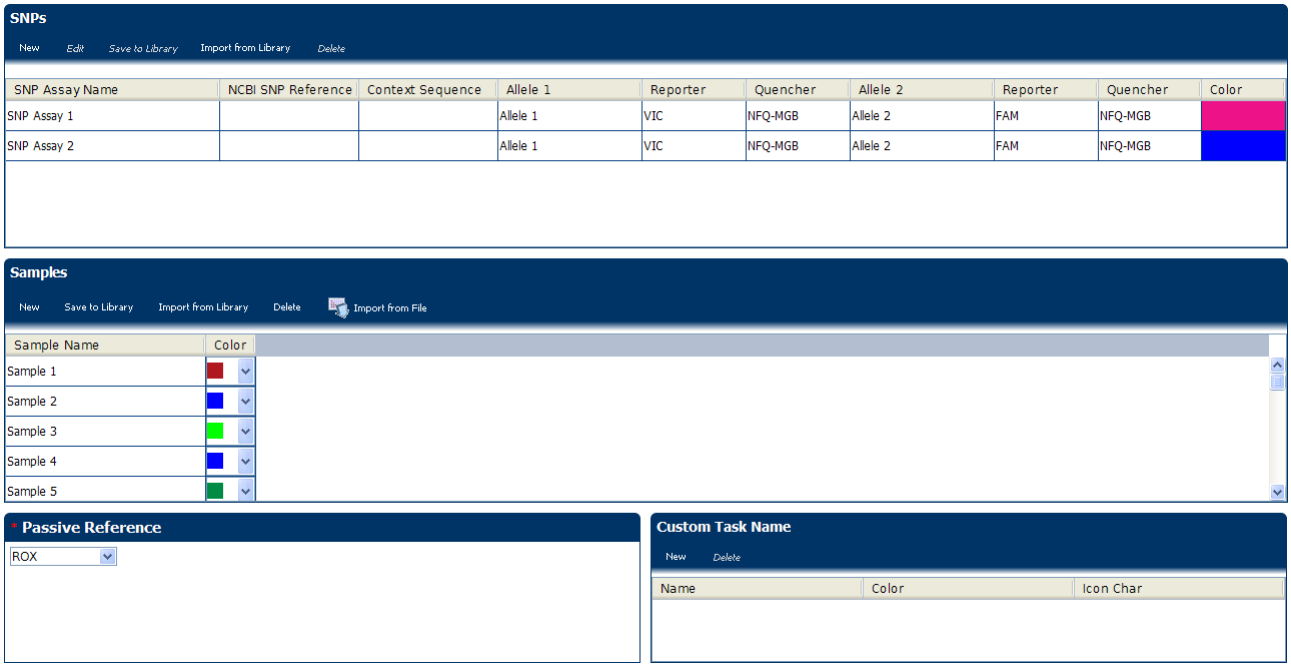
2. Samples

Sample name	Color	Sample name	Color	Sample name	Color
Sample 1		Sample 11		Sample 21	

Sample name	Color	Sample name	Color	Sample name	Color
Sample 2		Sample 12		Sample 22	
Sample 3		Sample 13		Sample 23	
Sample 4		Sample 14		Sample 24	
Sample 5		Sample 15		Sample 25	
Sample 6		Sample 16		Sample 26	
Sample 7		Sample 17		Sample 27	
Sample 8		Sample 18		Sample 28	
Sample 9		Sample 19		Sample 29	
Sample 10		Sample 20		Sample 30	

3. Dye to be used as a Passive Reference
   ROX
4. Custom Task Name
   Not applicable

Your Define screen should look like this:



Note: This example experiment does not define biological replicate groups. Leave Biological Replicate Groups blank.

Assign markers, samples, and controls

Click **Assign** to access the Assign screen. Enter the SNP assays and samples:

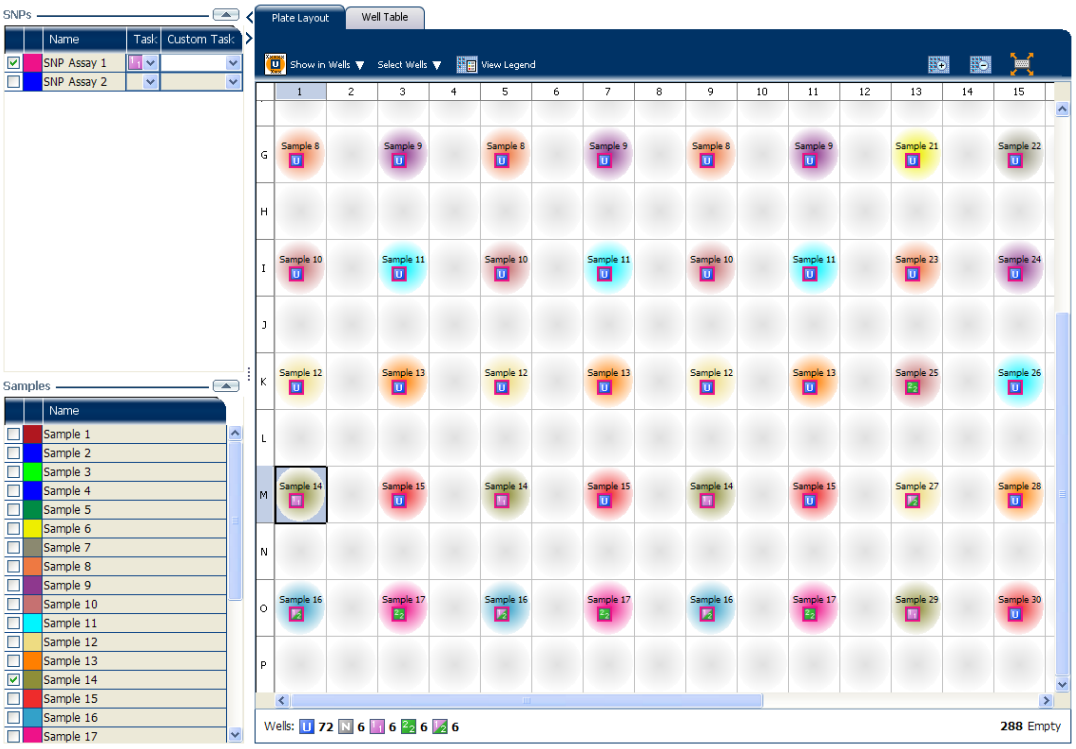
- SNP Assay 1

Target name	Well number	Task	Sample
SNP Assay 1	A3, A7, A11	Unknown	Sample 3
	C1, C5, C9	Unknown	Sample 4
	C3, C7, C11	Unknown	Sample 5
	E1, E5, E9	Unknown	Sample 6
	E3, E7, E11	Unknown	Sample 7
	G1, G5, G9	Unknown	Sample 8
	G3, G7, G11	Unknown	Sample 9
	I1, I5, I9	Unknown	Sample 10
	I3, I7, I11	Unknown	Sample 11
	K1, K5, K9	Unknown	Sample 12
	K3, K7, K11	Unknown	Sample 13
	M1, M5, M9	1/1	Sample 14
	M3, M7, M11	Unknown	Sample 15
	O1, O5, O9	1/2	Sample 16
	O3, O7, O11	2/2	Sample 17
SNP Assay 1	A1, A5, A9	Negative	NTC

- SNP Assay 2

Target name	Well number	Task	Sample
SNP Assay 2	A15, A19, A23	Unknown	Sample 1
	C13, C17, C21	Unknown	Sample 2
	C15, C19, C23	Unknown	Sample 18
	E13, E17, E21	Unknown	Sample 19
	E15, E19, E23	Unknown	Sample 20
	G13, G17, G21	Unknown	Sample 21
	G15, G19, G23	Unknown	Sample 22
	I13, I17, I21	Unknown	Sample 23
	I15, I19, I23	Unknown	Sample 24
	K13, K17, K21	2/2	Sample 25
	K15, K19, K23	Unknown	Sample 26
	M13, M17, M21	1/2	Sample 27
	M15, M19, M23	Unknown	Sample 28
	O13, O17, O21	1/1	Sample 29
	O15, O19, O23	Unknown	Sample 30
SNP Assay 2	A13, A17, A21	Negative	NTC

Your Assign screen should look like this:



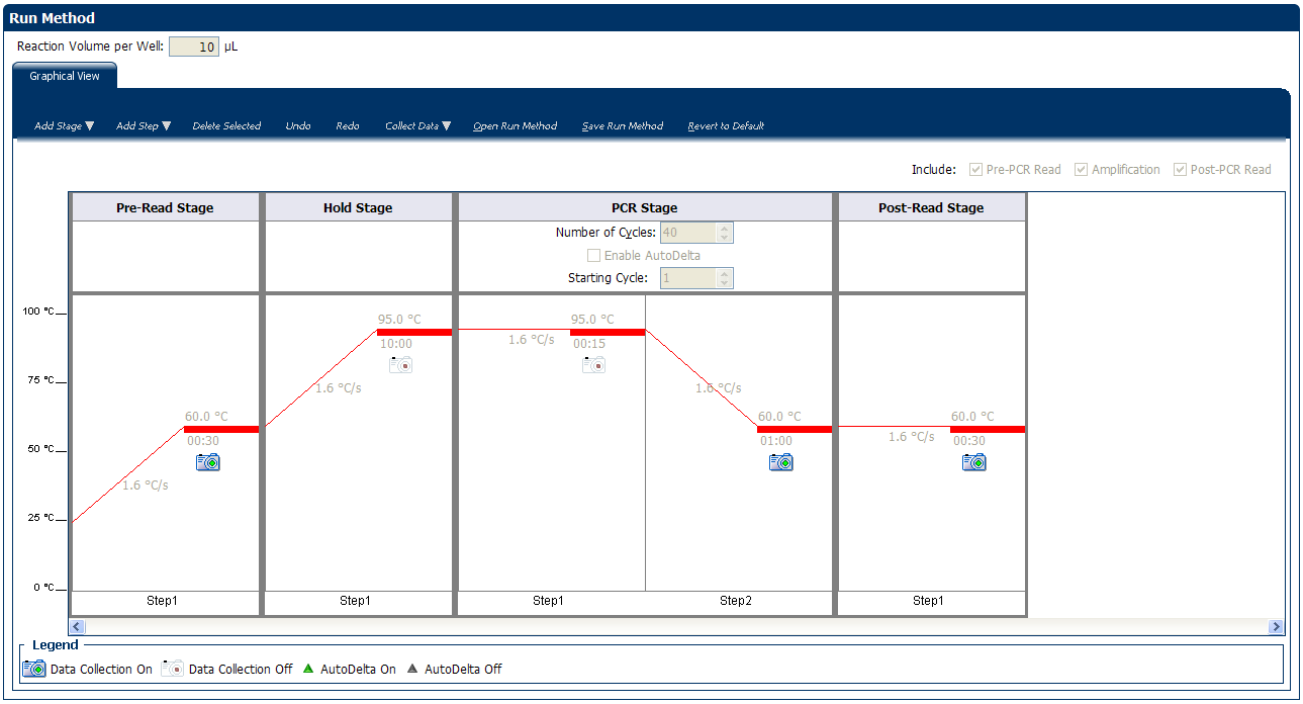
Set up the run method

Click **Run Method** to access the Run Method screen. Set the thermal profile under the Graphical View tab. Enter:

- Reaction Volume Per Well: 10 µL
- Thermal Profile

Stage	Step	Ramp rate	Temperature	Time
Pre-Read Stage	Step 1	1.6°C/s	60 °C	30 seconds
Hold Stage	Step 1	1.6°C/s	95°C	10 minutes
PCR Stage • Number of Cycles: 40 (default) • Enable AutoDelta: Unchecked (default) • Starting Cycle: Disabled when Enable AutoDelta is unchecked	Step 1	1.6°C/s	95°C	15 seconds
	Step 2	1.6°C/s	60°C	1 minute
Post-Read Stage	Step 1	1.6°C/s	60°C	30 seconds

Your Run Method screen should look like this:



For more information

For more information on...	Refer to...	Publication number
Consumables	Chapter 1 in Booklet 1, *Getting Started with QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Experiments* Appendix A in Booklet 7, *QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Experiments - Appendixes*	4489822
Data collection	Chapter 2 in Booklet 1, *Getting Started with QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Experiments*	4489822
Amplification efficiency	*Amplification Efficiency of TaqMan® Gene Expression Assays Application Note*	127AP05-03
Using Alternative Setup	Chapter 3 in Booklet 1, *Getting Started with QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Experiments*	4489822

3 Prepare the Reactions

This chapter explains how to prepare the PCR reactions for the Genotyping example experiment.

This chapter covers:

Assemble required materials

- Items listed in Booklet 1, *Getting Started with QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Experiments*
- Samples - Sample 1 - Sample 30
- Example experiment reaction mix components:
  - **TaqMan® Genotyping Master Mix (2X)**
  - SNP 1 Assay Mix (20×)
  - SNP 2 Assay Mix (20×)

Note: Fast Universal Master Mix is not recommended to be used with the 96-well (0.2 mL) reaction plates or reaction tubes and tube strips sealed with caps.

Prepare the sample dilutions

For the example experiment, two targets are assigned to 36 wells each. Each well contains 20 ng of Coriell DNA diluted from 100 ng/µL of stock.

To prepare the sample dilutions:

1. Label a separate microcentrifuge tube for each sample to be diluted.

2. Add 5µL of sample stock to each empty tube.
3. Add 45µL of sterile water (diluent) to each tube, such that each working stock tube has a final concentration of 10 ng/µL
4. Vortex each diluted sample for 3 to 5 seconds, then centrifuge the tubes briefly.
5. Place the diluted samples on ice until you prepare the reaction plate.

Prepare the reaction mix ("cocktail mix")

1. Label an appropriately sized tube for each reaction mix:
   - SNP 1 Reaction Mix
   - SNP 2 Reaction Mix
2. For SNP Assay 1, prepare a cocktail by adding the required volumes of each component to the SNP 1 reaction tube, as detailed below.

Reaction component	Reaction volume			
	Per well (µL)		36 Reactions + 10% excess (µL)	
	Dry	Wet	Dry	Wet
TaqMan® Genotyping Master Mix (2X)	10.0	10.0	400.0	400.0
SNP Assay Mix (20X)	1.0	1.0	40.0	40.0
H_2O, DNase-free	9.0	7.0	360.0	280.0
Total Reaction Mix Volume	20.00	18	800.0	720.0

3. Gently pipet the reaction mix up and down, then cap the tube.
4. Centrifuge the tube briefly.
5. Place the reaction mixes on ice until you prepare the reaction plate.
6. Repeat step 2 through 5 for the SNP 2 assay.

   Note: Do not add the sample at this time.

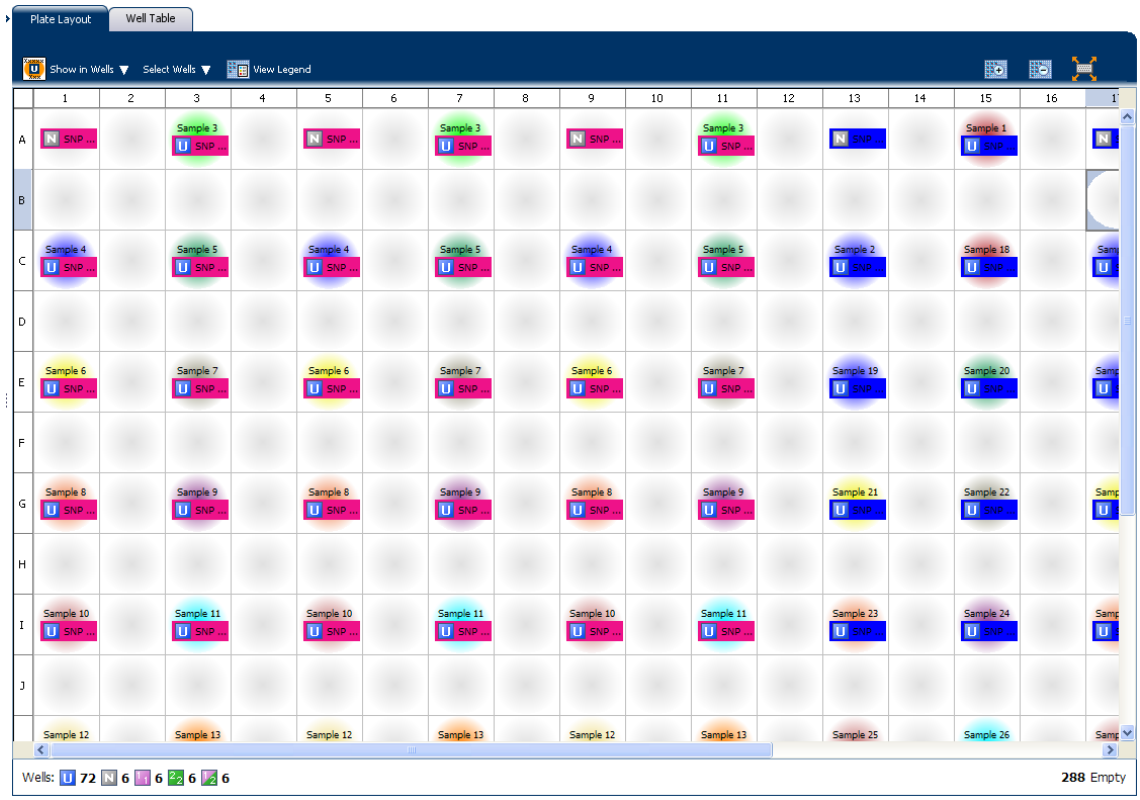
Prepare the reaction plate

Example experiment reaction plate components

The reaction plate for the Genotyping example experiment contains:

- A MicroAmp® Optical 384-Well Reaction Plate
- Reaction volume: 10 µL/well
- 72 Unknown wells 

The following is an image of the plate layout:



To prepare the reaction plate: dried gDNA

1. Pipet 2.0 µL of the appropriate sample (20 ng of purified genomic DNA) into each well of the reaction plate.

   All wells belonging to the same Genotyping assay must contain approximately the same quantity of sample or control.

   Note: While preparing the reaction plate for your own Genotyping experiment, add between 1 and 20 ng of purified DNA per reaction.

2. Dry down the samples by evaporation at room temperature in a dark, amplicon-free location. (Cover the reaction plate with a lint-free tissue while drying.)

3. Transfer 20 µL of reaction mix to each well.

   IMPORTANT! Make sure that no cross-contamination occurs from well to well.

4. Seal the reaction plate with adhesive film.

5. Vortex the reaction plate for 3 to 5 sec.

6. Briefly centrifuge the reaction plate.

7. Confirm that the liquid is at the bottom of each well of the reaction plate. If not, centrifuge the plate again at a higher speed and for a longer period of time.

To prepare the reaction plate: wet gDNA

1. Add 2 µL of DNA to the appropriate wells.
2. Add 2 µL of water to wells containing the NTCs.
3. Transfer 18 µL of reaction mix to the appropriate wells.
4. Seal the reaction plate with optical adhesive film.
5. Vortex the reaction plate for 3 to 5 seconds, then briefly centrifuge it.
6. Centrifuge the reaction plate briefly.
7. Until you are ready to perform the run, place the reaction plate at 4°C, in the dark.

Tips for preparing reactions for your own experiments

Tips for preparing samples

When you prepare the samples for your own experiment:

- Use DNAse-free water to dilute the samples.
- Use the same quantity of DNA per well for each experiment.

Tips for preparing the reaction mix

When you prepare the reaction mix for your own experiment, make sure you prepare the reactions for each SNP separately.

Prior to use:

- Mix the master mix thoroughly by swirling the bottle.
- Resuspend the assay mix by vortexing, then centrifuge the tube briefly.
- Thaw frozen samples by placing them on ice. When thawed, resuspend the samples by vortexing, then centrifuge the tubes briefly.

Tips for preparing the reaction plate

When you prepare the reaction plate for your own experiment:

- Make sure the reaction locations match the plate layout in the QuantStudio™ 6 and 7 Flex Real-Time PCR System Software.
- Load 1 to 20 ng of purified genomic DNA per reaction
- All wells belonging to the same Genotyping assay must contain approximately the same quantity of sample or control.
- Multiple assays may be run on one reaction plate, but must be analyzed separately.

For more information

For more information on...	Refer to...	Publication number
Assigning the reaction plate components	Chapter 2 in Booklet 1, *Getting Started with QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Experiments*	4489822
Sealing the reaction plate	Chapter 2 in Booklet 1, *Getting Started with QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Experiments*	4489822

4 Run the Experiment

This chapter explains how to run the example experiment on the QuantStudio™ 6 or 7 Instrument.

This chapter covers:

IMPORTANT! Run the experiment at the same ambient temperature at which you calibrated the instrument. Extreme variations in ambient temperature can affect the heating and cooling of the instrument and influence experimental results.

IMPORTANT! Do not attempt to open the access door during the run. The door is locked while the QuantStudio™ 6 or 7 Instrument is in operation.

Start the run

1. Open the Genotyping example file that you created using instructions in Chapter 2.
2. Load the reaction plate into the instrument.
3. Start the run.

   Note: To collect real-time data during a run, click the  button on the Run Method screen in the Experiment Setup menu.

Monitor the run

Monitor the example experiment run:

- From the QuantStudio™ 6 and 7 Flex Real-Time PCR System Software using the Run screen, while the experiment is in progress.
- From the Instrument Console of the QuantStudio™ 6 and 7 Flex Real-Time PCR System Software (to monitor an experiment started from another computer or from the QuantStudio™ 6 or 7 Instrument touchscreen).
- From the QuantStudio™ 6 or 7 Instrument touchscreen.

From the Instrument Console of the QuantStudio™ 6 and 7 Flex Real-Time PCR System Software

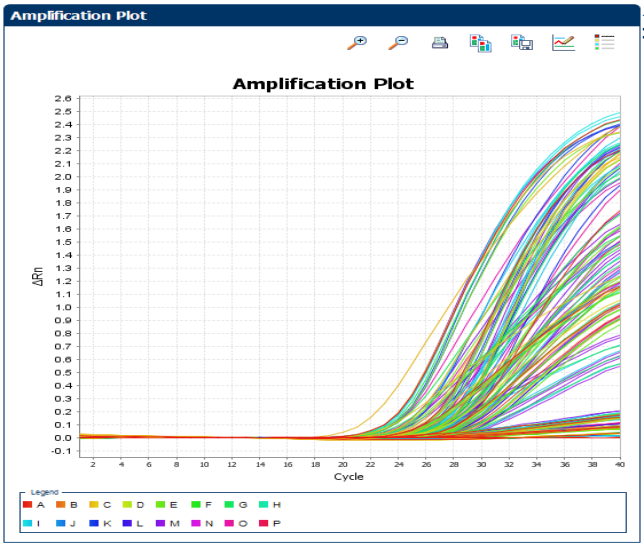
1. In the Instrument Console screen, select the instrument icon.
2. Click **Manage Instrument** or double-click on the instrument icon.
3. On the Manage Instrument screen, click **Monitor Running Instrument** to access the Run screen.

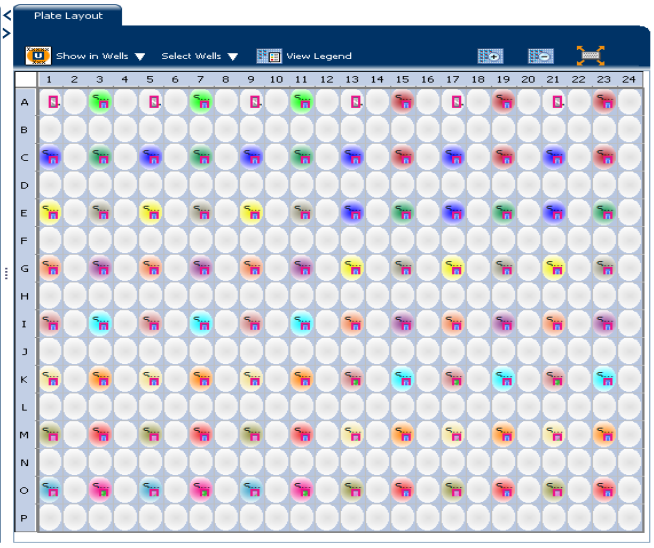
View the Amplification Plot

You can view the progress of the run in real time. During the run, periodically view all the three plots available from the QuantStudio™ 6 and 7 Flex Real-Time PCR System Software for potential problems.

Click **Amplification Plot** from the Run Experiment Menu, select the Plate Layout tab, then select the wells to view.

The following is an image of the Amplification Plot screen as it appears at the end of the example experiment.

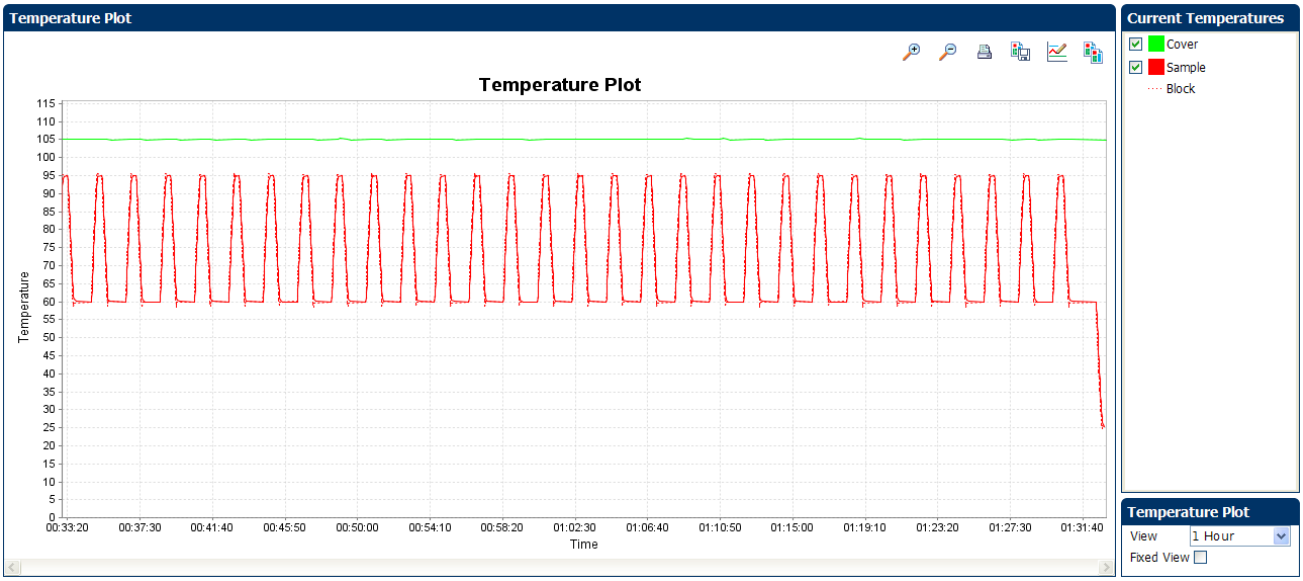




View the Temperature Plot

Click **Temperature Plot** from the Run Experiment Menu.

The following is an image of the Temperature Plot screen as it appears during the example experiment.

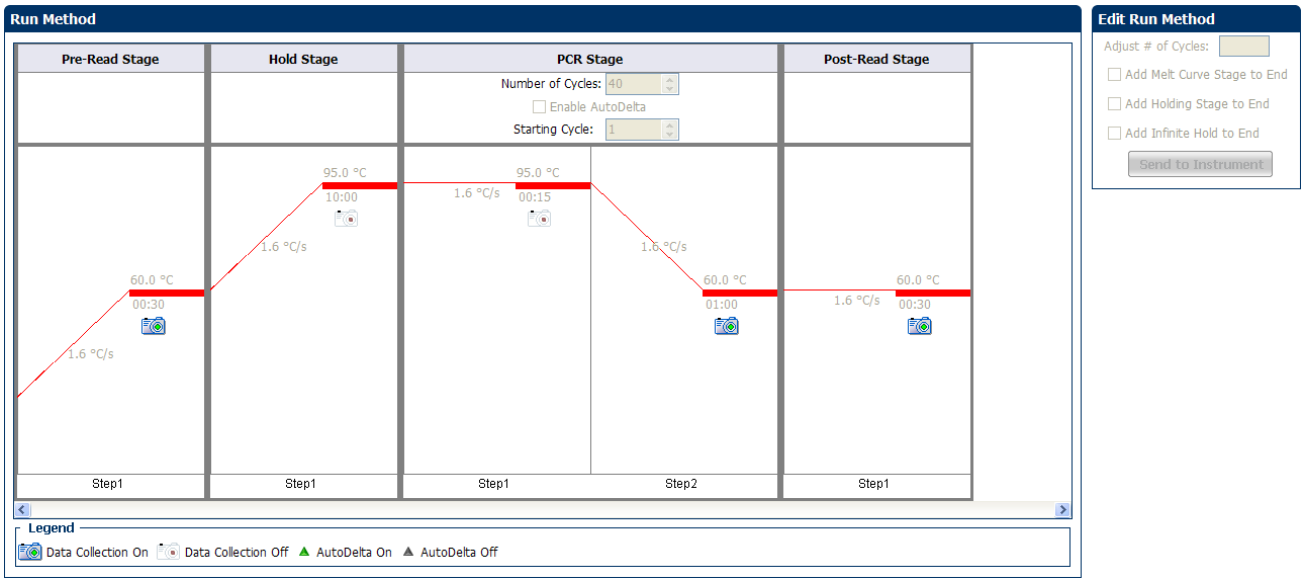


Note: The sample temperature displayed in the Current Temperatures group is a calculated value.

View the Run Method

Click **Run Method** from the Run Experiment Menu.

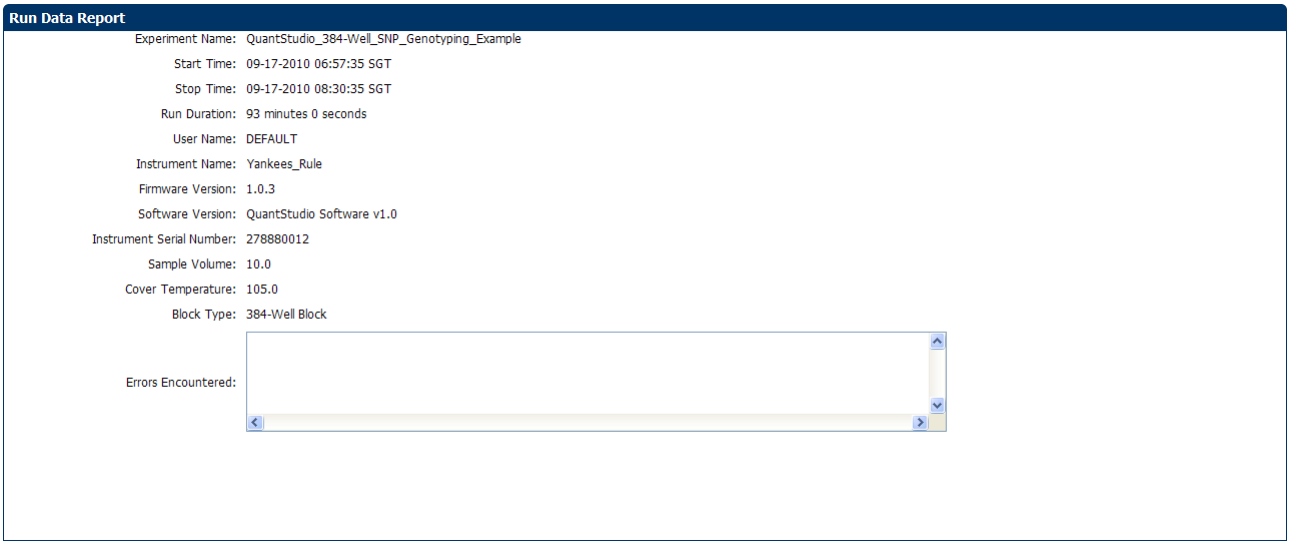
The following is an image of the Run Method screen as it appears in the example experiment.



View run data

Click **View Run Data** from the Run Experiment Menu.

The following is an image of the View Run Data screen as it appears in the example experiment.

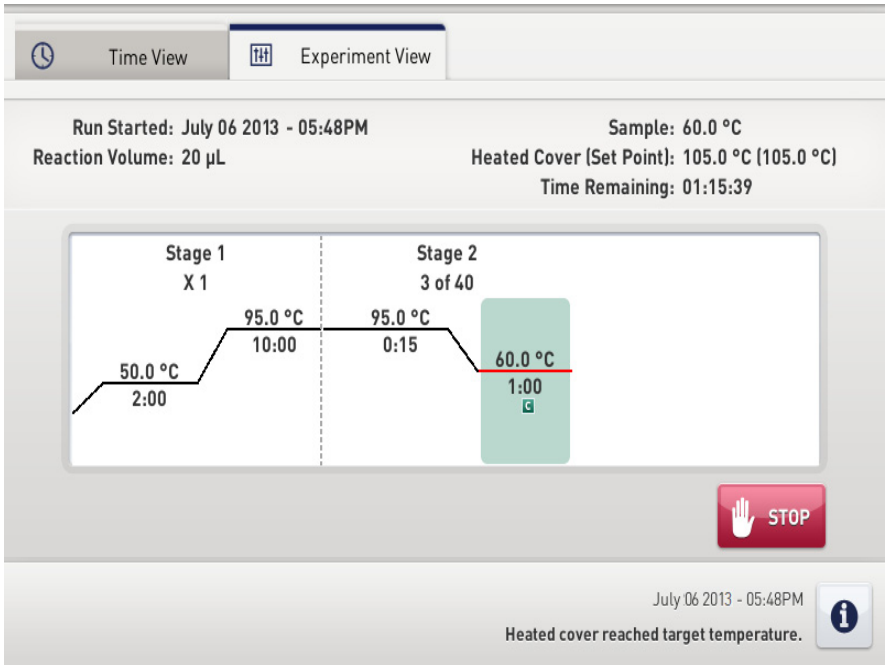


From the QuantStudio™ 6 or 7 Instrument touchscreen

You can also view the progress of the run from the QuantStudio™ 6 or 7 Instrument touchscreen.

The following are images of the Run Method screen on the QuantStudio™ 6 or 7 Instrument touchscreen:

Experiment View



Time View


Time View
Experiment View
Run Started: July 06 2013 - 05:48PM
Reaction Volume: 20 µL
Sample: 95.0 °C
Heated Cover (Set Point): 105.0 °C (105.0 °C)
Stage / Step / Cycle: 1 / 2 / 1
01:31:52
Remaining Time
Elapsed Time
July 06 2013 - 05:48PM
Heated cover reached target temperature.

5 Review Results and Adjust Experiment Parameters

In Section 5.1 of this chapter you review the analyzed data using several of the analysis screens and publish the data. Section 5.2 of this chapter walks you through the process of modifying experiment parameters to troubleshoot problems with experiment results before rerunning an experiment.

This chapter covers:

Section 5.1 Review Results

Analyze the example experiment

1. Open the example experiment file that you ran in Chapter 4.
2. Click **Analyze**. The software analyzes the data using the default analysis settings.

   Note: You can also access the experiment to analyze from the Home screen.

View clusters in the Allelic Discrimination Plot

The Allelic Discrimination Plot contrasts the normalized reporter dye fluorescence (Rn) for the allele-specific probes of the SNP assay.

View the allelic discrimination plot to identify:

- Clusters for the three possible genotypes (Allele 1 homozygous, Allele 2 homozygous, and Allele 1/2 heterozygous).
- A cluster for the negative controls.

Note: Including controls helps to improve the clustering algorithm, particularly in situations where a limited number of samples are run.

To view and assess the allelic discrimination plot

1. From the Experiment menu pane, select **Analysis ▸ Allelic Discrimination Plot.**
2. Click the **Plate Layout** tab, then click any empty well to select it.

   Note: In the Allelic Discrimination Plot, the software highlights all wells that are selected in the Plate Layout tab. If the plot displays a single color for all wells, then all wells in the plate layout are selected.
3. In the allelic discrimination plot, select **SNP Assay 1** from the SNP Assay menu, then enable Autocaller.

   The Allelic Discrimination Plot displays allele symbols for each sample evaluated for the selected SNP. The samples are grouped on the plot as follows:

Genotype	Symbol	Location
Homozygous for Allele 1 of the selected SNP assay.	● (red)	X-axis of the plot
Homozygous for Allele 2 of the selected SNP assay.	● (blue)	Y-axis of the plot
Heterozygous for both alleles of the selected SNP assay (Allele 1 and Allele 2).	● (green)	Midway between the homozygote clusters
A negative control.	■ (black)	Bottom-left corner of the plot
Undetermined.	✸ (black)	Anywhere on plot

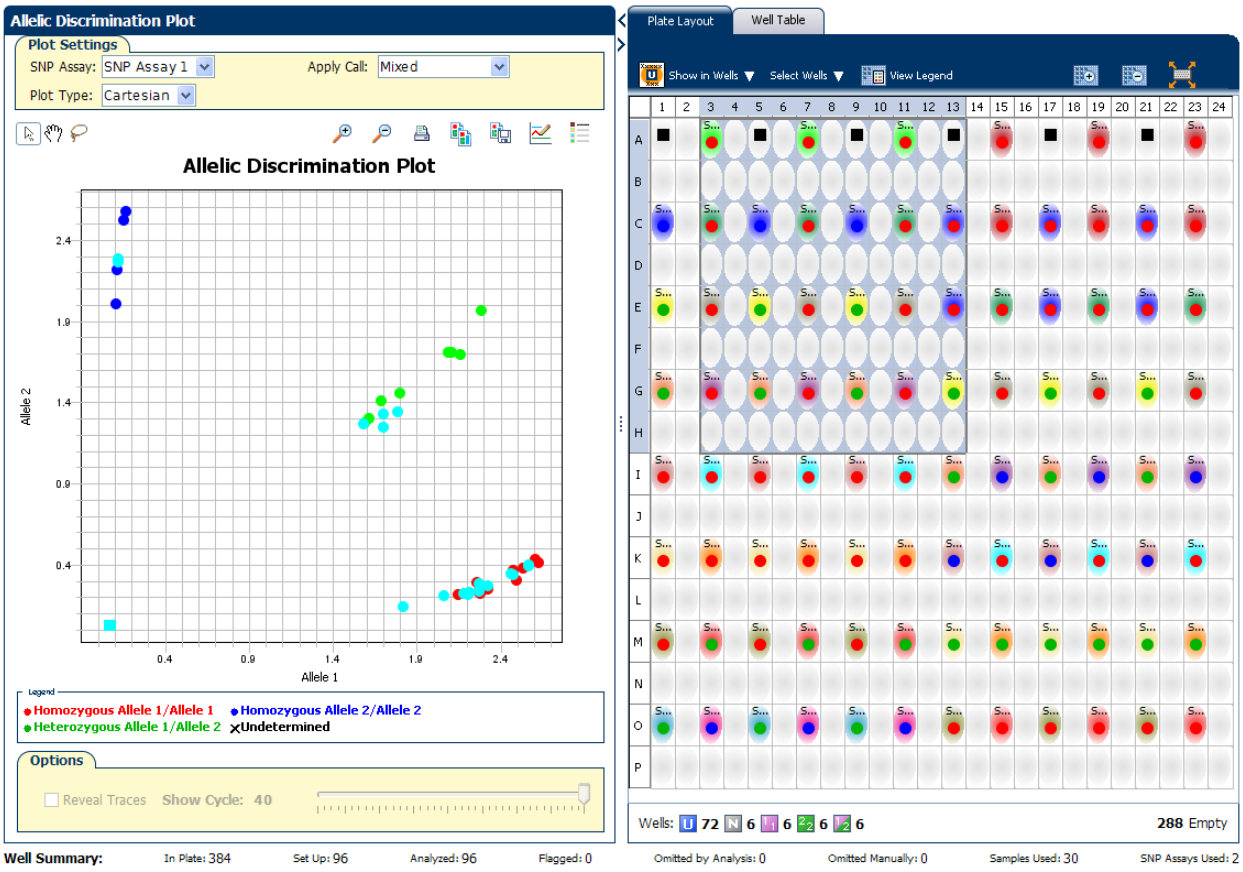
Note: If the Autocaller is not enabled, the Allelic Discrimination Plot displays a crossmark (X – Undetermined) for each sample.

4. Review each cluster in the plot:
   a. Click and drag a box around the cluster to select the associated wells in the plate layout and well table.
   b. Confirm that the expected wells are selected in the well table.

      For example, if you select the cluster at the bottom-left corner of the plot, only the negative controls should be selected. The presence of an unknown among the negative controls may indicate that the sample failed to amplify.
   c. Repeat steps a and b for all other clusters in the plot.
   d. The table below describes the elements of the Allelic Discrimination Plot.

Element	Description
SNP Assay drop-down menu	Determines the SNP assay data that the QuantStudio™ 6 and 7 Flex Software displays in the plot.
Plot Type drop-down menu	Determines the type of plot (Cartesian or Polar) that the QuantStudio™ 6 and 7 Flex Software uses to display the data.
Apply Call drop-down menu	When a datapoint is selected, this menu allows you to assign an allele call to the datapoint within the scatterplot.
Toolbar	Contains tools for manipulating the scatterplot: •  – Selection tool. •  – Selection tool. •  – Repositioning tool. •  – Zooms in. •  – Zooms out.
Legend	Explains the symbols in the scatterplot.
Options	The Reveal Traces option allows you to trace the clusters throughout the PCR process. This option is not activated for the example experiment. To activate the feature, see "Adjust analysis settings" on page 50.

The following is an image of the Allelic Discrimination plot for the example experiment:



Troubleshoot clustering on the Allelic Discrimination Plot

Do all controls have the correct genotype?

In the example experiment and in your own experiments, confirm that data points cluster as expected.

Clustering in positive controls

1. From the well table, select the wells containing a positive control to highlight the corresponding data points (symbols) in the Allelic Discrimination Plot.
2. Check that the data points for the positive controls cluster along the expected axis of the plot. For example, if you select the Positive Control Allele 1/Allele 1, then the controls should cluster along the X-axis.
3. Repeat steps 1 and 2 for the wells containing the other positive controls.

Failed amplification in the unknown samples

1. Select the data points of the cluster in the lower left corner of the Allelic Discrimination Plot to select the corresponding wells in the well table.
2. Check that the selected wells in the well table are negative controls, and not unknown samples.

Samples clustered with negative control

Samples that clustered with the negative controls may:

- Contain no DNA
- Contain PCR inhibitors
- Be homozygous for a sequence deletion

Confirm the results of these samples by retesting them.

Are outliers present?

If the Allelic Discrimination Plot contains clusters other than the three representative genotype clusters (heterozygous, homozygous allele 1, and homozygous allele 2), then those can be classified as outliers.

Confirm the results of the associated samples by retesting them.

Note: The results displays are synchronized. For example, selecting a well in the plate layout selects the corresponding data in the well table and Allelic Discrimination Plot.

Confirm setup accuracy using Plate Layout

Review the experiment results in the Plate Layout. The plate layout displays the assay-specific setup and analysis properties for the experiment in a well format corresponding to the type of reaction plate used for the run.

Example experiment plate layout values

For the example experiment, confirm that the QuantStudio™ 6 and 7 Flex Software called:

- 54 samples as Allele 1 homozygous (●)
- 12 samples as Allele 2 homozygous (●)
- 24 samples as heterozygous (●)
- 0 samples as undetermined (X)

Confirm that no wells of the reaction plate triggered QC flags (▲). The example experiment does not display any flags.

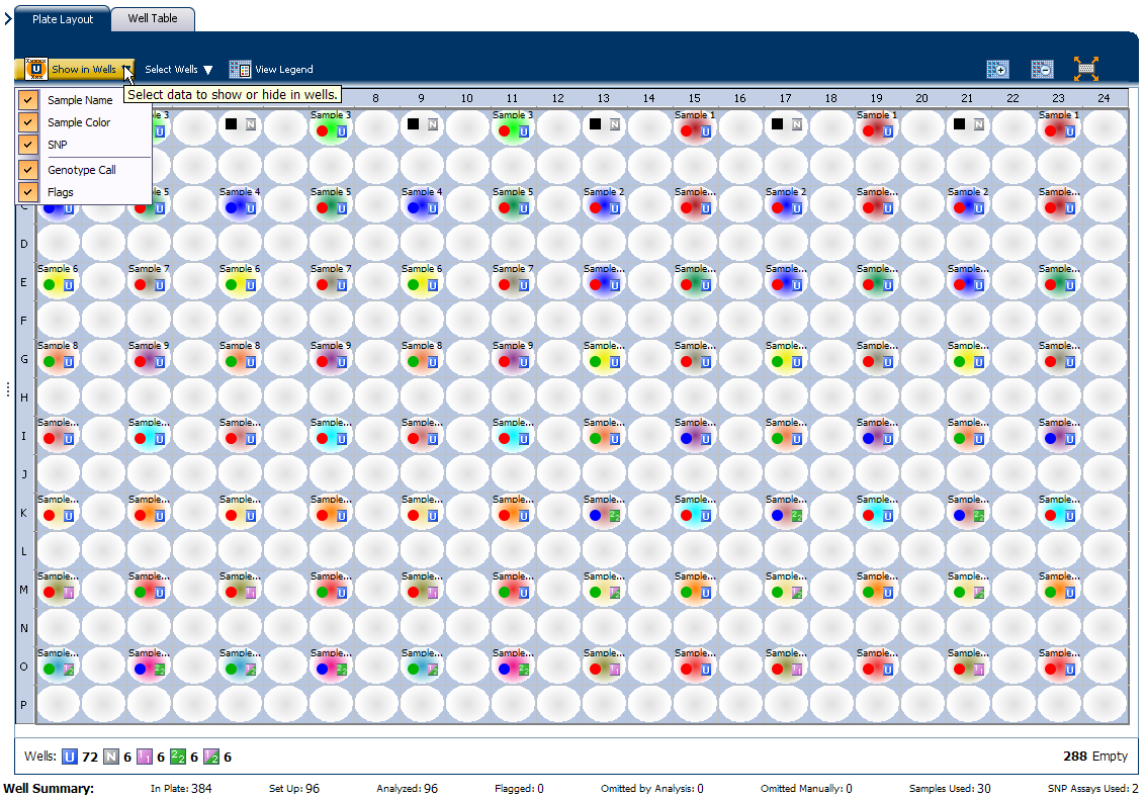
View the layout

1. Click the ❮ icon beside the Allelic Discrimination Plot to maximize the plate layout.
2. Click **Show in Wells**, then select or deselect a parameter that you want the wells to display. Repeat this step until the plate layout contains all of the desired parameters.

Parameter	Description
Sample Name	The name of the sample applied to the well.

Parameter	Description
Task	The task assigned to the well:  – Unknown  – Negative Control  – Positive Control - Allele 1  – Positive Control - Allele 2  – Positive Control - Allele 1/2
SNP Assay Name	The name of the SNP evaluated by the well.
Assay ID	The Assay ID number of the SNP evaluated by the well.
Allele 1 / Allele 2	The name of the associated allele for the SNP evaluated by the well
Allele 1 Dyes / Allele 2 Dyes	The name of the reporter and quencher dyes of the associated allele for the SNP evaluated by the well
SNP Assay Color	The color of the SNP evaluated by the well.
Sample Color / Task Color	The color of the sample or task applied to the well.
Genotype Call	The allele call assigned to the sample: • ● Homozygous 1/1 • ● Homozygous 2/2 • ● Heterozygous 1/2 • ■ Negative Control • **X** Undetermined
Flag	The number of QC flags the well triggered as listed in the ▲ symbol.

The following is an image of the plate layout of the example Genotyping experiment.



Tips for troubleshooting plate setup in your own experiment

You can adjust your view of the plate layout:

- Note the location of any samples that trigger QC flags (▲). Understanding the position of errors can aid in diagnosing any failures that may occur.
- You can select the entire reaction plate, areas of the reaction plate, or specific wells:
  - Click the upper left corner of the reaction plate to select all 384 wells.
  - Left-click the mouse and drag across the area to select it.
  - Select **Sample**, **SNP Assay**, or **Task** from the Select Wells menu in the Plate Layout tab to select wells of a specific type using the well-selection criteria.
- Use the (Zoom In), (Zoom Out), and (Fit Plate) buttons to magnify or compress the view of the wells shown.
- Use the arrow tabs to expand the plate layout to cover the entire screen.

Assess amplification results using the Amplification Plot

IMPORTANT! Amplification plots are not used to make SNP calls. Examine the plots to help with troubleshooting and quality control.

If you collected real-time data for your experiment, review the amplification data to further understand the flags triggered by the experiment data.

About amplification plots

The Amplification Plot screen displays amplification of all samples in the selected wells. Use the amplification plots to confirm the results of the experiment:

- **ΔRn vs. Cycle** – ΔRn is the difference in normalized fluorescence signal generated by the reporter between the pre-PCR read and the post-PCR read. This plot displays Δ_{Rn} as a function of cycle number. You can use this plot to identify and examine irregular amplification and to view threshold and baseline values for the run.

  Note: Viewing the ΔRn vs. Cycle plot is discussed in this booklet as an example of how to view the plot.

- **Rn vs. Cycle** – Rn is the fluorescence signal from the reporter dye normalized to the fluorescence signal from the passive reference. This plot displays Rn as a function of cycle number. You can use this plot to identify and examine irregular amplification.
- **C_T vs. Well** – C_T is the PCR cycle number at which the fluorescence meets the threshold in the amplification plot. This plot displays C_T as a function of well position. You can use this plot to locate outlying amplification (outliers).

Each plot can be viewed as a linear or log10 graph type.

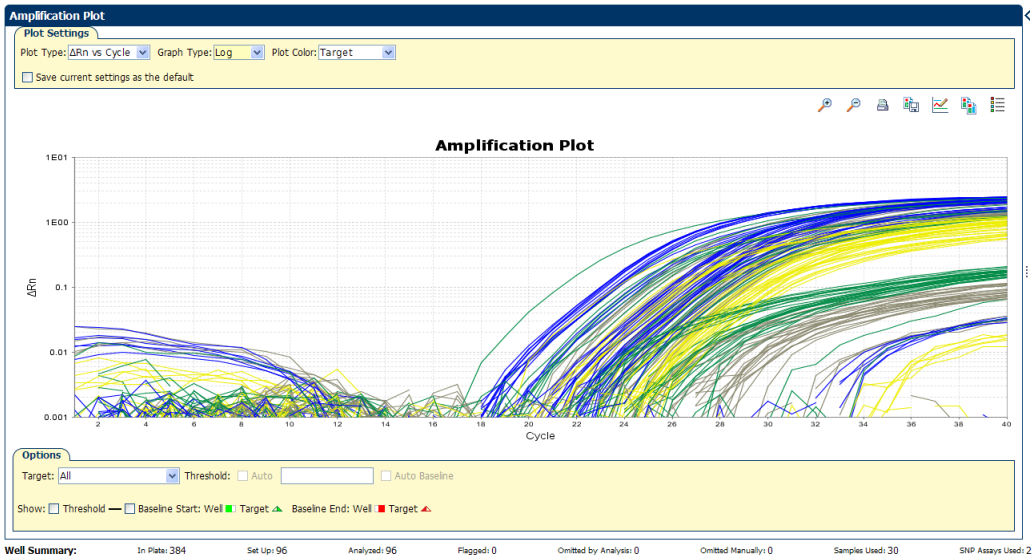
View the ΔRn vs. Cycle plot

1. From the Experiment Menu pane, select **Analysis** ▸ **Amplification Plot**.

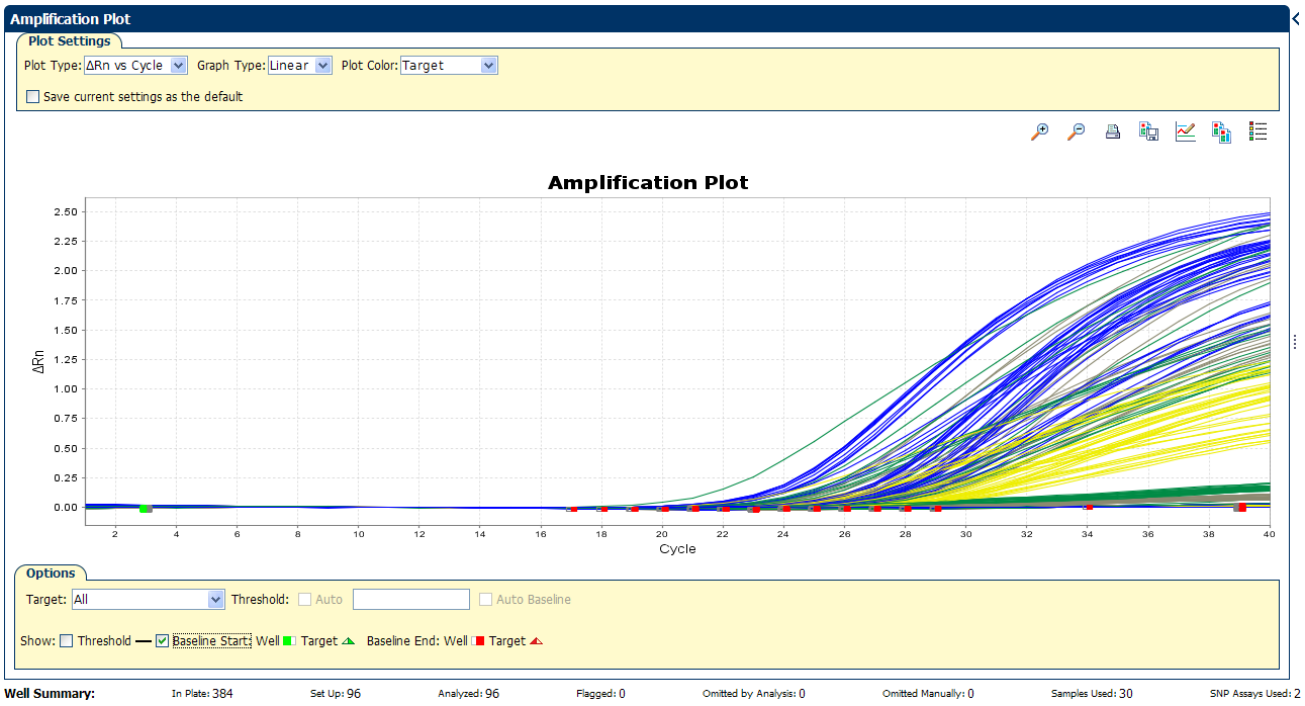
   Note: If no data are displayed, click **Analyze**.

2. Select the plot type and format:

Menu	Selection
Plot Type	ΔRn vs. Cycle
Plot Color	Target
 (This is a toggle button. When the legend is displayed, the button changes to Hide the plot legend.)	Check (default)

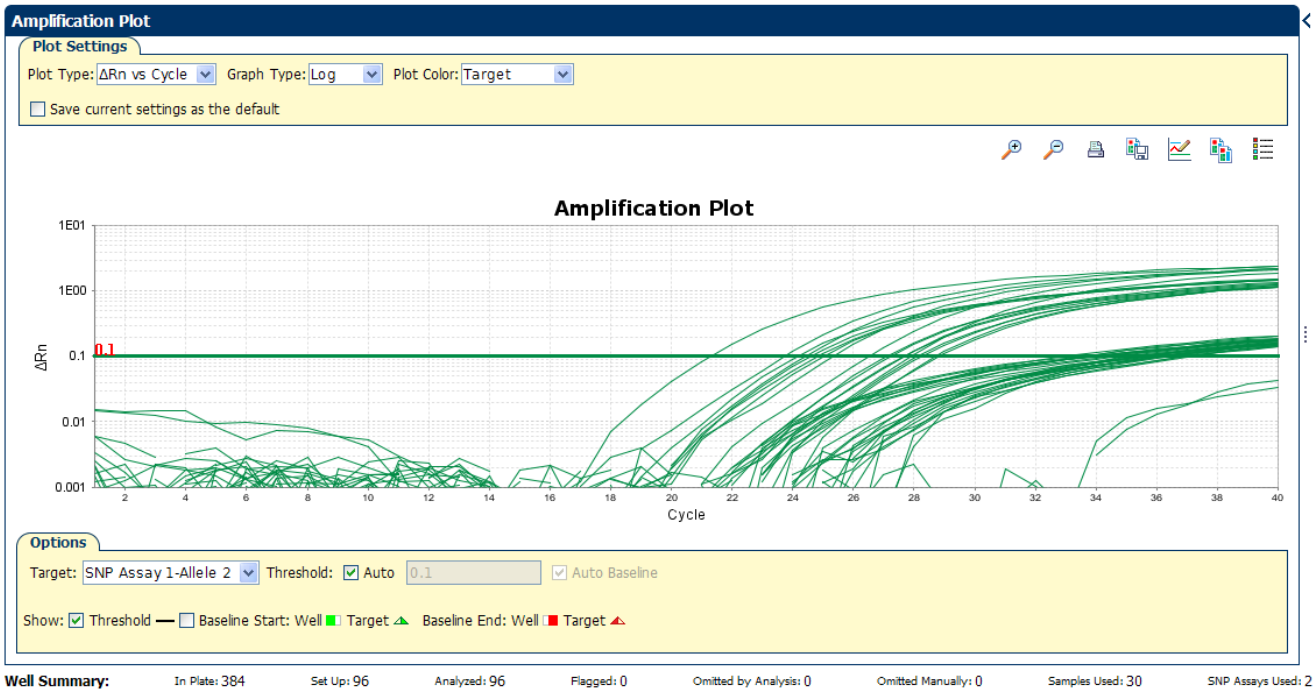


3. View the baseline values:
   a. From the Graph Type drop-down menu, select **Linear**.
   b. Select Baseline to show the start cycle and end cycle.



4. View the threshold values:
   a. From the Graph Type drop-down menu, select **Log**.
   b. From the Target drop-down menu, select **SNP Assay 1-Allele 2**.

c. Select the Threshold check box to show the threshold.



Identify well problems using the Well Table

Review the details of the experiment results in the Well Table and identify any flagged wells. The Well Table displays the assay-specific setup and analysis properties for the experiment in a tabular format.

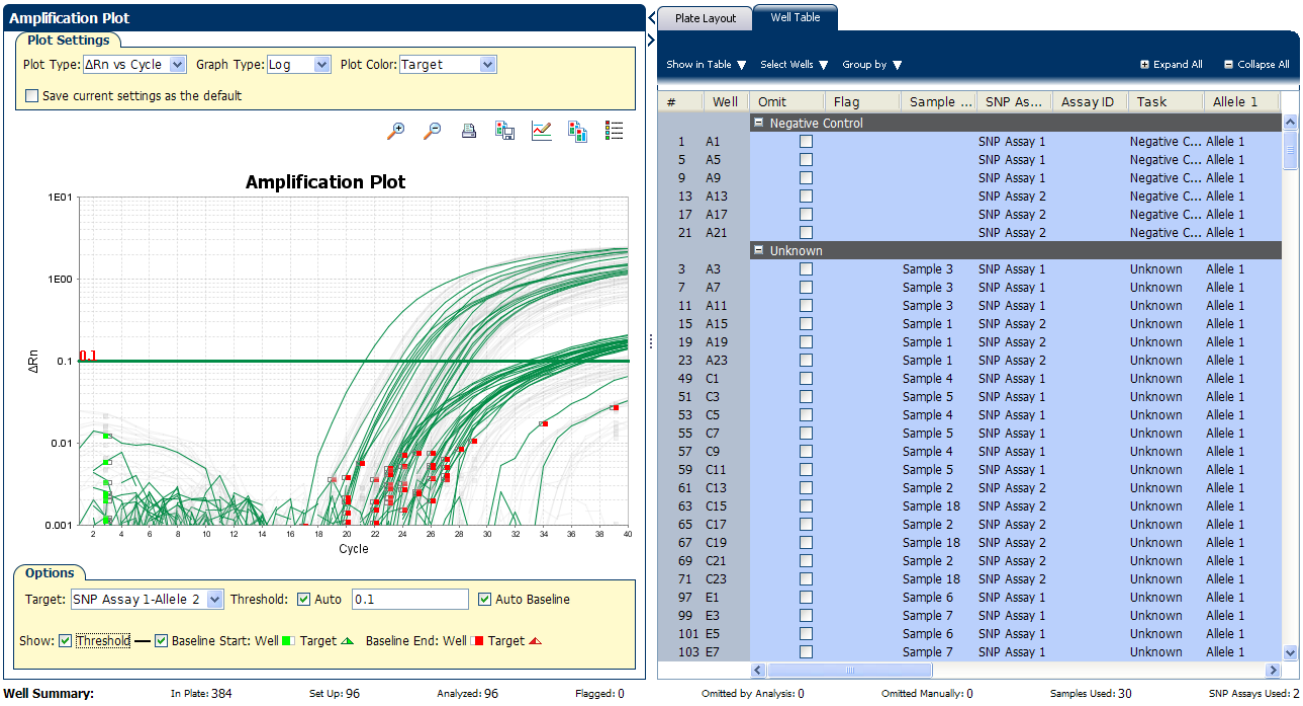
Example experiment values and flags

For the example experiment, look for wells that triggered QC flags (▲). The example experiment has no flags.

View the well table

1. Select the **Well Table** tab.
2. Click the **Flag** column header to sort the data so that the wells that triggered flags appear at the top of the table.
3. Confirm the integrity of the controls:
   a. From the Group By menu, select **Task** to organize the table rows by their function on the reaction plate.
   b. Confirm that each of the controls do not display flags (▲1).
   c. Click the ⊟ icon to collapse the negative and positive controls.

The following is an image of the well table of the example Genotyping experiment.



The following table gives the names and description of the columns in the well table:

Column	Description
Well	The position of the well on the reaction plate.
Omit	A check mark indicates that the well has been removed from the analysis.
Flag	A 1 indicates that the well triggered the number of flags listed inside the symbol.
Sample Name	The name of the sample.
SNP Assay Name	The name of the SNP assay evaluated by the well.
Assay ID	The Assay ID number of the SNP evaluated by the well.
Task	The task assigned to the well (Unknown, Negative Control, or Positive Control).
Allele 1 / 2	The name of the associated allele for the SNP evaluated by the well.
Allele 1 / 2 Dyes	The name of the reporter and quencher dyes of the associated allele for the SNP evaluated by the well.
Allele 1 / 2 $_{Rn}$	Normalized signal ($_{Rn}$) of the reporter dye of the associated allele for the SNP evaluated by the well.
Pass Ref	The signal of the passive reference dye for the well.

Column	Description
Call	The allele call assigned to the sample, where possible calls are: • ● Homozygous 1/1 - Homozygous for allele 1 • ● Homozygous 2/2 - Homozygous for allele 2 • ● Heterozygous 1/2 - Heterozygous • ■ Negative Control • ✕ Undetermined
Quality(%)	The quality value calculated for the genotype call.
Method	The method used to assign the call to the sample (Auto if assigned by the QuantStudio™ 6 and 7 Flex Real-Time PCR System Software, or Manual if applied by a user).
Comments	Comments entered for the associated sample well.
Allele 1 / 2 C_T	Threshold cycle (C_T) of the sample for the associated allele for the SNP evaluated by the well.

Identify quality control (QC) problems

The Well Table displays columns for QC flags that are triggered by the experimental data. If the experiment data does not trigger a QC flag, then the QuantStudio™ 6 and 7 Flex Real-Time PCR System Software does not display a corresponding column for the flag.

A (▲) in one of the following columns indicates that the associated well triggered the flag.

Flag	Description
BADROX	The well produced a passive reference signal greater than the limit defined in the analysis settings.
OFFSCALE	The well produced a level of fluorescence greater than the QuantStudio™ 6 or 7 Instrument can measure.
NOSIGNAL	The well did not produce a detectable level of fluorescence.
CLUSTER#	For the SNP evaluated by the well, the number of clusters generated from the experiment data is greater than the limit defined in the analysis settings.
PCFAIL	The positive control did not produce an $_{Rn}$ for the associated allele greater than the limit defined in the analysis settings indicating that the control may have failed to amplify.
SMCLUSTER	The number of data points in the associated cluster is less than the limit defined in the analysis settings.
AMPNC	The negative control has produced a $_{Rn}$ greater than the limit defined in the analysis settings indicating possible amplification.
NOAMP	The well did not produce an $_{Rn}$ for either allele that is greater than the limit defined in the analysis settings indicating that the well may have failed to amplify.
NOISE	The background fluorescence (noise) produced by the well is greater than the other wells on the reaction plate by a factor greater than the limit defined in the analysis settings.

Flag	Description
SPIKE	The amplification plot for the well contains one or more data points inconsistent with the other points in the plot.
EXPFAIL	The software cannot identify the exponential region of the amplification plot for the well.
BLFAIL	The software cannot calculate the best fit baseline for the data for the well.
THOLDFAIL	The software cannot calculate a threshold for the associated well.
CTFAIL	The software cannot calculate a threshold cycle (C_T) for the associated well.

Tips for analyzing your own experiments

Confirm the integrity of positive controls

When you analyze the example experiment or your own experiment, if you are using positive controls, confirm the integrity of the positive controls:

1. From the Group By menu, select **Task** to organize the table rows by their function on the reaction plate
2. Confirm that the positive controls do not display flags (▲) and that their normalized reporter dye fluorescence (R_n) is appropriate for the genotype (for example, if evaluating the Positive Control Allele 1/Allele 1, you would expect to see significant increase in R_n for the Allele 1 probe and very little for the Allele 2 probe).

Adjust the Well Table

- Review the data for the Unknown samples. For each row that displays (▲) in the Flag column, note the data and the flag(s) triggered by the associated well.
- Select areas of the table or wells of a specified type by:
  - Left-clicking the mouse and dragging across the area you want to select an area of the table.
  - Selecting **Sample**, **SNP Assay**, or **Task** from the Select Wells menu in the Well Table tab to select wells of a specific type using the well-selection tool.
- Group the rows of the plate layout by selecting an option from the Group By menu. You can then collapse or expand the lists either by clicking the **+/-** icon next to individual lists, or by clicking **Collapse All** or **Expand All**.
- Omit a well from the analysis by selecting the **Omit** check box for that well. To include the well in the analysis, deselect the **Omit** check box.

  Note: You must reanalyze the experiment each time you omit or include a well.

Confirm accurate dye signal using the Multicomponent Plot

The Multicomponent Plot screen displays the complete spectral contribution of each dye in a selected well over the duration of the PCR run.

Purpose

In the example experiment, you review the Multicomponent Plot screen for:

- ROX™ dye (passive reference)
- FAM™ dye (reporter)
- VIC® dye (reporter)
- Spikes, dips, and/or sudden changes
- Amplification in the negative control wells

View the Multicomponent Plot

1. From the Experiment Menu pane, select **Analysis ▸ Multicomponent Plot**.

   Note: If no data are displayed, click **Analyze**.

2. Display the unknown wells in the plate layout to display the corresponding data in the Multicomponent Plot screen:

   a. Click the **Plate Layout** tab.

   b. Select one well in the plate layout; the well is shown in the Multicomponent Plot screen.

      Note: If you select multiple wells, the Multicomponent Plot screen displays the data for all selected wells simultaneously.

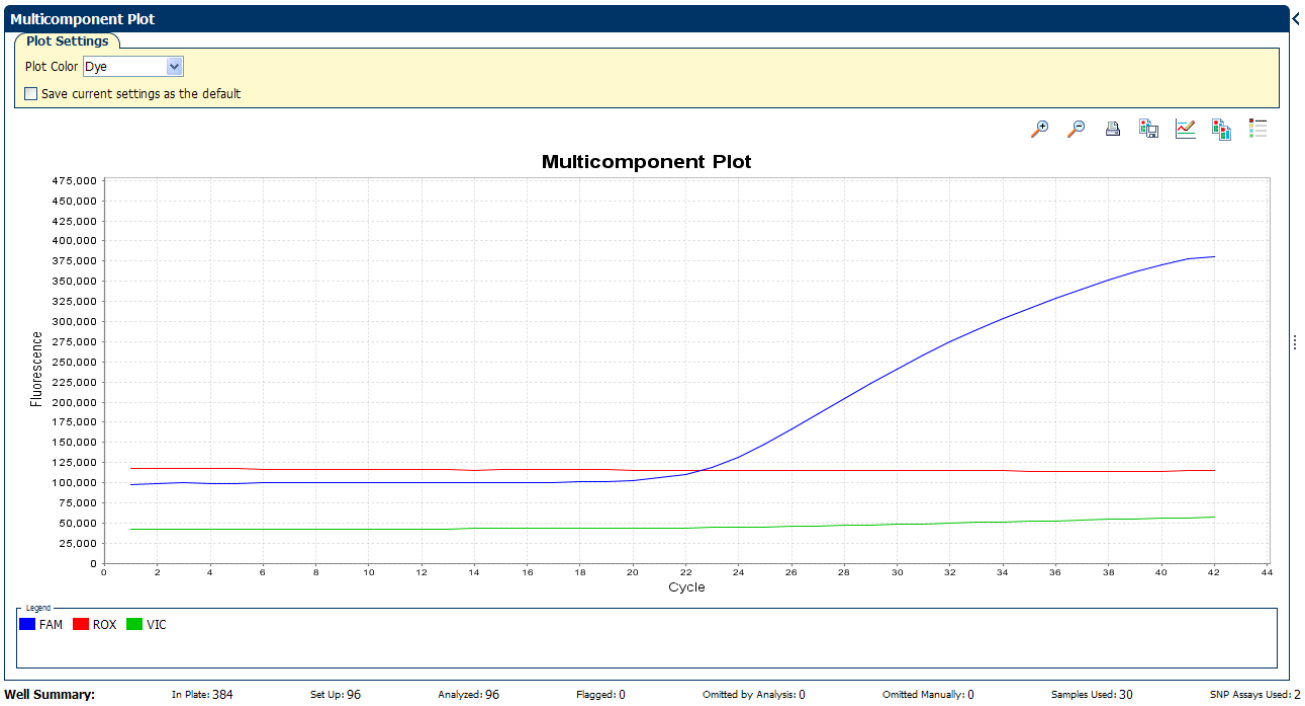
3. From the Plot Color drop-down menu, select **Dye**.

4. Click  **Show a legend for the plot** (default).

   Note: This is a toggle button. When the legend is displayed, the button changes to Hide the plot legend.

5. Check the ROX dye signal. In the example experiment, the ROX dye signal remains constant throughout the PCR process; a constant ROX dye signal indicates typical data.

6. Check the FAM dye signal. In the example experiment, the FAM dye signal increases throughout the PCR process; increase in FAM dye signal indicates normal amplification.



7. Select the negative control wells one at time and check for amplification. Wells with the negative control should not show amplification. The example experiment does not have negative controls.

Tips for confirming dye accuracy in your own experiment

When you analyze your own Genotyping experiment, look for:

- **Passive Reference** – The passive reference dye fluorescence level should remain relatively constant throughout the PCR process.
- **Reporter Dye** – The reporter dye fluorescence level should display a flat region corresponding to the baseline, followed by a rapid rise in fluorescence as the amplification proceeds.
- **Irregularities in the signal** – There should not be any spikes, dips, and/or sudden changes in the fluorescent signal.
- **Negative Control wells** – There should not be any amplification in the negative control wells.

Determine signal accuracy using the Raw Data Plot

The Raw Data Plot screen displays the raw fluorescence signal (not normalized) for each optical filter for the selected wells during each cycle of the real-time PCR.

Purpose

In the Genotyping example experiment, you review the Raw Data Plot screen for a stable increase in signal (no abrupt changes or dips) from the appropriate filter.

View the Raw Data Plot

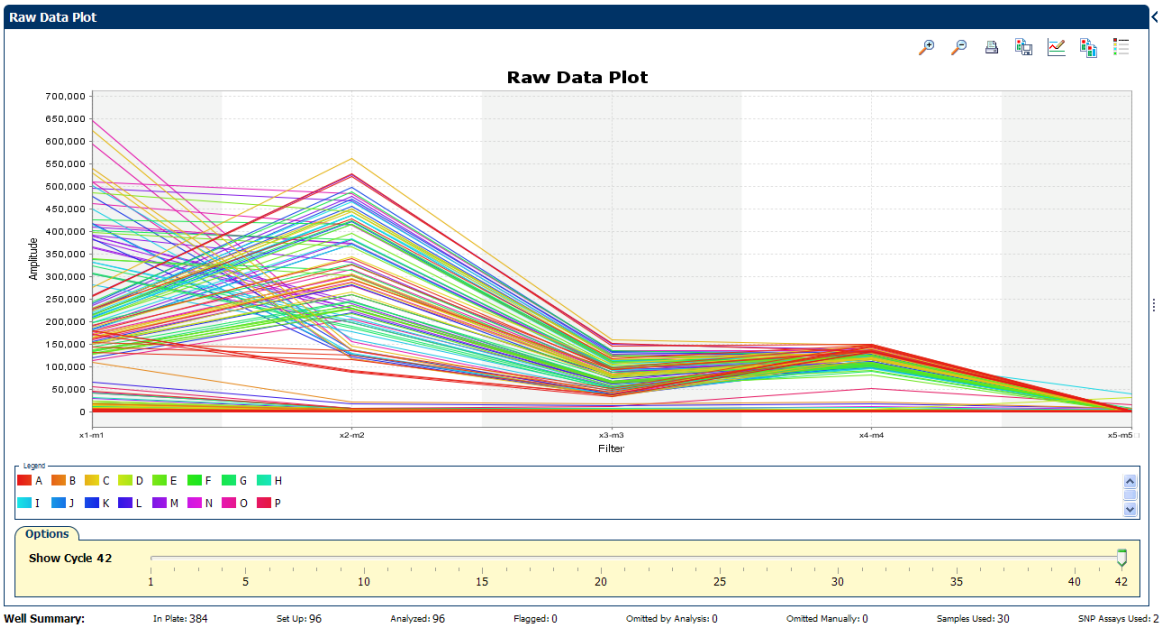
1. From the Experiment Menu pane, select **Analysis ▸ Raw Data Plot**.

   Note: If no data are displayed, click **Analyze**.

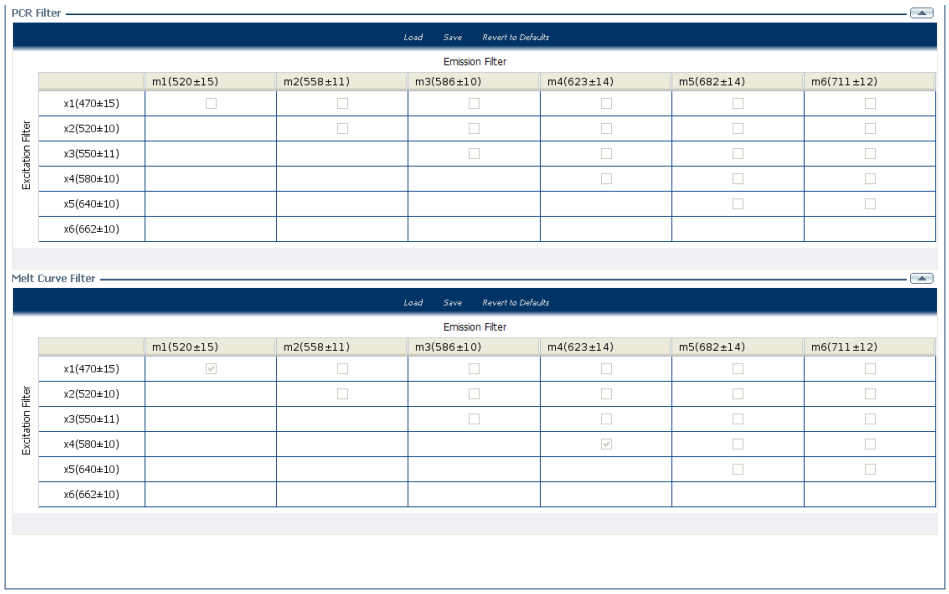
2. Display all 384 wells in the Raw Data Plot screen by clicking the upper left corner of the plate layout in the Plate Layout tab.

3. Click  **Show a legend for the plot** (default). The legend displays the color code for each row of the reaction plate (see the legend in the Raw Data Plot shown below).

4. Click and drag the Show Cycle pointer from cycle 1 to cycle 42. In the example experiment, there is a stable increase in signal from filter 1, which corresponds to the FAM™ dye filter.



The filters are:



Tips for determining signal accuracy in your own experiment

When you analyze your own Genotyping experiment, look for the following in each filter:

- Characteristic signal growth
- No abrupt changes or dips

Review the flags in the QC Summary

The QC Summary screen displays a list of the QuantStudio™ 6 and 7 Flex Real-Time PCR System Software flags, including the flag frequency and location for the open experiment.

For Genotyping experiments, flag appearance is triggered by experiment data or the assay. If a flag has been triggered by the assay, the Plate Layout does not display the ▲ icon. The flag details appear in the QC Summary.

In the example experiment, there are no flags.

View the QC Summary

1. From the Experiment Menu pane, select **Analysis** ▸ **QC Summary**.

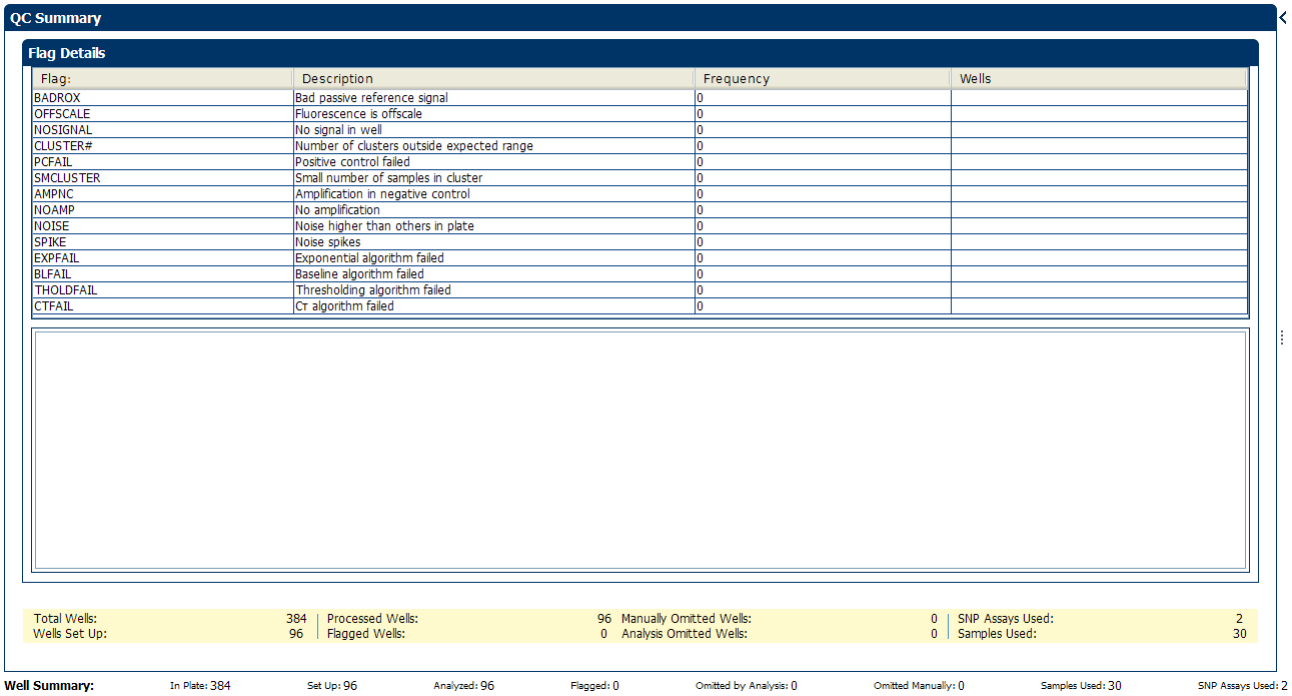
   Note: If no data are displayed, click **Analyze**.

2. Review the Flags Summary.

   Note: A 0 displayed in the Frequency column indicates that the flag does not appear in the experiment. If the frequency is >0, the flag appears somewhere in the experiment; the well position is listed in the Wells column.

3. In the Flag Details table, click each flag with a frequency >0 to display detailed information about the flag.

4. *(Optional)* For those flags with frequency >0, click each flag with a frequency >0 to display detailed information about the flag.



Possible flags

The flags listed below may be triggered by the experiment data or the assay.

Flag	Description
Pre-processing flag	
OFFSCALE	Fluorescence is offscale
Primary analysis flags	
BADROX	Bad passive reference signal
NOAMP	No amplification
NOISE	Noise higher than others in plate
SPIKE	Noise spikes
NOSIGNAL	No signal in well
EXPFAIL	Exponential algorithm failed
BLFAIL	Baseline algorithm failed
THOLDFAIL	Thresholding algorithm failed
CTFAIL	C_T algorithm failed
Secondary analysis flags	
AMPNC	Amplification in negative control
CLUSTER#	Number of clusters outside expected range
PCFAIL	Positive Control failed
SMCLUSTER#	Small number of samples in clusters

For more information

For more information on...	Refer to...	Publication number
Publishing data	Chapter 2 in Booklet 1, *Getting Started with QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Experiments*	4489822

Section 5.2 Adjust parameters for re-analysis of your own experiments

Adjust analysis settings

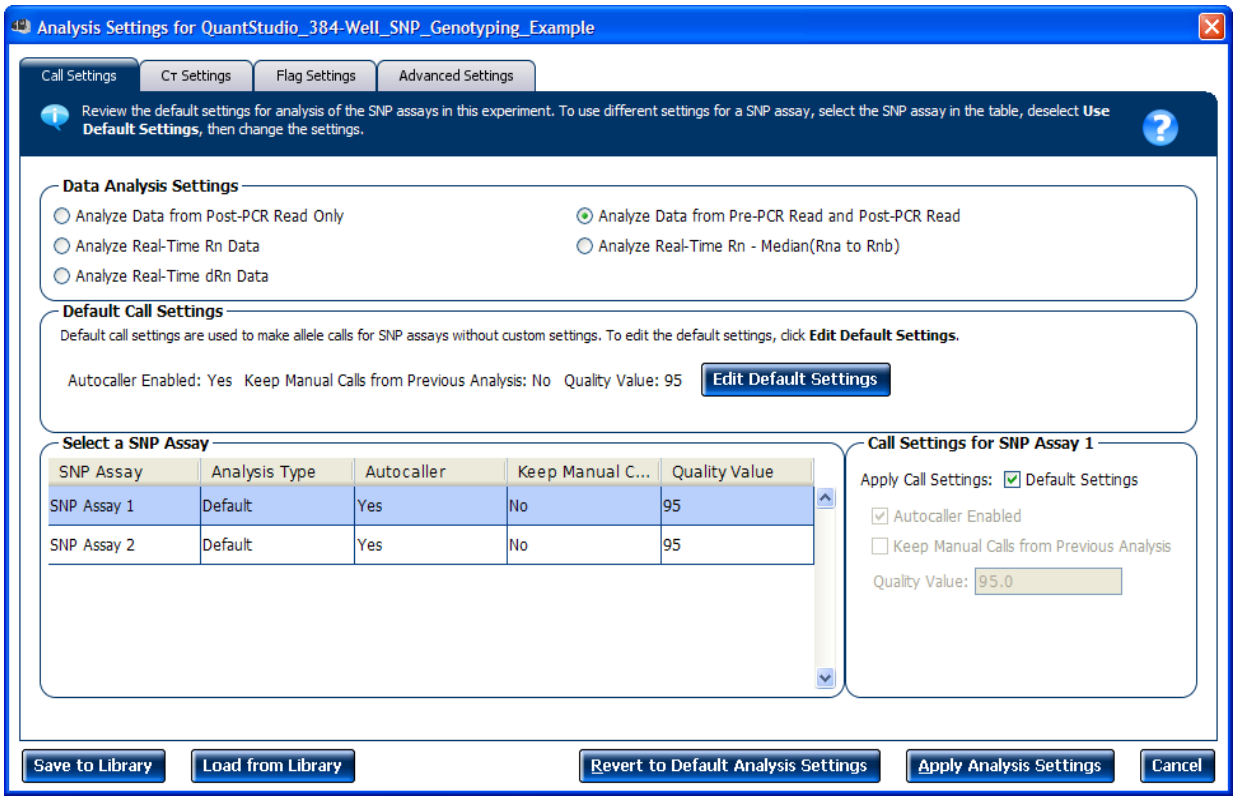
The Analysis Settings dialog box displays the analysis settings for the call, threshold cycle (C_T), flags, and advanced options.

You can change the settings in the Analysis Settings dialog box, then reanalyze your experiment.

View the analysis settings

1. From the Experiment Menu pane, select **Analysis**.
2. Click **Analysis** ▸ **Analysis Settings** to open the Analysis Settings dialog box.
   In the example experiment, the default analysis settings are used for each tab:
   - Call Settings
   - C_T Settings
   - Flag Settings
   - Advanced Settings

   The following is an image of the Analysis Settings dialog box for a Genotyping experiment:



3. View and, if necessary, change the analysis (see "Adjust analysis settings" below).

   Note: You can save the changes to the analysis settings to the Analysis Settings Library for later use. For more information, see About the Analysis Settings Library in Booklet 1, *Getting Started with QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Experiments*.

4. Click **Apply Analysis Settings** to apply the current analysis settings.

   Note: You can go back to the default analysis settings, by clicking **Revert to Default Analysis Settings**.

Adjust analysis settings

Call Settings

Use the Call Settings tab to:

- Change the default data analysis settings. You can select from:
  - Analyze data from Post-PCR Read only - Select if you do not want to use data from the pre-PCR read to determine genotype calls.
  - Analyze data from Pre-PCR Read and Post-PCR Read - If you included the pre-PCR read in the run, select if you want to use data from the pre-PCR read to determine genotype calls.
  - Analyze Real-Time Rn Data - If you included amplification in the run, select if you want to use the normalized reporter (Rn) data from the cycling stage to determine genotype calls.
  - Analyze data from Rn - Avg (Rna to Rnb) - If you included amplification in the run, select if you want to use the subtracted median of the normalized reporter (Rn) data from the cycling stage to determine genotype calls, where Rna to Rnb refers to all the cycles from the Start Cycle Number to the End Cycle Number. The average subtraction provides improved data accuracy.

    Note: To activate the Reveal Traces feature on the Allelic Discrimination Plot scree, select either **Analyze Real-Time Rn Data** or **Analyze data from Rn - Avg (Rna - Rnb)**.
- Edit the default call settings. Click **Edit Default Settings**, then specify the default settings:
  - **Autocaller Enabled** - Select for the software to make genotype calls using the autocaller algorithm.
  - **Keep Manual Calls from Previous Analysis** - If the autocaller is enabled, select to maintain manual calls after reanalysis
  - **Quality Value** - Enter a value to use to make genotype calls. If the confidence value is less than the call setting, the call is undetermined.
- Use custom call settings for a SNP assay.
  - Select one or more SNP assays in the table, then deselect the **Default Settings** check box.
  - **Define the custom call settings**.

C_T Settings

- **Data Step Selection**

  Use this feature to select one stage/step combination for C_T analysis when there is more than one data collection point in the run method.
- **Algorithm Settings**

Use the Baseline Threshold algorithm to determine the C_T values.

The Baseline Threshold algorithm is an expression estimation algorithm that subtracts a baseline component and sets a fluorescent threshold in the exponential region for gene quantification.

- **Default C_T Settings**

  Use the default C_T settings feature to calculate C_T for the alleles that do not have custom settings. To edit the default settings, click **Edit Default Settings**.

- **C_T Settings for Target**

  When you manually set the threshold and baseline, Life Technologies recommends:

Setting	Recommendation
Threshold	Enter a value for the threshold so that the threshold is: • Above the background. • Below the plateau and linear regions of the amplification curve. • Within the exponential phase of the amplification curve.
Baseline	Select the Start Cycle and End Cycle values so that the baseline ends before significant fluorescent signal is detected.

Note: This setting is applicable only to the Baseline Threshold algorithm.

Note: Selecting Automatic Threshold implies selection of automatic setting of the baseline. However, if Automatic Threshold is deselected, then you can choose between setting the baseline either automatically or manually.

Flag Settings

Use the Flag Settings tab to:

- Adjust the sensitivity so that more wells or fewer wells are flagged.
- Change the flags that are applied by the QuantStudio™ 6 and 7 Flex Real-Time PCR System Software.

  To adjust the flag settings:

To adjust the flag settings:

1. In the Use column, select the check boxes for flags to apply during analysis.
2. *(Optional)* If an attribute, condition, and value are listed for a flag, specify the setting for applying the flag.

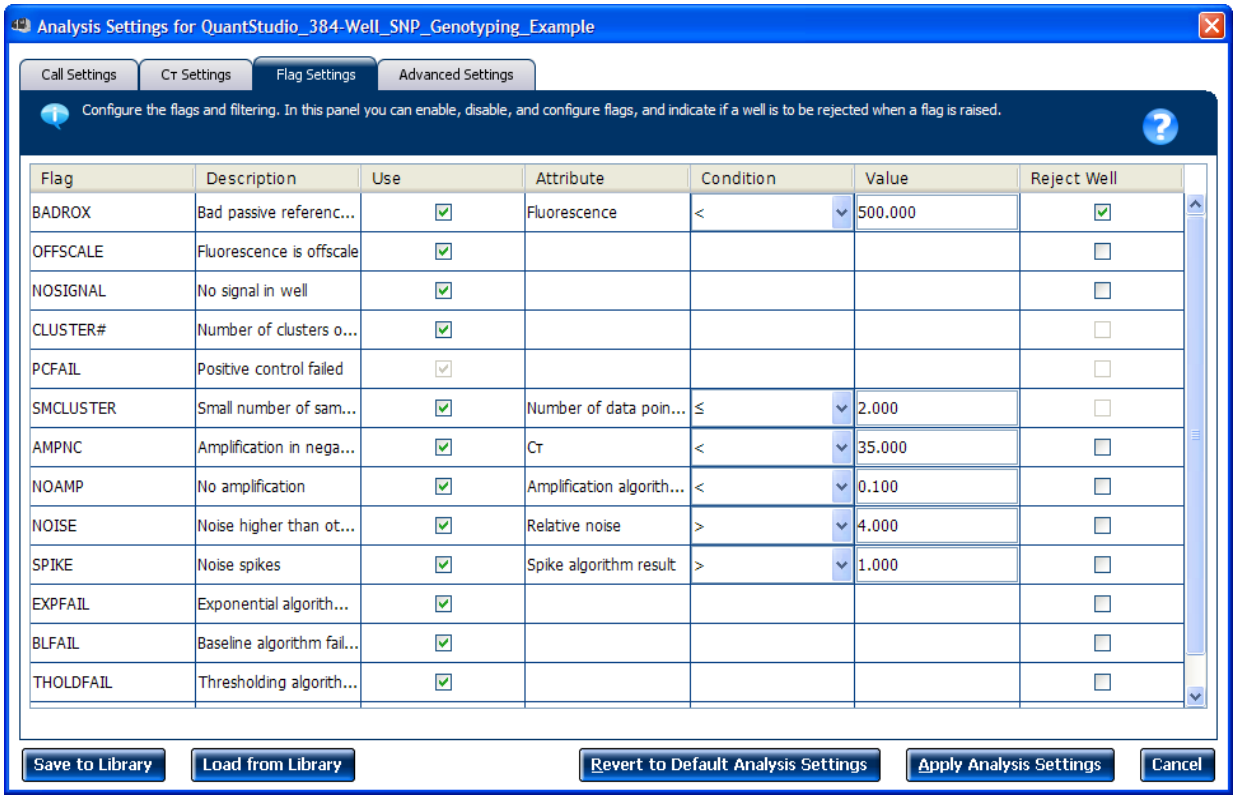
   Note: If you choose to adjust the setting for applying a flag, make minor adjustments as you evaluate the appropriate setting.

3. In the Reject Well column, select the check boxes if you want the software to reject wells with the flag.

   Note: After you have rejected the flagged wells, analysis results depend on factors such as the experiment type and flag type. For example, rejecting wells flagged by HIGHSD in experiments using the Standard Deviation calculations may change the result of C_T SD. For some flags, analysis results calculated before the well is rejected are maintained.

4. Click **Apply Analysis Settings** in the Analysis Settings dialog box. If the run status is complete, the data are reanalyzed.

The following is an image of the Flag Settings tab:



Advanced Settings

Use the Advanced Settings tab to change baseline settings well-by-well.

To use custom baseline settings for a well-target combination:

1. Select one or more well-target combinations in the table.
2. Deselect the **Use C_T Settings Defined for Target** check box.
3. Define the custom baseline settings:
   - For automatic baseline calculations, select the **Automatic Baseline** check box.
   - To define the baseline, deselect the **Automatic Baseline** check box, then enter the baseline start cycle and baseline end cycle.

For more information

For more information on...	Refer to...	Publication number
Amplification efficiency	*Amplification Efficiency of TaqMan® Gene Expression Assays Application Note*.	127AP05-03

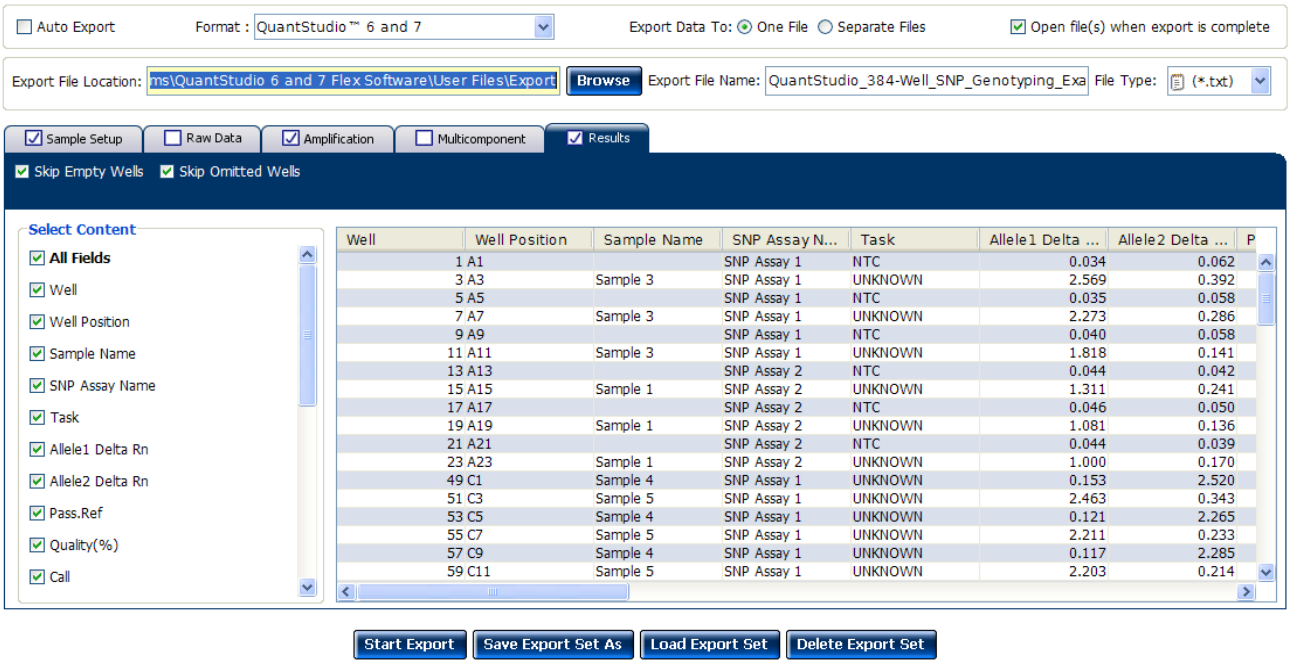
6 Export Analysis Results

1. Open the Genotyping example experiment file that you analyzed in Chapter 5.
2. In the Experiment Menu, click  **Export.**

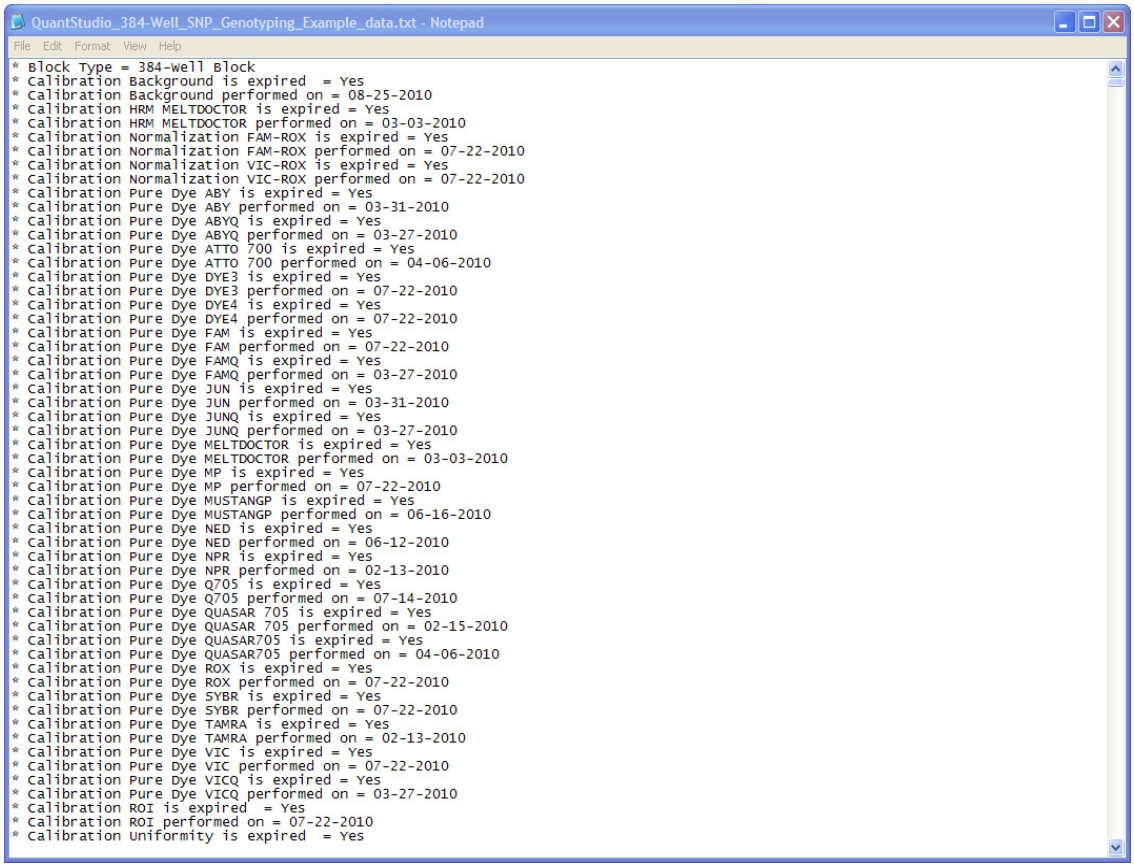
   Note: To export data automatically after analysis, select the **Auto Export** check box during experiment setup or before running the experiment. Auto export is unchecked for the example experiment.
3. Select **QuantStudio™ 6 and 7** format.
4. Complete the Export dialog box as shown below:

Field or Selection	Entry
Select Data to export/ Select Content	Results
Export Data To	One File
Export File Name	QuantStudio_384-Well_SNP_Genotyping_Example_data
File Type	*.txt
Export File Location	C:\Applied Biosystems\QuantStudio 6 and 7 Flex Software\User Files\Export

Your Export screen should look like this:



Your exported file when opened in Notepad should look like this:



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USER GUIDE

applied biosystems®
by life technologies™

QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Getting Started Guide for Presence/ Absence Experiments

Booklet 5

Publication Number 4489822
Revision A

life technologies™

Contents

1 About Presence/ Absence Experiments

This chapter covers:

IMPORTANT! First-time users of the QuantStudio™ 6 and 7 Flex Real-Time PCR System Software please read Booklet 1, *Getting Started with QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Experiments* and Booklet 7, *QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Experiments - Appendixes* of this binder thoroughly. The booklet provides information and general instructions that are applicable to all the experiments described in this binder

Note: For more information about any of the topics discussed in this guide, access the Help from within QuantStudio™ 6 and 7 Flex Real-Time PCR System Software by pressing **F1**, clicking in the toolbar, or selecting **Help ▸ QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Help**.

About data collection

Presence/Absence experiments are end-point experiments that are performed to detect a target nucleic acid sequence in a sample.

You can collect the experiment data at the end of the run or continuously in real time.

End-point PCR Data

The QuantStudio™ 6 and 7 Instruments collect data at an end-point, that is after the process has completed.

The data collected is the normalized intensity of the reporter dye, or Rn.

Note: Some end-point experiments also include pre-PCR (data collected before the amplification process) datapoints. If so, the software calculates the delta Rn (ΔRn) value per the following formula:

ΔRn = Rn (post-PCR read) – Rn (pre-PCR read), where Rn = normalized readings.

Real-Time PCR Data

The QuantStudio™ 6 and 7 Instruments provide the option of collecting real-time data, during the PCR process.

Note: Real-time data collection is used only for troubleshooting, and not for Presence/Absence analysis.

Setting up PCR reactions

With Presence/Absence experiments, you prepare PCR reactions that contain primers and probes to amplify the target and a reagent to detect amplification of the target. You can set up the PCR reactions for the Presence/Absence experiments two different ways.

Note: The example experiment uses IPC setup for setting up the PCR reactions.

IPC setup

Use an internal positive control (IPC) to monitor the PCR progress and ensure that a negative result is not caused by failed PCR in the sample. PCR reactions contain two primer/probe sets: One to detect the unknown target (unknown target primer set and TaqMan® probe to detect the unknown target) and one to detect the IPC (IPC primer set and a VIC dye-labeled TaqMan® probe to detect the IPC template). With this setup, there are three well types:

- **Unknown-IPC wells** contain sample template and IPC template; the presence of the target is not known.
- **Negative control-IPC wells** contain IPC template and water or buffer instead of sample template in the PCR reaction. Only the IPC template should amplify in negative control-IPC wells because the reaction contains no sample template. Also called **IPC+**.
- **Negative control-blocked IPC wells** do not contain sample template in the PCR reaction. Amplification is prevented by a blocking agent. As a result, no amplification should occur in negative control-blocked IPC wells because the reaction contains no sample template and amplification of the IPC is blocked. Negative control-blocked IPC is called *no amplification control (NAC)*.

  If the run method includes amplification, real-time data are plotted in an amplification plot.

No IPC, singleplex setup

Omit the IPC from your Presence/Absence experiment. PCR reactions contain one primer/probe set. PCR reactions do not contain the IPC. With this setup, there are two well types:

- **Unknown wells** – Wells contain sample template; the presence of the target is not known.
- **Negative controls** – Wells contain water or buffer instead of sample template.

About the instrument run

With Presence/Absence experiments, the instrument runs can include:

- **Pre-PCR read** – Perform the pre-PCR read on the QuantStudio™ 6 and 7 Flex Real-Time PCR System Software before PCR amplification to collect baseline fluorescence data.
- **Amplification** – Perform amplification on the QuantStudio™ 6 and 7 Flex Real-Time PCR System Software to collect fluorescence data during PCR amplification. If you do not include amplification in the run method, perform amplification on another instrument.
- **Post-PCR read** – To determine the results for Presence/Absence experiments, perform the post-PCR read on the instrument after PCR amplification to collect endpoint fluorescence data.

Fluorescence data collected during the instrument run are stored in an experiment data file (.eds).

About the analysis

Data from the instrument run are used to determine Presence/Absence calls. Results are plotted in a Presence/Absence plot. If the experiment includes amplification, results are plotted in an amplification plot.

- **Pre-PCR read** – If included, the data collected from the pre-PCR read can be used to normalize data collected from the post-PCR read.
- **Amplification** – If included, the data collected from the amplification can be used to troubleshoot.
- **Post-PCR read** – The data collected from the post-PCR read are used to make Presence/Absence calls:
  - **Presence** – The target amplified above the target's threshold. The target is present in the sample.
  - **Absence** – The target did not amplify above the target's threshold. The target is absent in the sample.
  - **Unconfirmed** – The data collected is below the target threshold, and the intensity of IPC is below the IPC threshold.

With the IPC setup, the data collected form the post-PCR read are used to make the following calls:

- **IPC Failed** – The IPC target did not amplify in the IPC wells and/or the IPC target amplified in the blocked IPC wells.
- **IPC Succeeded** – The IPC target amplified in the IPC wells and the IPC target did not amplify in the blocked IPC wells.

About the example experiment

To illustrate how to perform Presence/Absence experiments, this guide leads you through the process of designing and performing an example experiment. The example experiment represents a typical setup that you can use to quickly familiarize yourself with a QuantStudio™ 6 and 7 Flex Real-Time PCR System Software.

The objective of the Presence/Absence example experiment is to determine if a pathogen is present or absent in each batch of ground beef.

In the Presence/Absence example experiment:

- DNA is extracted from samples using the PrepMan® Ultra Sample Preparation Reagent (Part no. 4318930). The DNA is extracted from each of the four samples of ground beef or from the bacteria found in the ground beef.
- The target is a pathogen.
- The experiment is designed for duplex PCR, where each reaction contains two primer/probe sets. One set detects the pathogen sequence, TGFB (primer set and FAM™ dye-labeled probe to detect the TGFB sequence). The other primer/probe set detects the IPC primer set and VIC® dye-labeled TaqMan® probe detects the IPC template.

2 Design the Experiment

This chapter explains how to design the example experiment from the Experiment Setup menu.

This chapter covers:

Note: To automatically export the analyzed data to a specified location, select the Auto Export task from the Export screen, before running the experiment. For more information on Auto Export, refer to Chapter 2 in Booklet 1, *Getting Started with QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Experiments*.

Define the experiment properties

Click **Experiment Setup** ▸ **Experiment Properties** to create a new experiment in the QuantStudio™ 6 and 7 Flex Software. Enter:

Field or Selection	Entry
Experiment Name	QuantStudio_384-Well_Presence-Absence_Example
Barcode	Leave field empty
User Name	Example User
Comments	Presence/Absence example
Instrument type	QuantStudio™ 6 Flex System
Block	384-Well Block
Experiment Type	Presence/ Absence
Reagents	TaqMan® Reagents
Ramp speed	Standard
Reagent information	NA

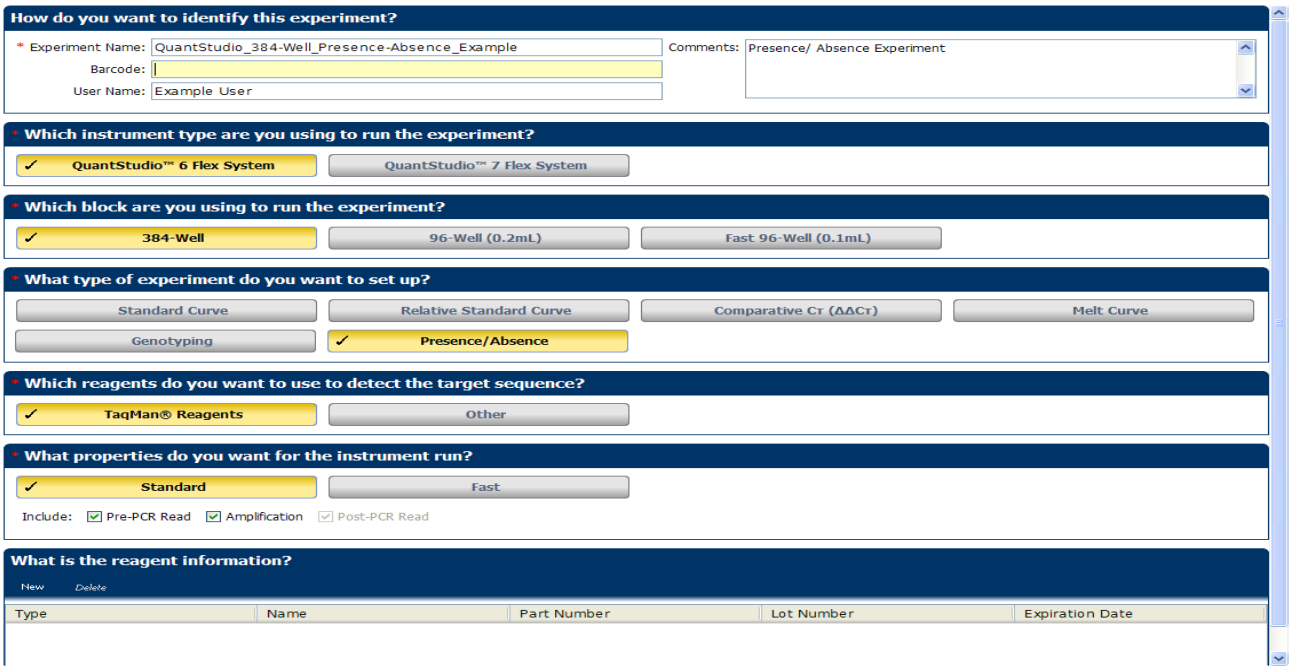
Select all three data-collection check boxes: Pre-PCR, Amplification, and Post-PCR collection methods:

Pre-PCR Read	Checked

Amplification	Checked
Post-PCR Read	Checked

Save the experiment.

Your Experiment Properties screen should look like this:



Define targets and samples

Click **Define** to access the Define screen. Enter:

1. Targets

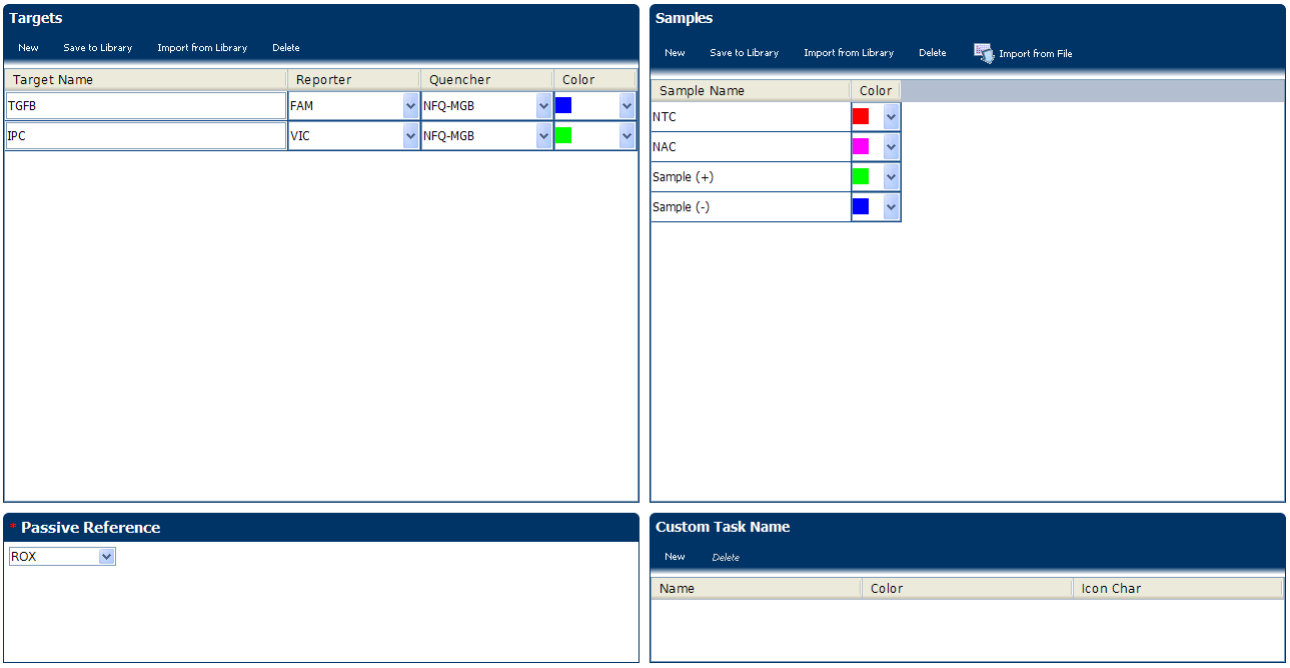
Target Name	Reporter	Quencher	Color
TGFB	FAM	NFQ-MGB	
IPC	VIC	NFQ-MGB	

2. Samples

Sample Name	Color
NTC	
NAC	
Sample (+)	
Sample (-)	

3. Dye to be used as a Passive Reference
   ROX
4. Custom Task Name
   Not applicable

Your Define screen should look like this:

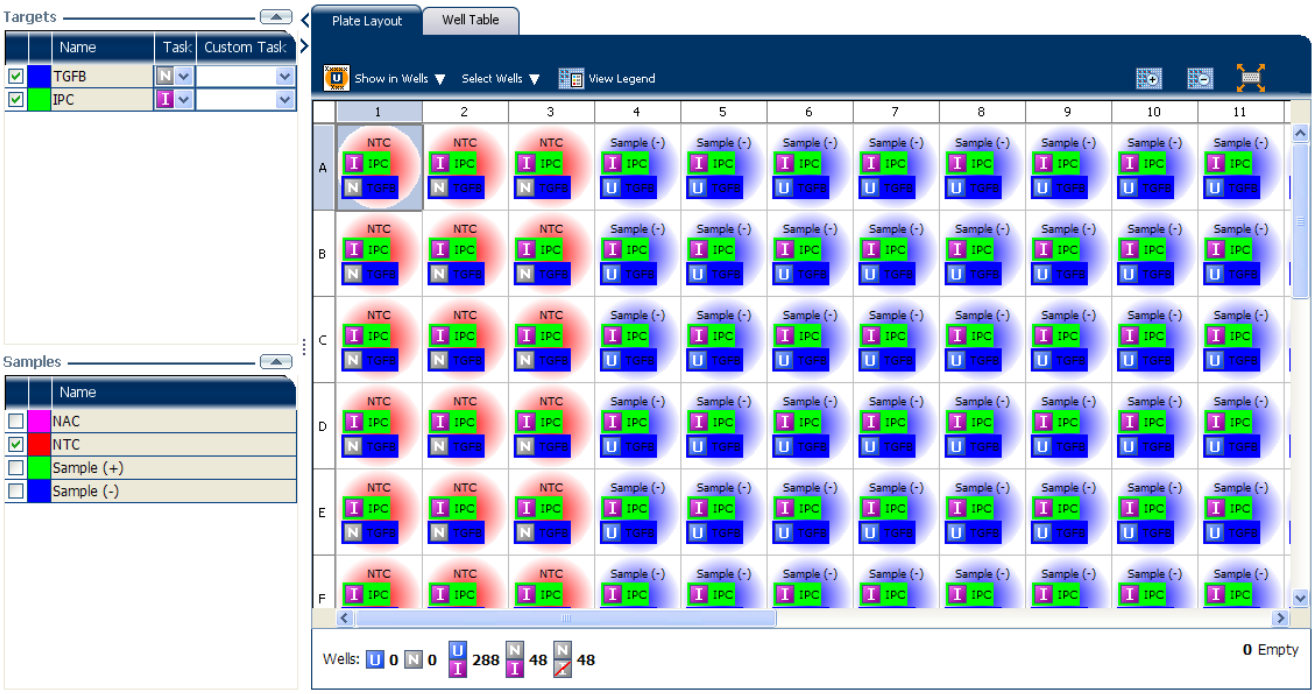


Assign targets and samples

Click **Assign** to access the Assign screen. Enter the targets and samples:

Target Name	Well Number	Task	Sample
TGFB IPC	A1 - P3 (Columns 1 - 3)	Negative IPC	NTC
TGFB IPC	A4 - P12 (Columns 4 - 12)	Unknown IPC	Sample (-)
TGFB IPC	A13 - P15 (Columns 13 - 15)	NTC Blocked IPC	NAC
TGFB IPC	A16 - P24 (Columns 16 - 24)	Unknown IPC	Sample (+)

Your Assign screen should look like this:



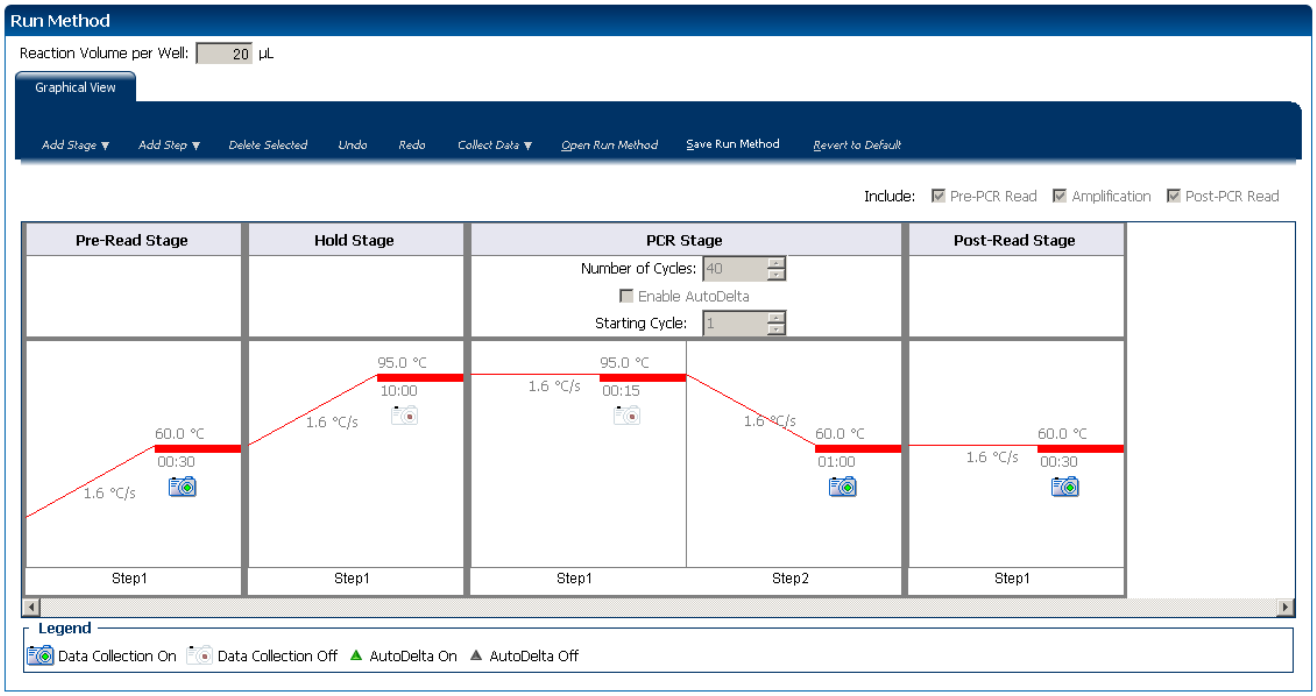
Set up the run method

Click **Run Method** to access the Run Method screen. Set the thermal profile under the Graphical View tab. Enter:

- Reaction Volume Per Well: 20 µL
- Thermal Profile

Stage	Step	Ramp rate	Temperature	Time
Pre-Read Stage	Step 1	1.6°C/s	60°C	30 seconds
Hold Stage	Step 1	1.6°C/s	95°C	10 minutes
PCR Stage • Number of Cycles: 40 (default) • Enable AutoDelta: Unchecked (default) • Starting Cycle: Disabled when Enable AutoDelta is unchecked	Step 1	1.6°C/s	95°C	15 seconds
	Step2	1.6°C/s	60°C	1 minute
Post-Read Stage	Step 1	1.6°C/s	60°C	30 seconds

Your Run Method screen should look like this:



For more information

For more information on...	Refer to	Publication number
Consumables	Chapter 3 in Booklet 1, *Getting Started with QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Experiments* Appendix A in Booklet 7, *QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Experiments - Appendixes*	4489822
Data collection	Chapter 2 in Booklet 1, *Getting Started with QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Experiments*	4489822
Amplification efficiency	*Amplification Efficiency of TaqMan® Gene Expression Assays Application Note*	127AP05-03
Using Alternative Setup	Chapter 3 in Booklet 1, *Getting Started with QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Experiments*	4489822

3 Prepare the Reactions

This chapter explains how to prepare the PCR reactions for the Presence/Absence example experiment.

This chapter covers:

Assemble required materials

- Items listed in Booklet 1, *Getting Started with QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Experiments*
- Samples - DNA extracted from ground beef (100 ng/µL)
- Example experiment reaction mix components:
  - TaqMan® Universal PCR Master Mix
  - 10X IPC Mix
  - 50X IPC DNA
  - 20X Primer/ Probe Mix

Note: Fast Universal Master Mix is not recommended to be used with the 96-well (0.2 mL) reaction plates or reaction tubes and tube strips sealed with caps.

Prepare the reaction mix (“cocktail mix”)

For the Presence/ Absence example experiment, four cocktail mixes are used; one each for:

- Sample (+)
- Sample (-)
- NTC/ IPC+
- NAC/IPC-

The following tables list the universal assay conditions (volume and final concentration) for using the TaqMan® Universal PCR Master Mix for the four cocktail mixes.

Cocktail Mix	Reaction component	Volume for 1 reaction (µL)	Volume for 160 reactions (144 wells + 10% excess) (µL)
Cocktail Mix 1 for Sample (+)	TaqMan® Universal PCR Master Mix (2.0X)	12.50	2000.0
	10X IPC Mix	2.50	400.0
	50X IPC DNA	0.50	80.0
	20X Primer/ Probe Mix	1.25	200.0
	Water/ Buffer	5.75	920.0
	Diluted unknown 1	2.5	400
	Total reaction mix volume	25.0	4000.0
Cocktail Mix 2 for Sample (-)	TaqMan® Universal PCR Master Mix (2.0X)	12.50	2000.0
	10X IPC Mix	2.50	400.0
	50X IPC DNA	0.50	80.0
	20X Primer/ Probe Mix	1.25	200.0
	Water/ Buffer	5.75	920.0
	Diluted unknown 2	2.5	400
Cocktail Mix 3 for NTC/ IPC+	TaqMan® Universal PCR Master Mix (2.0X)	12.50	662.5
	10X IPC Mix	2.50	132.5
	50X IPC DNA	0.50	26.5
	20X Primer/ Probe Mix	1.25	66.25
	Water/ Buffer	8.25	304.75
	Total reaction mix volume	25.0	1325.0
	TaqMan® Universal PCR Master Mix (2.0X)	12.50	662.5
Cocktail Mix 4 for NAC/ IPC-	TaqMan® Universal PCR Master Mix (2.0X)	12.50	662.5
	10X IPC Mix	2.50	132.5
	50X IPC DNA	0.50	26.5
	20X Primer/ Probe Mix	1.25	66.25
	IPC Block	2.5	132.5
	Water/ Buffer	5.75	304.75
	Total reaction mix volume	25.0	1325.0

To prepare the reaction mix for each of the four types:

1. Label four appropriately sized tubes for the reaction mixes: **Sample (+), Sample (-), NTC, NAC.**
2. Add the required volumes of each cocktail mix component to the tube.

3. Mix the cocktail thoroughly by gently pipetting up and down several times, then cap the tube.
4. Centrifuge the tube briefly to remove air bubbles.
5. Place the cocktail mix on ice until you prepare the reaction plate.

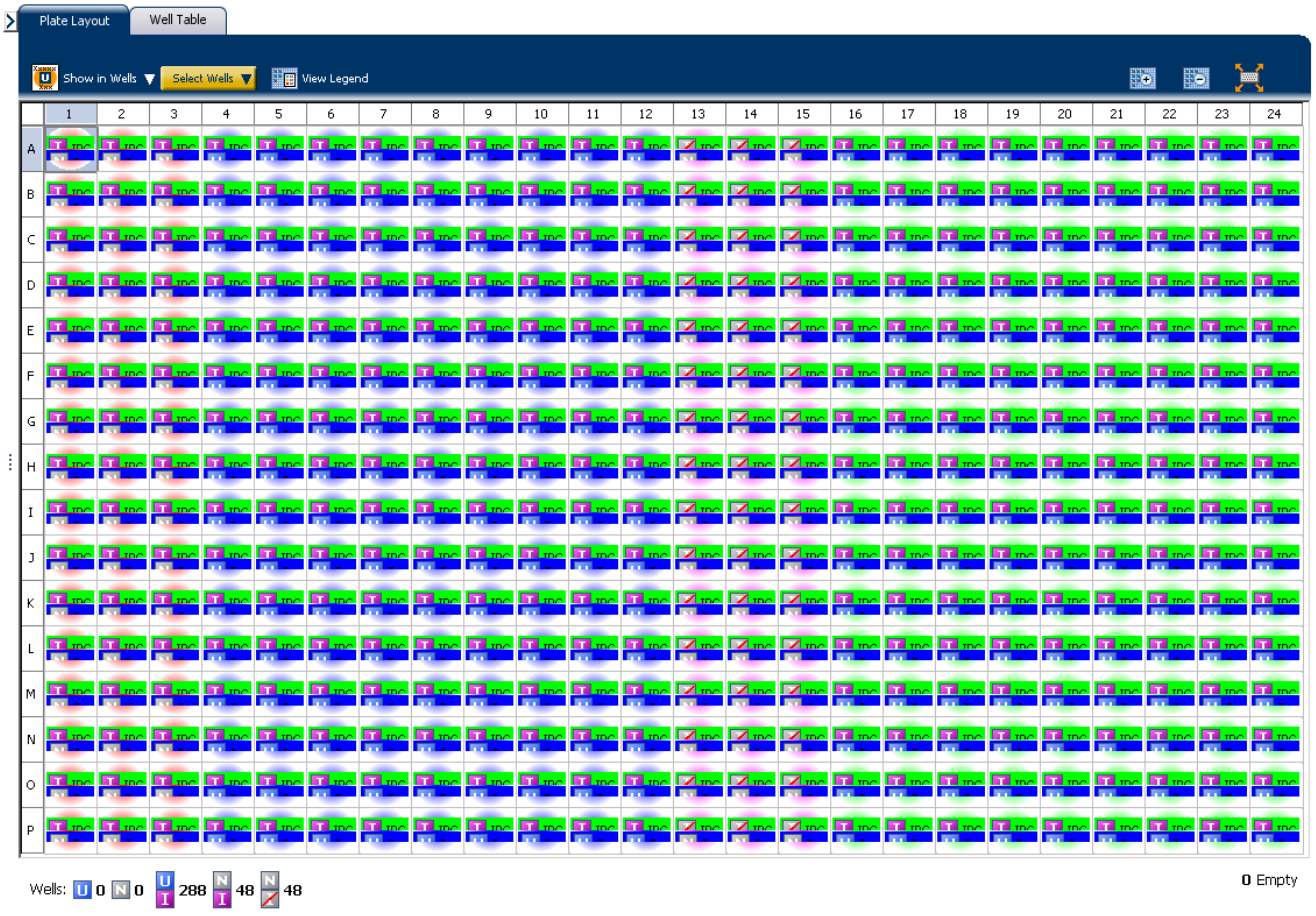
   Note: You can separately add the sample to the reaction plate, as opposed to preparing individual reaction mixes for each sample.

Prepare the reaction plate

The reaction plate for the Presence/Absence example experiment contains:

- A MicroAmp® Optical 384-Well Reaction Plate
- Reaction volume of 20 µL/well
- 144 Sample (+) wells
- 144 Sample (-) wells
- 48 NTC/ IPC+
- 48 NAC/IPC-

The following is an image of the plate layout:



To prepare the reaction plate:

1. Add 25 µL of Cocktail mix 1 to wells A16–P24.
2. Add 25 µL of Cocktail mix 2 to wells A4–P12.
3. Add 25 µL of Cocktail mix 3 to wells A13–P15.
4. Add 25 µL of Cocktail mix 4 to wells A1–P3.
5. Seal the reaction plate with optical adhesive film.
6. Centrifuge the reaction plate briefly to remove air bubbles.
7. Confirm that the liquid is at the bottom of each well of the reaction plate. If not, centrifuge the reaction plate again at a higher speed and for a longer period of time.
8. Until you are ready to perform the run, place the reaction plate at 4°C, in the dark.

For more information

For more information on...	Refer to...	Publication number
Assigning the reaction plate components	Chapter 2 in Booklet 1, *Getting Started with QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Experiments*	4489822
Sealing the reaction plate	Chapter 2 in Booklet 1, *Getting Started with QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Experiments*	4489822

4 Run the Experiment

This chapter explains how to run the example experiment on the QuantStudio™ 6 or 7 Instrument.

This chapter covers:

IMPORTANT! Run the experiment at the same ambient temperature at which you calibrated the instrument. Extreme variations in ambient temperature can affect the heating and cooling of the instrument and influence experimental results.

IMPORTANT! Do not attempt to open the access door during the run. The door is locked while the QuantStudio™ 6 or 7 Instrument is in operation.

Start the run

1. Open the Presence/Absence example file that you created using instructions in Chapter 2.
2. Load the reaction plate into the instrument.
3. Start the run.

Monitor the run

Monitor the example experiment run:

- From the QuantStudio™ 6 and 7 Flex Real-Time PCR System Software using the Run screen, while the experiment is in progress.
- From the Instrument Console of the QuantStudio™ 6 and 7 Flex Real-Time PCR System Software (to monitor an experiment started from another computer or from the QuantStudio™ 6 or 7 Instrument touchscreen).
- From the QuantStudio™ 6 or 7 Instrument touchscreen.

From the Instrument Console of the QuantStudio™ 6 and 7 Flex Real-Time PCR System Software

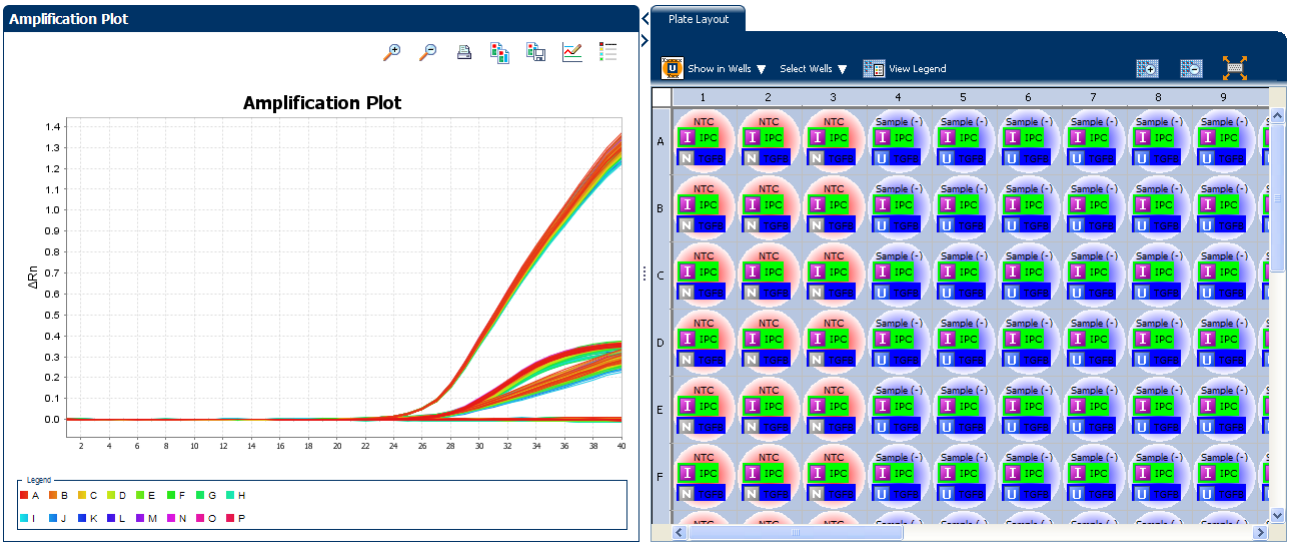
1. In the Instrument Console screen, select the instrument icon.
2. Click **Manage Instrument** or double-click on the instrument icon.
3. On the Manage Instrument screen, click **Monitor Running Instrument** to access the Run screen.

View the Amplification Plot

You can view the progress of the run in real time. During the run, periodically view all the three plots available from the QuantStudio™ 6 and 7 Flex Real-Time PCR System Software for potential problems.

Click **Amplification Plot** from the Run Experiment Menu, select the Plate Layout tab, then select the wells to view.

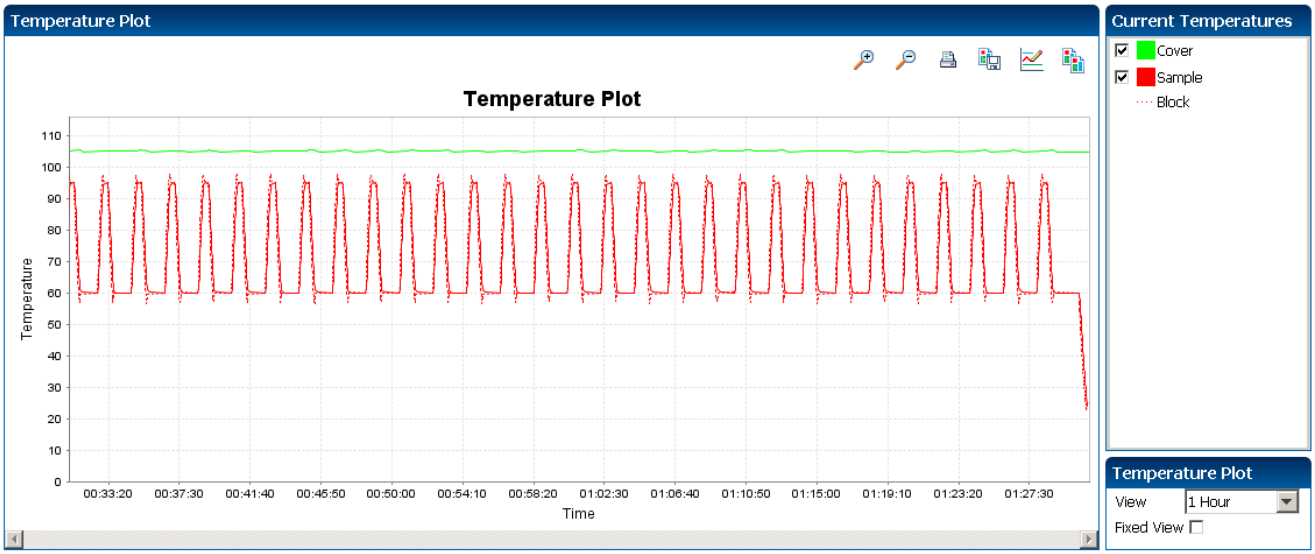
The following is an image of the Amplification Plot screen as it appears at the end of the example experiment.



View the Temperature Plot

Click **Temperature Plot** from the Run Experiment Menu.

The following is an image of the Temperature Plot screen as it appears during the example experiment.

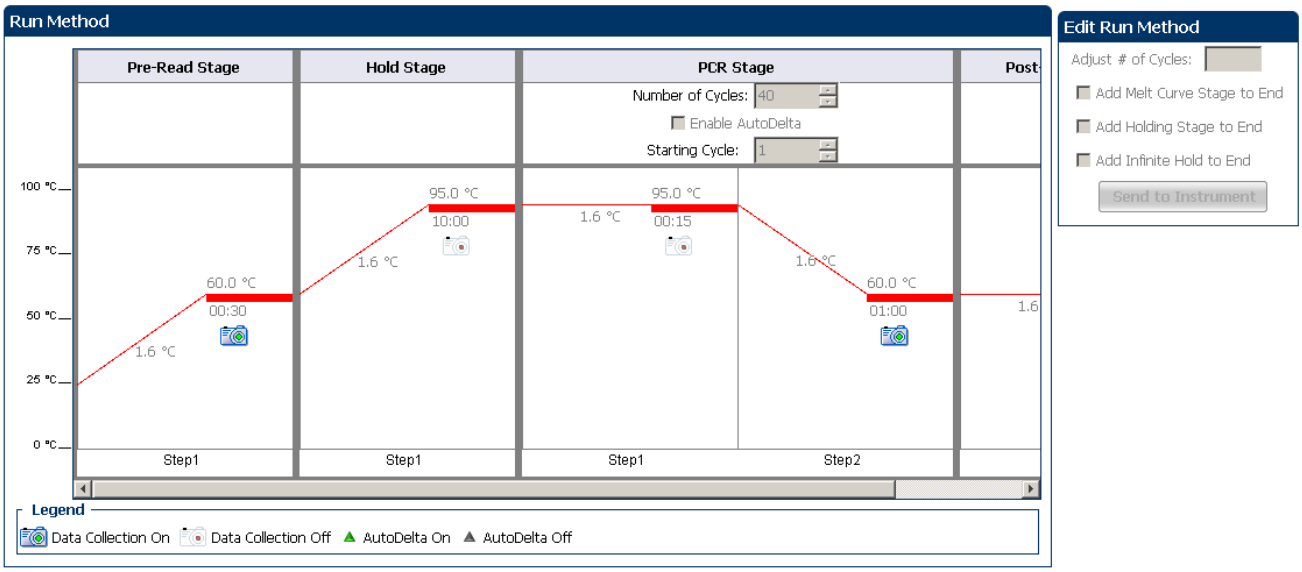


Note: The Sample temperature displayed in the Current Temperatures group is an estimated value.

View the Run Method

Click **Run Method** from the Run Experiment Menu.

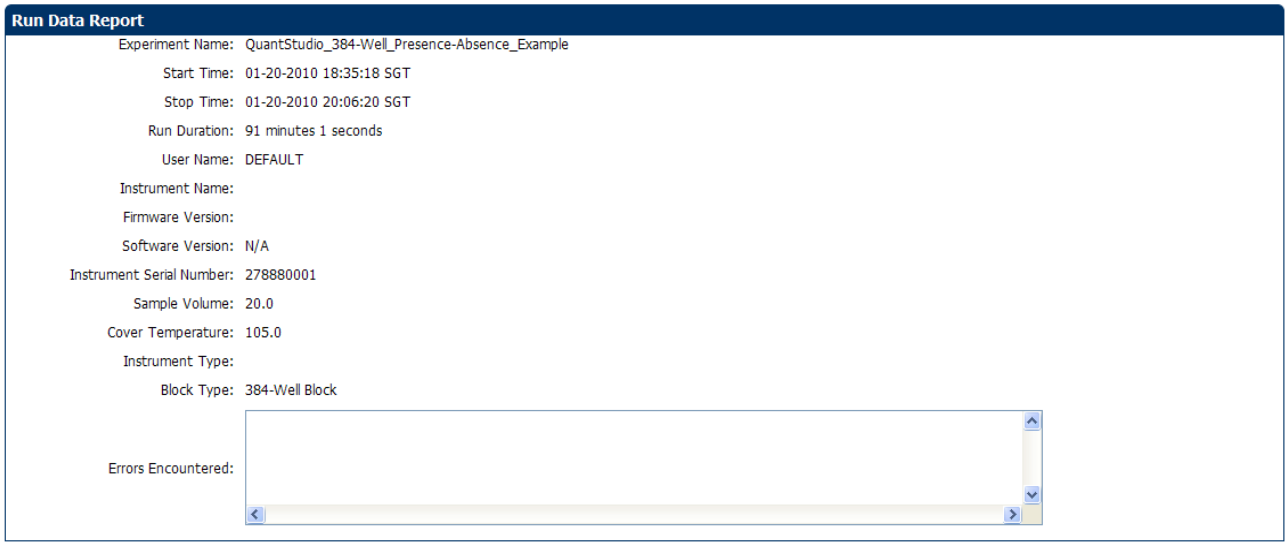
The following is an image of the Run Method screen as it appears in the example experiment.



View run data

Click **View Run Data** from the Run Experiment Menu.

The following is an image of the View Run Data screen as it appears in the example experiment.

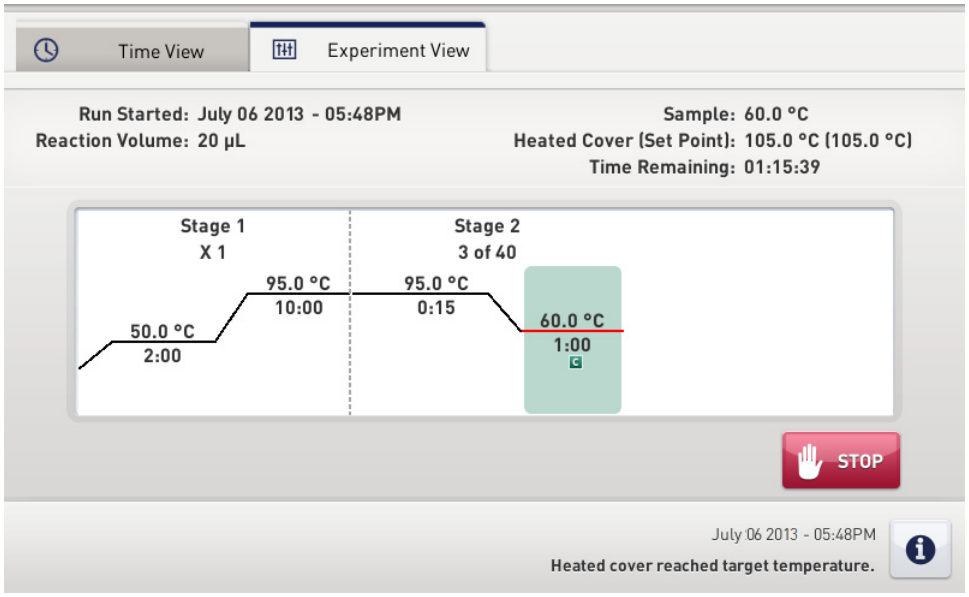


From the QuantStudio™ 6 or 7 Instrument touchscreen

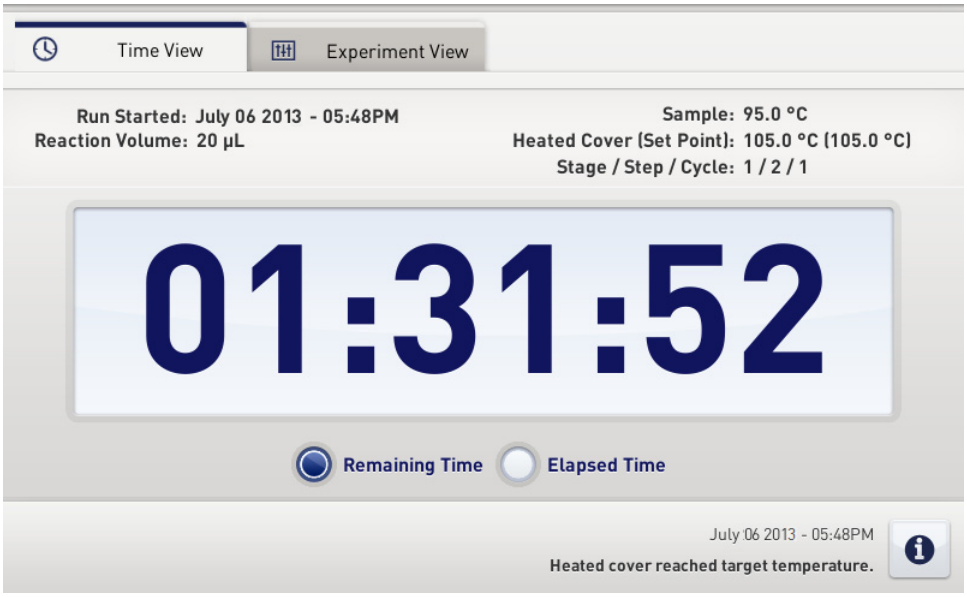
You can also view the progress of the run from the QuantStudio™ 6 or 7 Instrument touchscreen.

The following are images of the Run Method screen on the QuantStudio™ 6 or 7 Instrument touchscreen:

Experiment View



Time View


Time View
Experiment View
Run Started: July 06 2013 - 05:48PM
Reaction Volume: 20 µL
Sample: 95.0 °C
Heated Cover (Set Point): 105.0 °C (105.0 °C)
Stage / Step / Cycle: 1 / 2 / 1
01:31:52
Remaining Time
Elapsed Time
July 06 2013 - 05:48PM
Heated cover reached target temperature.

5 Review Results and Adjust Experiment Parameters

In Section 5.1 of this chapter you review the analyzed data using several of the analysis screens and publish the data. Section 5.2 of this chapter walks you through the process of modifying experiment parameters to troubleshoot problems with experiment results before rerunning an experiment.

This chapter covers:

Section 5.1 Review Results

Analyze the example experiment

1. Open the Presence/Absence example experiment file that you ran in Chapter 4.
2. Click **Analyze**. The software analyzes the data using the default analysis settings.

   Note: You can also access the experiment to analyze from the Home screen.

View the Presence/Absence Plot

The Presence/Absence Plot displays the intensity of the fluorescence for each well position. There are four Presence/Absence plot views available:

- All Calls
- Presence calls only
- Absence calls only
- Unconfirmed calls

For each view you can choose to:

- Show IPC
- Show Controls

Purpose

The purpose of viewing the Presence/Absence Plot for the example experiment is to confirm that:

- The target is absent in samples NTC and Sample (-).
- The target is present in Sample (+).
- There are no unconfirmed wells.
- The IPC succeeded in all wells.
- There is no amplification in NAC wells.

To view and assess the Presence/ Absence Plot

From the Experiment menu pane, select **Analysis ▸ Presence/Absence Plot.**

Note: If no data are displayed, click **Analyze**.

1. Display all 384 wells in the Presence/Absence Plot screen by clicking the upper left corner of the plate layout in the **Plate Layout** tab.
2. Enter the Plot Settings:

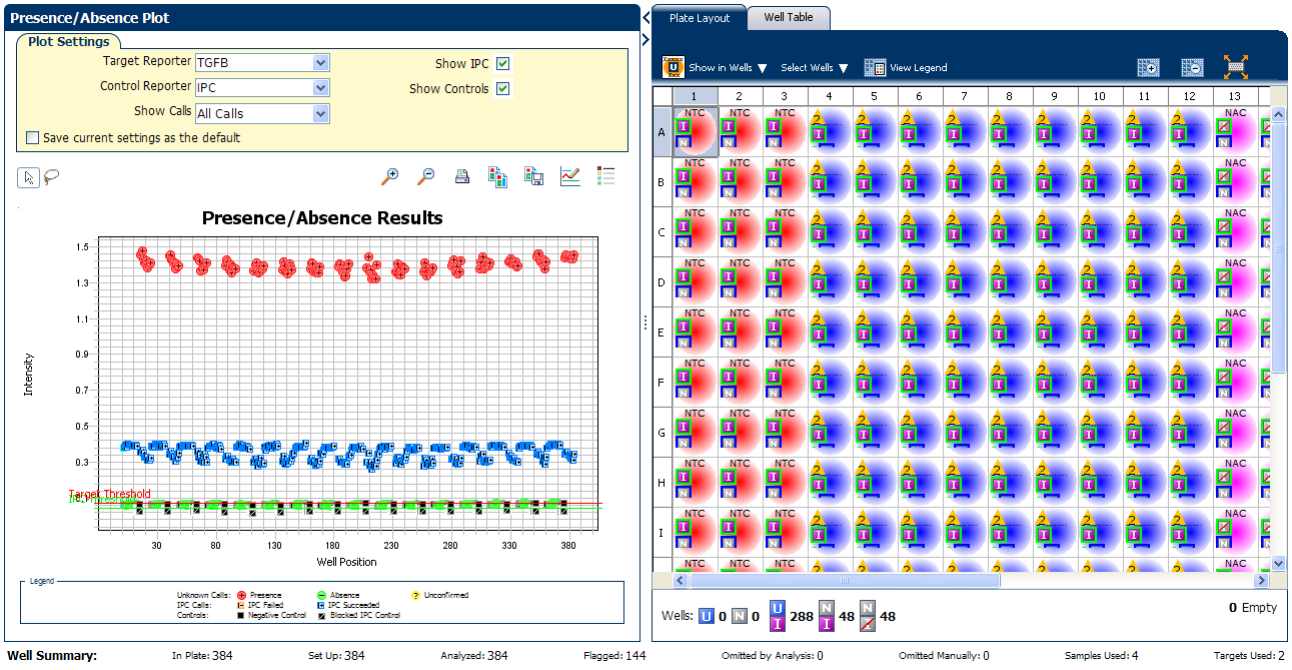
Menu	Selection
Target Reporter	TGFB
Control Reporter	IPC
Show Calls	All Calls

Menu	Selection
 (This is a toggle button. When the legend is displayed, the button changes to Hide the plot legend.)	Check (default)

3. Click the **Show IPC** check box to view the fluorescence intensity of the IPC target in the Unknown-IPC wells.
4. Click the **Show Controls** check box to view the fluorescence intensity of the IPC target in the Negative Control-IPC wells and the Negative Control-Blocked IPC wells.
5. To view the fluorescence intensity of:
   - Presence calls– select **Presence** from the Show Calls drop-down menu.
   - Absence calls– select **Absence** from the Show Calls drop-down menu.
   - Unconfirmed calls– select **Unconfirmed** from the Show Calls drop-down menu.

     Note: The Presence/Absence example experiment does not contain any unconfirmed calls.

The following is an image of the Presence/Absence Plot for the example experiment:



Tips for viewing presence/absence plots in your own experiments

- The **IPC threshold** is calculated from the Negative Control- Blocked IPC reactions.
- **The Target Threshold** is calculated from the Negative Control- IPC reactions. If the target's intensity is:
  - Above the target threshold, the call is present (regardless of the intensity of the IPC).
  - Below the target threshold, and the IPC's intensity is above the IPC threshold, the call is absent.
  - Below the target threshold, and the IPC's intensity is below the IPC threshold, the call is unconfirmed.
- T**arget Calls**:
  - Presence
  - Absence
  - Unconfirmed
- **IPC Calls**:
  - IPC Succeeded
  - IPC Failed
- **Control Well Calls**:
  - Negative Control - IPC
  - Negative Control - Blocked IPC

Assess amplification results using the Amplification Plot

IMPORTANT! Amplification plots are not used to make presence/absence calls. Examine the plots to help with troubleshooting and quality control.

Amplification plots available for viewing

The Amplification Plot displays amplification of all samples in the selected wells. There are three amplification plot views available:

- **ΔRn vs Cycle** – ΔRn is the difference in normalized fluorescence signal generated by the reporter between the pre-PCR read and the post-PCR read. This plot displays ΔRn as a function of cycle number. You can use this plot to identify and examine irregular amplification and to view threshold and baseline values for the run.
- **Rn vs Cycle** – Rn is the fluorescence signal from the reporter dye normalized to the fluorescence signal from the passive reference. This plot displays Rn as a function of cycle number. You can use this plot to identify and examine irregular amplification.
- **C_T vs Well** – C_T is the PCR cycle number at which the fluorescence meets the threshold in the amplification plot. This plot displays C_T as a function of well position. You can use this plot to locate outlying amplification (outliers).

Each plot can be viewed as a linear or log10 graph type.

Purpose

The purpose of viewing the amplification plot for the example experiment is to review the target to identify:

- Correct baseline and threshold values
- Irregular amplification
- Outliers

View the Amplification Plot

1. From the Experiment menu pane, select **Analysis ▸ Amplification Plot.**

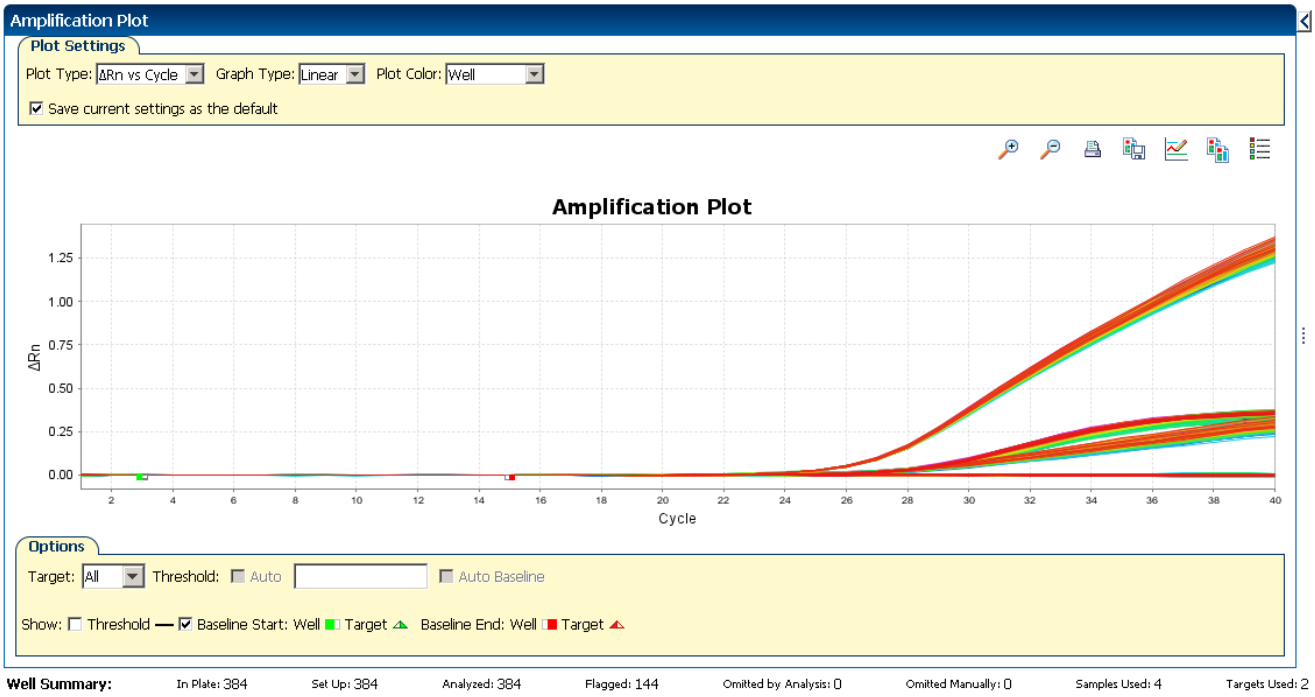
   Note: If no data are displayed, click **Analyze**.

2. Display all 384 wells in the amplification plot by clicking the upper left corner of the plate layout in the Plate Layout tab.
3. Expand the Plate Layout tab by clicking the left facing arrow that is left of the tab.
4. In the Amplification Plot screen, enter:

Menu	Selection
Plot Type	ΔRn vs Cycle
Plot Color	Well
 (This is a toggle button. When the legend is displayed, the button changes to Hide the plot legend.)	Check (default)

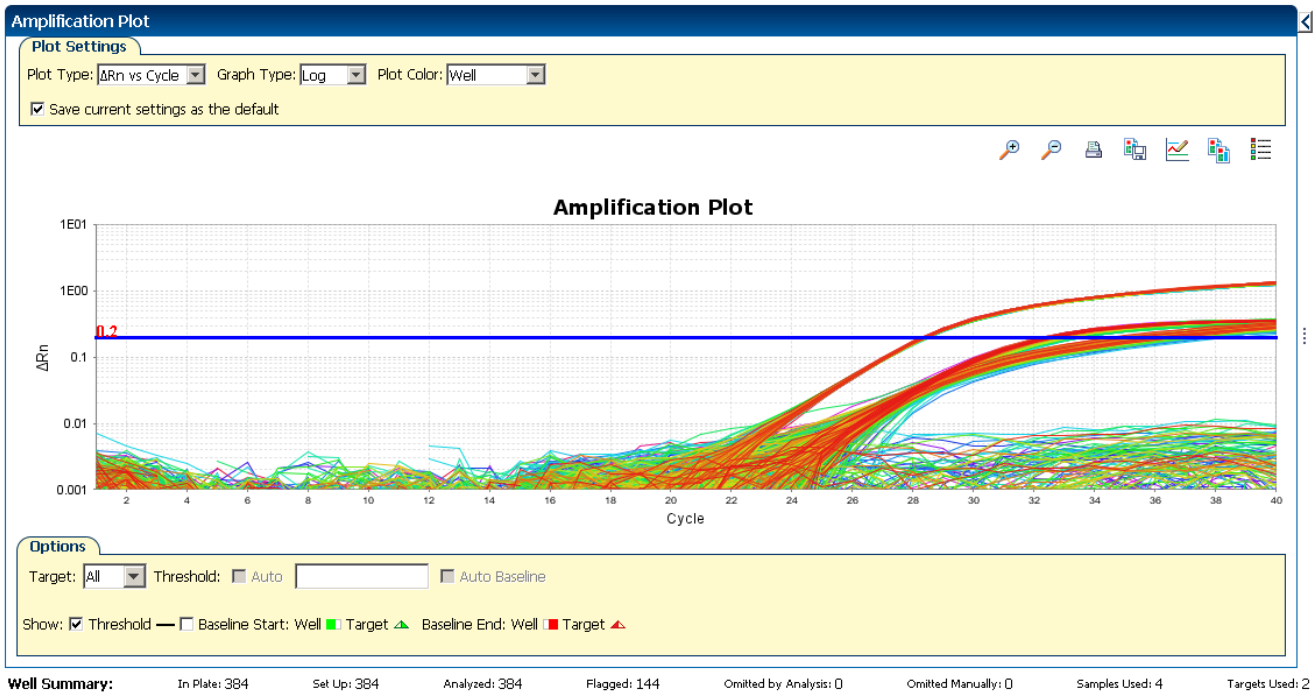
5. View the baseline values:
   a. From the Graph Type drop-down menu, select **Linear**.
   b. Select the **Baseline** check box to show the start cycle and end cycle.
   c. Verify that the baseline is set correctly: The end cycle should be set a few cycles before the cycle number where significant fluorescent signal is detected. In the example experiment, the baseline is set correctly.

Your screen should look like this:



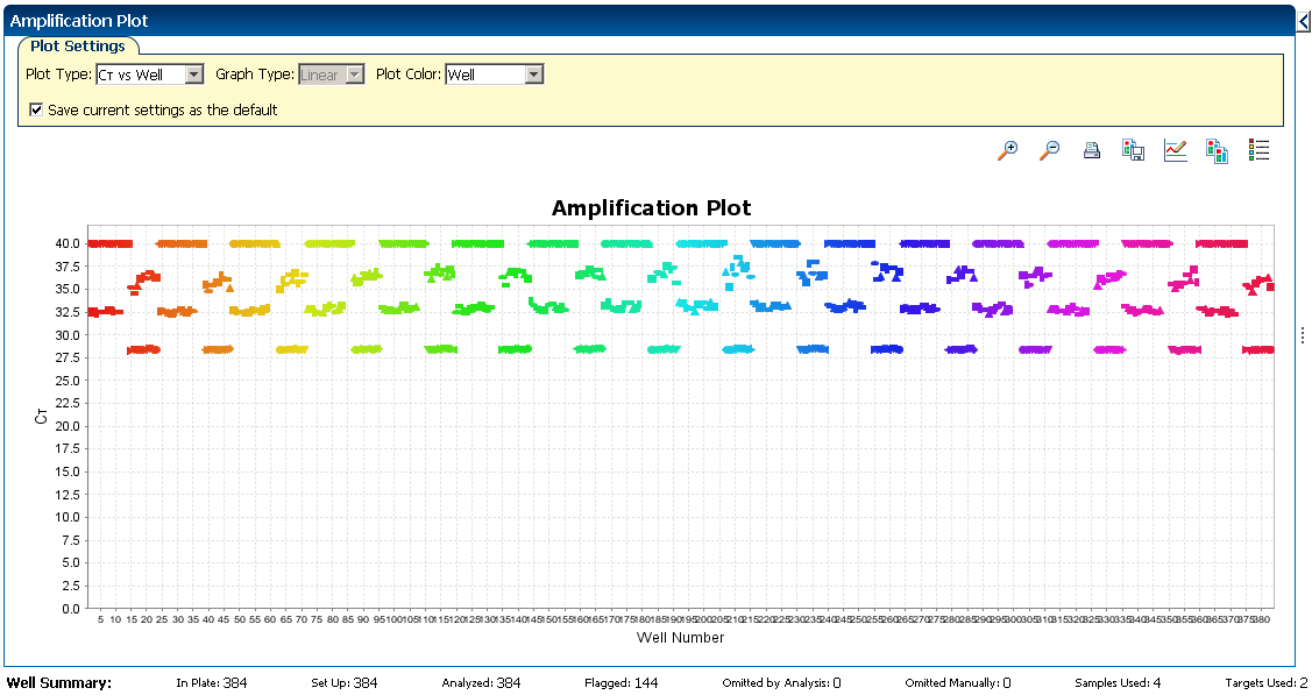
6. View the threshold values:
   a. From the Graph Type drop-down menu, select **Log**.
   b. Select the **Threshold** check box to show the threshold.
   c. Verify that the threshold is set correctly.

Your screen should look like this:



7. Locate any outliers:
   a. From the Plot Type drop-down menu, select **C_T vs Well**.
   b. Look for outliers from the amplification plot. In the example experiment, there are no outliers for IPC.

Your screen should look like this:

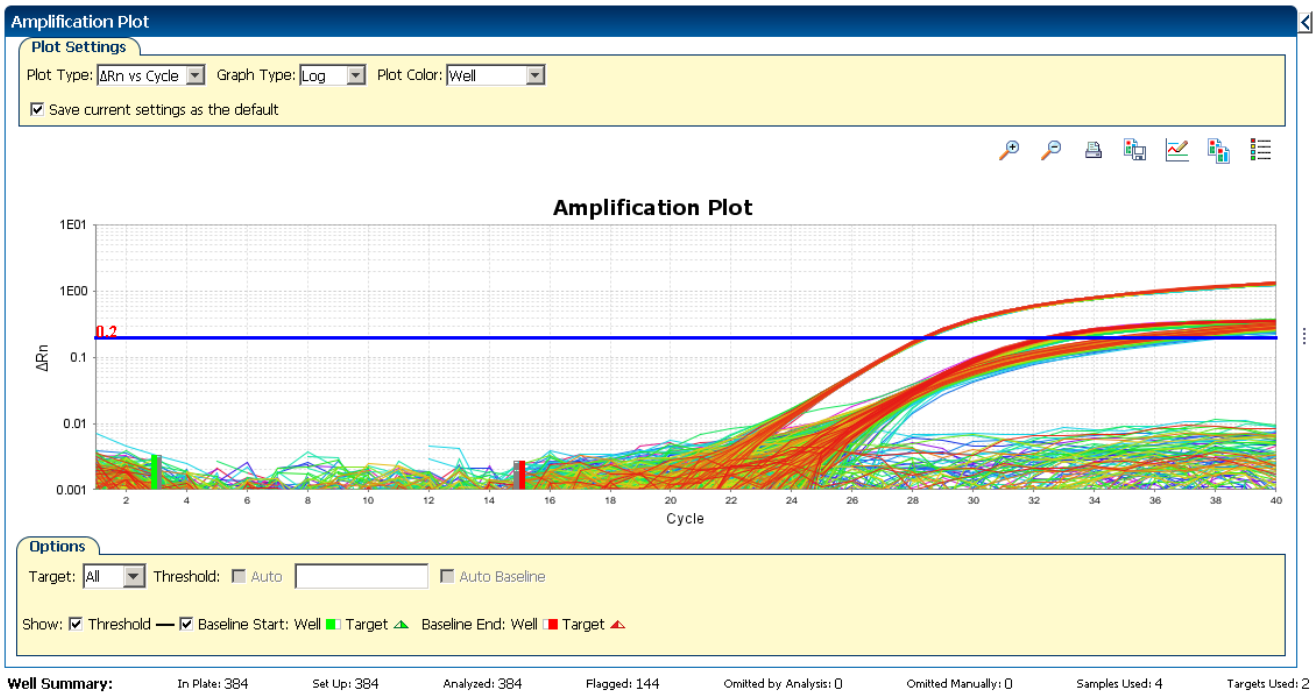


Tips for viewing amplification plots in your own experiments

When you analyze your own Presence/ Absence experiment, look for:

- **Outliers**
- **A typical amplification plot** – The QuantStudio™ 6 and 7 Flex Software automatically calculates baseline and threshold values based on the assumption that the data exhibit a *typical* amplification plot. A typical amplification plot has four distinct sections:
  - Plateau phase
  - Linear phase
  - Exponential (geometric phase)
  - Baseline

A following is an image of a typical amplification plot:



IMPORTANT! Experimental error (such as contamination or pipetting errors) can produce atypical amplification curves that can result in incorrect baseline and threshold value calculations by the QuantStudio™ 6 and 7 Flex Software. Life Technologies recommends that you examine the Amplification Plot screen and review the assigned baseline and threshold values for each well after analysis.

View the Well Table

The well table displays results data for each well in the reaction plate, including:

- The well number, sample name, target name, task, and dyes
- The calculated values: ΔRn, ΔRn mean, and ΔRn SD

  Note: ΔRn, ΔRn mean, and ΔRn SD are calculated only when the analysis call settings specify to analyze data from the pre-PCR read and the post-PCR read.
- Target and IPC thresholds, Call, Comments
- Flags

Purpose

In the Presence/Absence example experiment, you review the well table for:

- Call
- ΔRn
- Flag

To view the Well Table

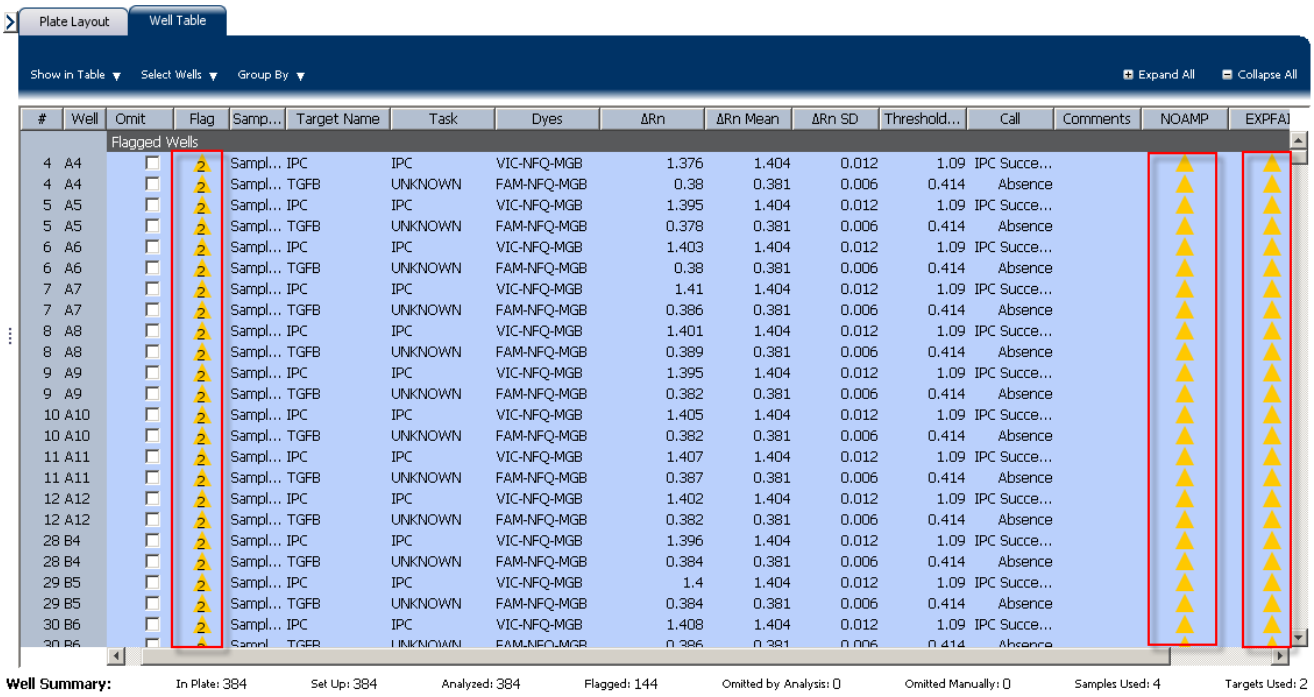
1. From the Experiment Menu pane, select **Analysis** ▸ **Amplification Plot**, then select the **Well Table** tab.

   Note: If no data are displayed, click **Analyze**.

2. Use the Group By drop-down menu to group wells by a specific category. For the example experiment, group the wells by flag, call, and ΔRn value.

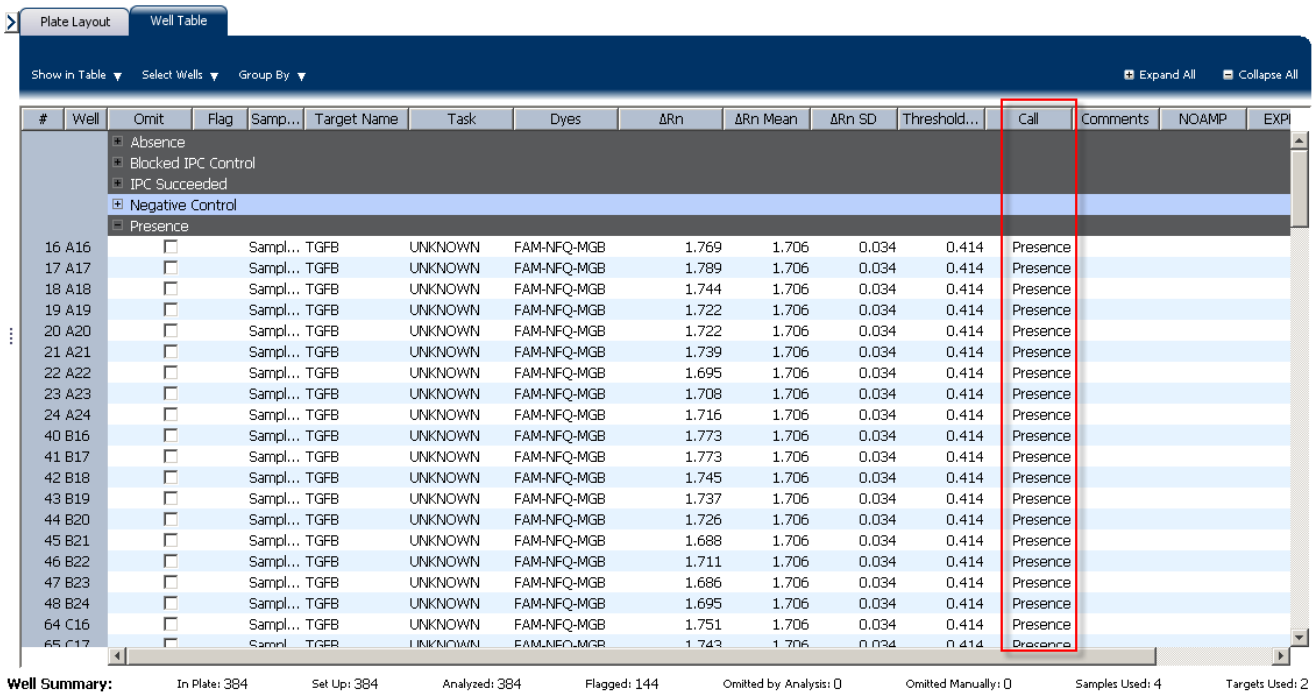
   Note: You can select only one category at a time.

   a. From the Group By drop-down menu, select **Flag**:
      - 144 wells are listed under Flagged.
      - 240 wells are listed under Unflagged.

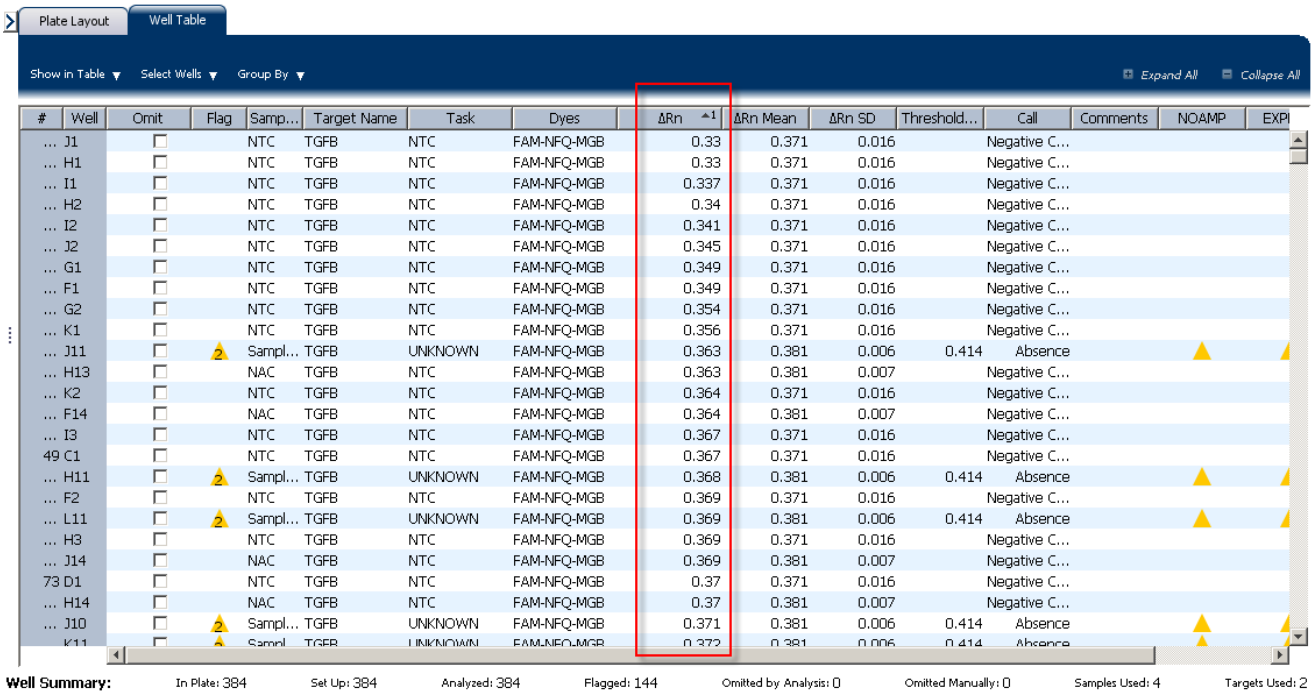


   b. From the Group By drop-down menu, select **Call**. Wells are listed in the order:
      - Absence
      - Blocked IPC Control
      - IPC Succeeded

- Negative Control
- Presence



c. From the Group By drop-down menu, select **None**. In the table, click the column heading **ΔRn**. Wells are listed in order of increasing ΔRn. Click the column heading again to reverse the sort order.



Tips for analyzing your own experiments

When you analyze your own Presence/Absence experiment, group the wells by:

- **Flag** –The software groups the flagged and unflagged wells. A flag indicates that the software has found an error in the flagged well. For a description of the QuantStudio™ 6 and 7 Flex Software flags, see "Review the flags in the QC Summary" on page 40.
- **Call** – The software groups the wells by call: Negative Control, Blocked-IPC, Presence, Absence, Unconfirmed, IPC Succeeded, and IPC Failed.

Confirm accurate dye signal using the Multicomponent Plot

The Multicomponent Plot screen displays the complete spectral contribution of each dye in a selected well over the duration of the PCR run.

Purpose

In the Presence/Absence example experiment, you review the Multicomponent Plot screen for:

- ROX™ dye (passive reference)
- FAM™ dye (reporter)
- VIC® dye (reporter)
- Spikes, dips, and/or sudden changes
- Amplification in the negative control wells

View the Multicomponent Plot

1. From the Experiment Menu pane, select **Analysis** ▸ **Multicomponent Plot**.

   Note: If no data are displayed, click **Analyze**.

2. Display the wells **one at a time** in the Multicomponent Plot screen:
   a. Click the **Plate Layout** tab.
   b. Select one well in the plate layout; the well is shown in the Multicomponent Plot screen.

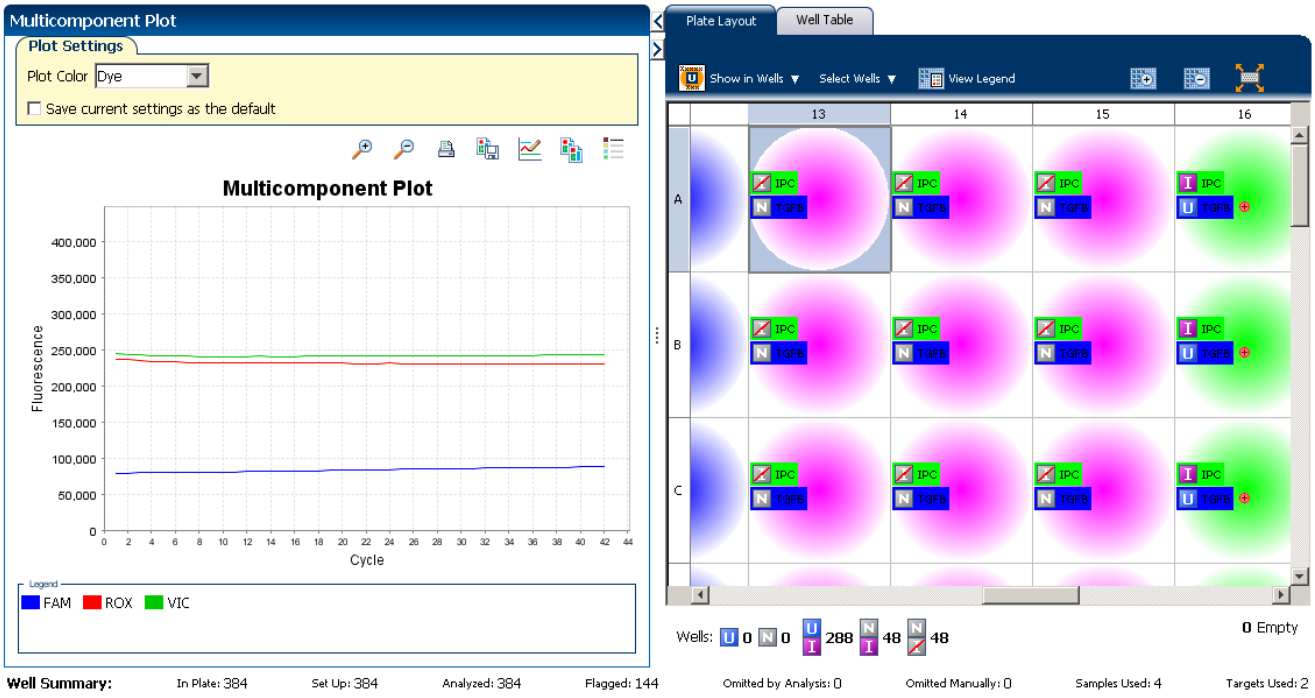
      Note: If you select multiple wells, the Multicomponent Plot screen displays the data for all selected wells simultaneously.

3. From the Plot Color drop-down menu, select **Dye**.
4. Click  **Show a legend for the plot** (default).

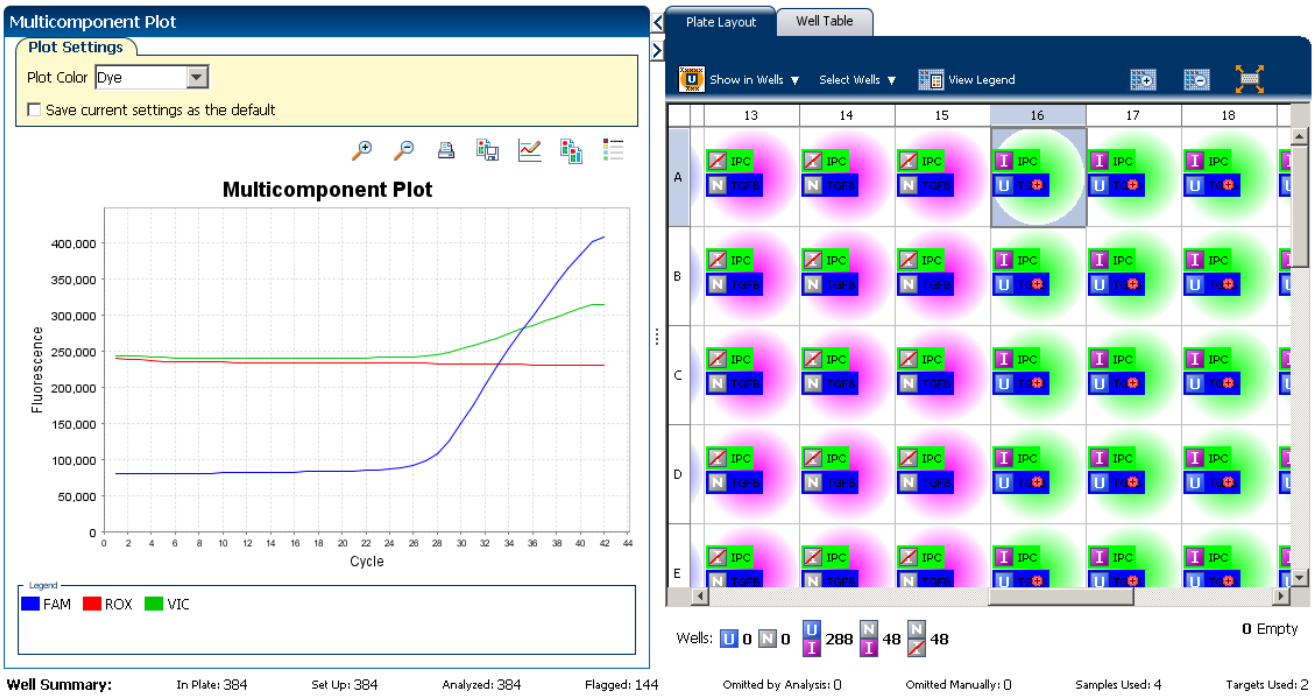
   Note: This is a toggle button. When the legend is displayed, the button changes to Hide the plot legend.

5. Check the ROX dye signal. In the example experiment, the ROX dye signal remains constant throughout the PCR process; a constant ROX dye signal indicates typical data.

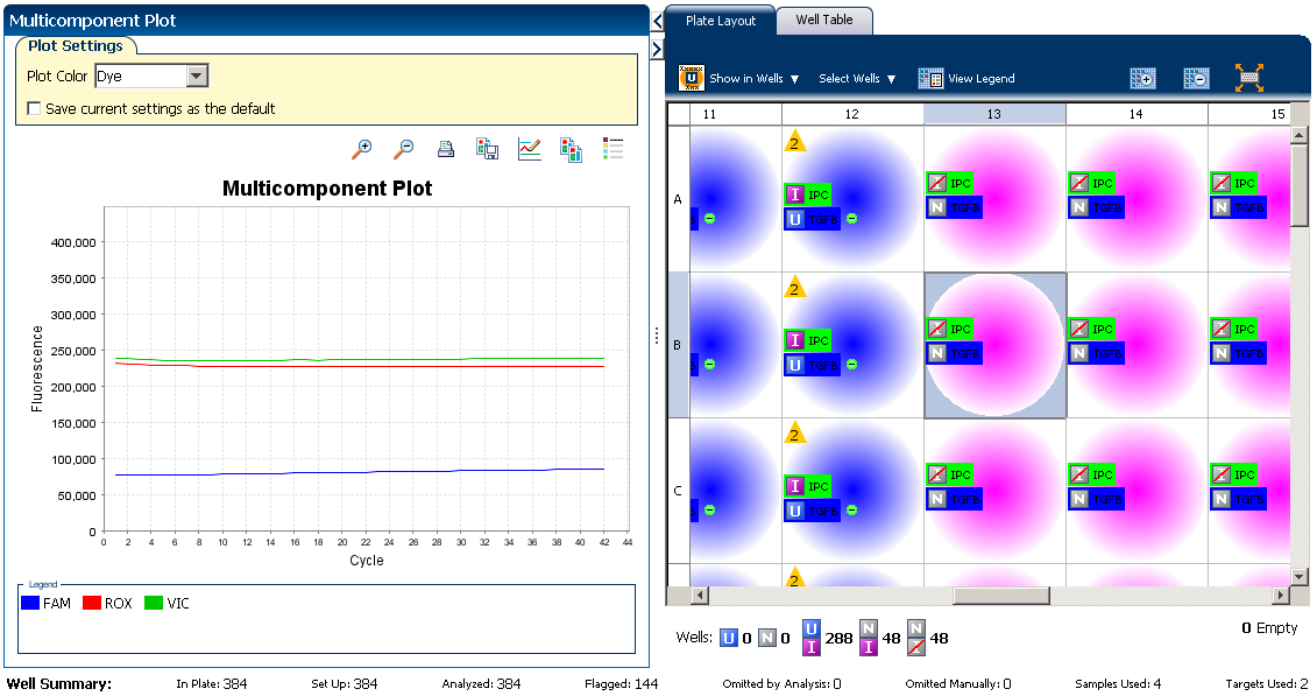
6. Check the VIC dye signal. In the example experiment the VIC dye signal should not amplify for NC-Blocked IPC wells or if the IPC call for the Unknown-IPC well is IPC Failed.



7. Check the FAM dye signal. In the example experiment, for the Sample (+), the FAM dye signal increases throughout the PCR process; increase in FAM dye signal indicates normal amplification.



8. Select the negative control (NAC) wells one at time and check for amplification. In the example experiment, there is no amplification in the negative control wells.



Tips for confirming dye accuracy in your own experiment

When you analyze your own Presence/Absence experiment, look for:

- **Passive reference (ROX)**– The passive reference dye fluorescence level should remain relatively constant throughout the PCR process.
- **Reporter dye (FAM)** – The reporter dye fluorescence level should display a flat region corresponding to the baseline. If target is present in the sample (a Presence call is made), the baseline will be followed by a rapid rise in fluorescence as the amplification proceeds.
- **Irregularities in the signal** – There should not be any spikes, dips, and/or sudden changes in the fluorescent signal.
- **Negative control wells** – There should not be any amplification in the negative control wells.

Determine signal accuracy using the Raw Data Plot

The Raw Data Plot screen displays the raw fluorescence signal (not normalized) for each optical filter for the selected wells during each cycle of the real-time PCR.

Purpose

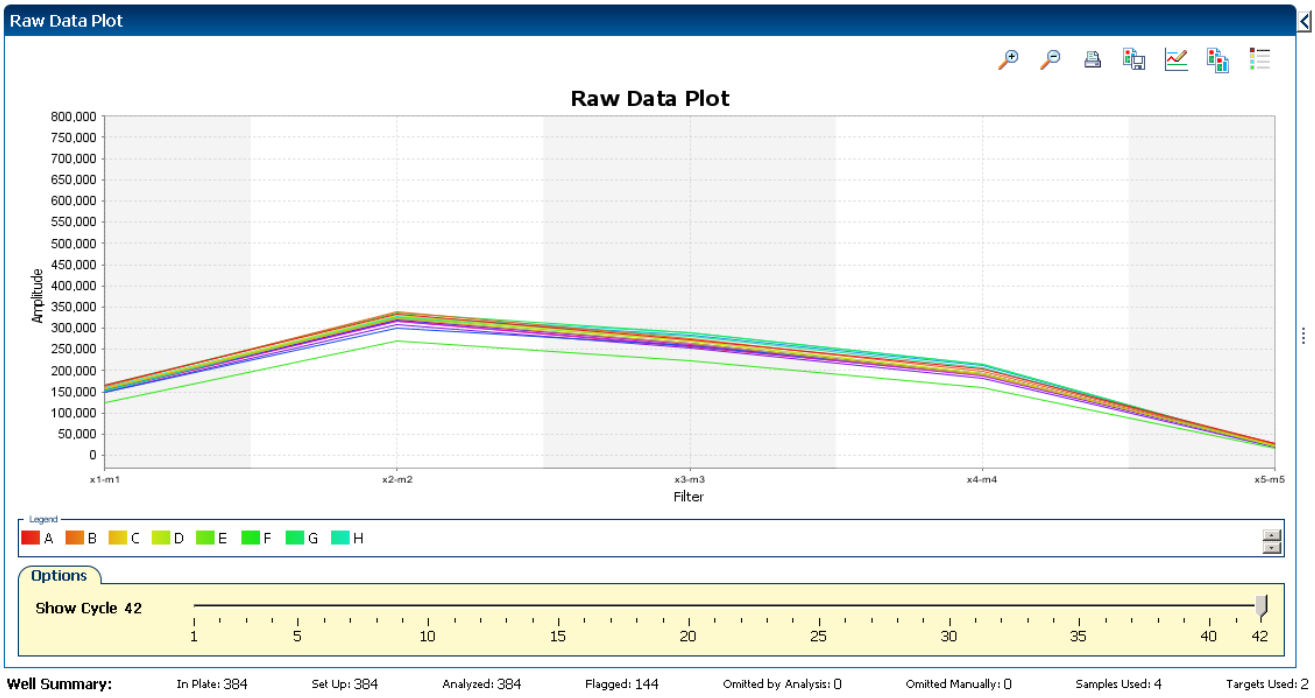
In the Presence/Absence example experiment, review the Raw Data Plot screen for a stable increase in signal (no abrupt changes or dips) from the appropriate filter.

View the Raw Data Plot

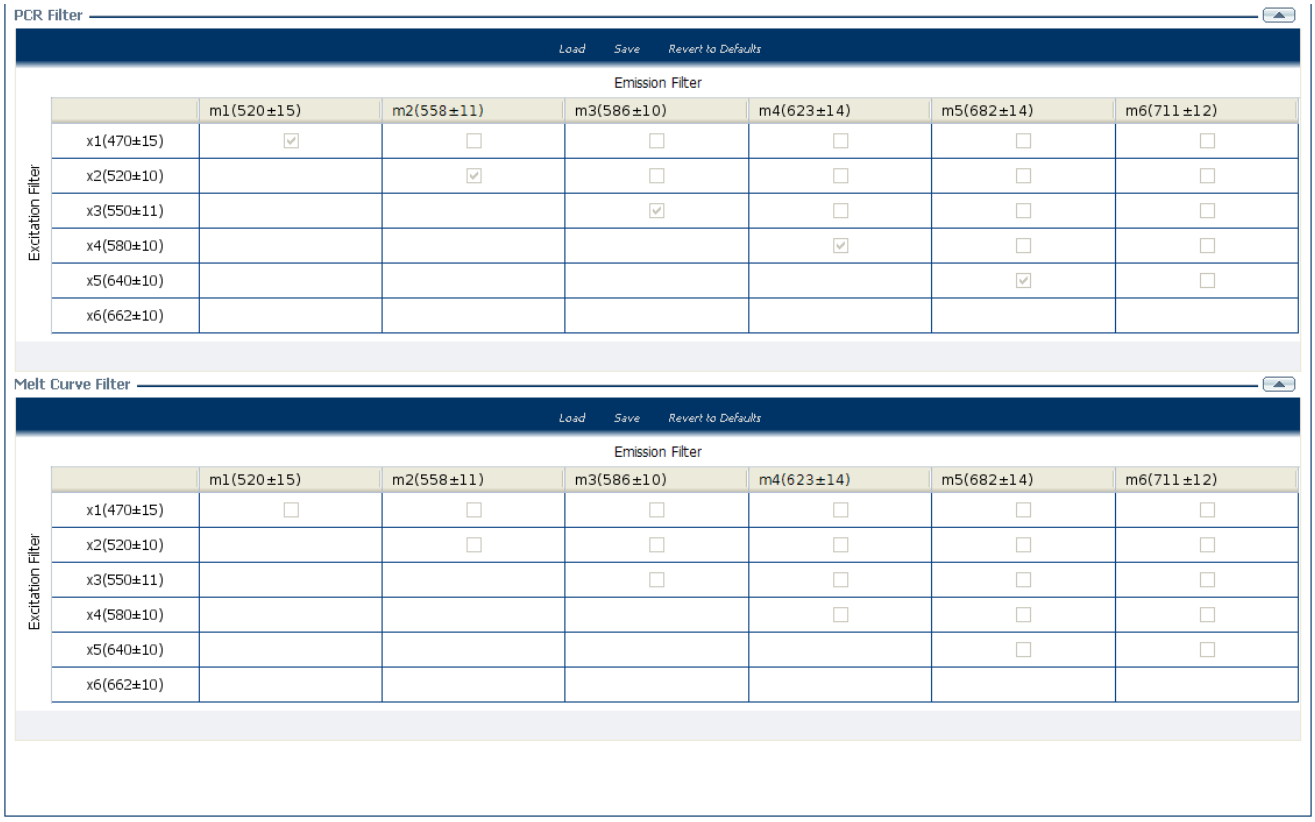
1. From the Experiment Menu pane, select **Analysis** ▸ **Raw Data Plot**.

   Note: If no data are displayed, click **Analyze**.

2. Click **Show a legend for the plot** (default). The legend displays the color code for each row of the reaction plate (see the legend in the Raw Data Plot shown below).
3. Display all 384 wells in the Raw Data Plot screen by clicking the upper left corner of the plate layout in the Plate Layout tab.
4. Select wells corresponding to a replicate group:
   - Sample (-) wells: From the Select Wells with drop-down menus, select Sample (-).
   - Sample (+) wells: From the Select Wells with drop-down menus, select Sample (+).
   - Negative control-IPC wells: Select wells A1 - P1, A2 - P2, and A3 - P3.
   - Negative control-blocked IPC wells: Select wells A13 - P13, A14 -P14, and A15 - P15.
5. Click and drag the Show Cycle pointer from cycle 1 to cycle 42. In the example experiment, there is a stable increase in signal from filter 1, which corresponds to the FAM™ dye filter.



The filters used for the example experiment are:



Tips for determining signal accuracy in your own experiments

When you analyze your own Presence/ Absence experiment, look for the following in each filter:

- Characteristic signal growth
- No abrupt changes or dips

Review the flags in the QC Summary

The QC Summary screen displays a list of the QuantStudio™ 6 and 7 Flex Software flags, including the flag frequency and location for the open experiment. In the example experiment, 144 flags have been triggered.

Note: The flags triggered in the example experiment are seen in the Sample (-) wells. The flags, NOAMP and EXPFAIL indicate that the wells containing the Sample (-) did not amplify and that the software could not identify the exponential region of the amplification plot (as amplification did not take place). The occurrence of these flags in the example experiment is valid because it indicates the absence of the target in the sample.

View the QC Summary

1. From the Experiment Menu pane, select **Analysis** ▸ **QC Summary**.

   Note: If no data are displayed, click **Analyze**.

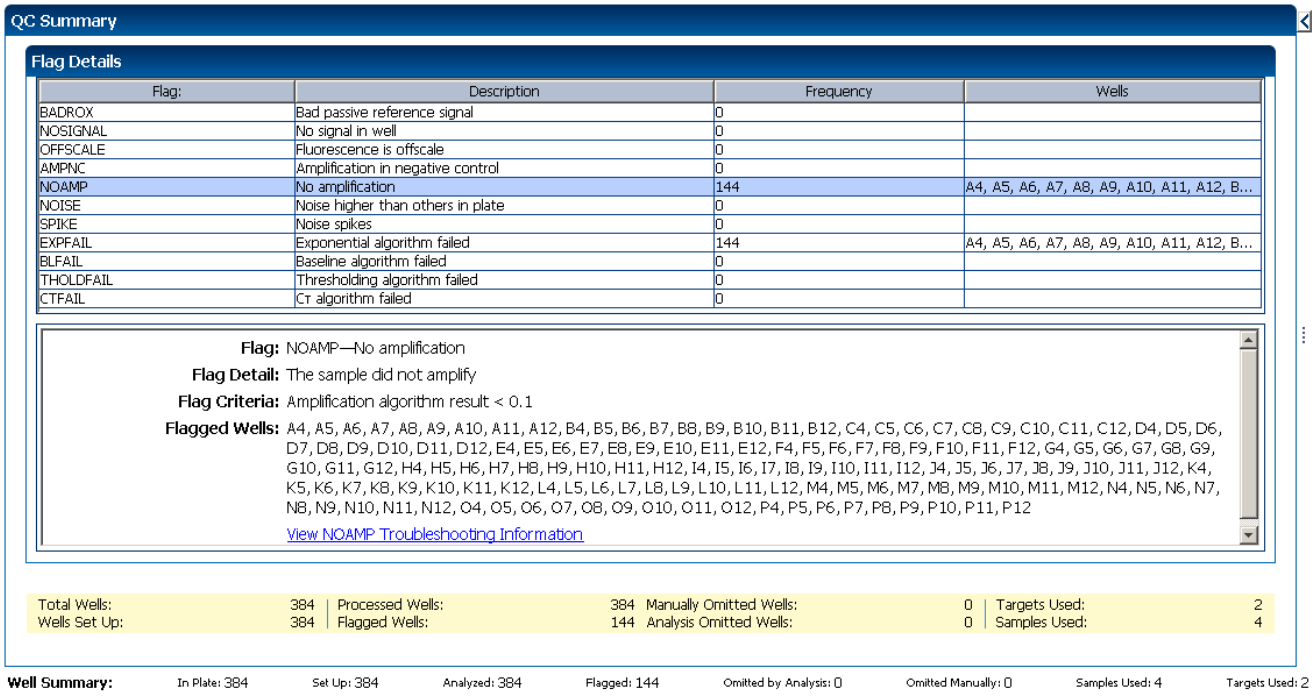
2. Review the Flags Summary

   Note: A 0 displayed in the Frequency column indicates that the flag does not appear in the experiment. If the frequency is >0, the flag appears somewhere in the experiment; the well position is listed in the Wells column.

   In the example experiment, there are 144 flagged wells.

3. In the Flag Details table, click each flag with a frequency >0 to display detailed information about the flag. In the example experiment:
   - The NOAMP flag appears 144 times, in the wells A4 - P4, A5 - P5, A6 - P6, A7 - P7, A8 - P8, A9 - P9, A10 - P10, A11 - P11, and A12 - P12.
   - The EXPFAIL flag appears 144 times, in the same wells as the NOAMP flag, that is, A4 - P4, A5 - P5, A6 - P6, A7 - P7, A8 - P8, A9 - P9, A10 - P10, A11 - P11, and A12 - P12.

4. *(Optional)* For those flags with frequency >0, click the troubleshooting link to view information on correcting the flag.



Possible flags

For Presence/Absence experiments, the flags listed below may be triggered by the experiment data.

Flag	Description
Pre-processing flag	
OFFSCALE	Fluorescence is offscale
Primary analysis flags	
BADROX	Bad passive reference signal
NOAMP	No amplification

Flag	Description
NOISE	Noise higher than others in plate
SPIKE	Noise spikes
NOSIGNAL	No signal in well
EXPFAIL	Exponential algorithm failed
BLFAIL	Baseline algorithm failed
THOLDFAIL	Thresholding algorithm failed
CTFAIL	C_T algorithm failed

Secondary analysis flags	
AMPNC	Amplification in negative control

Note: If the experiment does not include amplification, then the only flags are BADROX, NOSIGNAL, and OFFSCALE.

For more information

For more information on...	Refer to...	Publication number
Publishing data	Chapter 2 in Booklet 1, *Getting Started with QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Experimentss*	4489822

Section 5.2 Adjust parameters for re-analysis of your own experiments

Adjust analysis settings

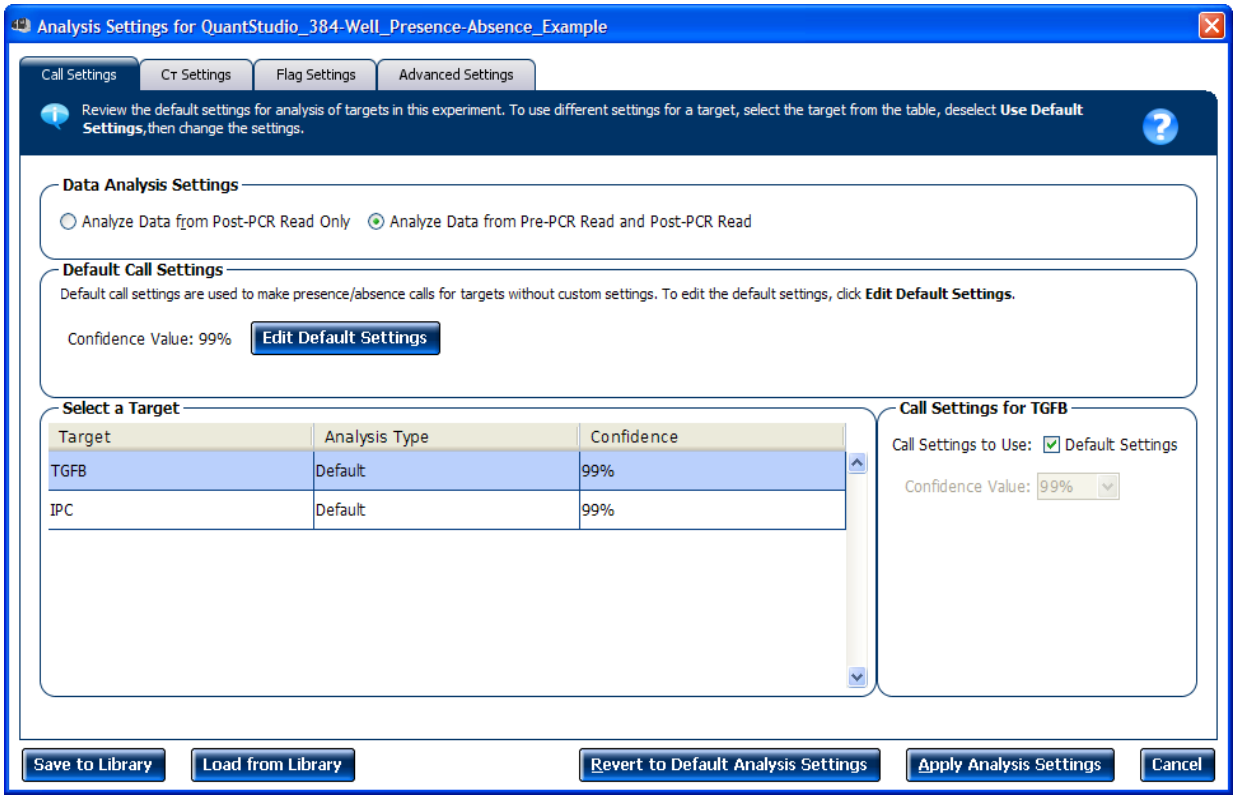
The Analysis Settings dialog box displays the analysis settings for the call, threshold cycle (C_T), flags, and advanced options.

If the default analysis settings in the QuantStudio™ 6 and 7 Flex Software are not suitable for your own experiment, you can change the settings in the Analysis Settings dialog box, then reanalyze your experiment.

View the analysis settings

1. From the Experiment Menu pane, select **Analysis**.
2. Click **Analysis** ▸ **Analysis Settings** to open the Analysis Settings dialog box.
   In the example experiment, the default analysis settings are used for each tab:
   - Call Settings
   - C_T Settings
   - Flag Settings
   - Advanced Settings

   The following is an image of the Analysis Settings dialog box for a Presence/Absence experiment:



3. View and, if necessary, change the analysis settings (see "Adjust analysis settings" below).

   Note: You can save the changes to the analysis settings to the Analysis Settings Library for later use. For more information, see About the Analysis Settings Library in Booklet 1, *Getting Started with QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Experiments*.

4. Click **Apply Analysis Settings** to apply the current analysis settings.

   Note: You can go back to the default analysis settings, by clicking **Revert to Default Analysis Settings**.

Adjust analysis settings

Call Settings

Use the Call Settings tab to:

- Change the default data analysis settings. You can select from:
  - Analyze data from Post-PCR Read only
  - Analyze data from Pre-PCR Read and Post-PCR Read
- Edit the default call settings.
  - Click **Edit Default Settings**, then select the confidence value to use to make presence/absence calls. If the confidence value is less than the call setting, the call is unconfirmed.
  - Click **Save Changes**.
- Use custom call settings for a target.
  - Select one or more targets in the table, then deselect the **Default Settings** check box.
  - Select the confidence value to use to make presence/absence calls for the selected target(s).

C_T Settings

- **Data Step Selection**

  Use this feature to select one stage/step combination for C_T analysis when there is more than one data collection point in the run method.
- **Algorithm Settings**

  Use the Baseline Threshold algorithm to determine the C_T values.

  The Baseline Threshold algorithm is an expression estimation algorithm that subtracts a baseline component and sets a fluorescent threshold in the exponential region for gene quantification.
- **Default C_T Settings**

  Use the default C_T settings feature to calculate C_T for the targets that do not have custom settings. To edit the default settings, click **Edit Default Settings**.

- **C_T Settings for Target**

  When you manually set the threshold and baseline, Life Technologies recommends:

Setting	Recommendation
Threshold	Enter a value for the threshold so that the threshold is: • Above the background. • Below the plateau and linear regions of the amplification curve. • Within the exponential phase of the amplification curve.
Baseline	Select the Start Cycle and End Cycle values so that the baseline ends before significant fluorescent signal is detected.

  Note: This setting is applicable only to the Baseline Threshold algorithm.

  Note: Selecting Automatic Threshold implies selection of automatic setting of the baseline. However, if Automatic Threshold is deselected, then you can choose between setting the baseline either automatically or manually.

Flag Settings

Use the Flag Settings tab to:

- Adjust the sensitivity so that more wells or fewer wells are flagged.
- Change the flags that are applied by the QuantStudio™ 6 and 7 Flex Software.

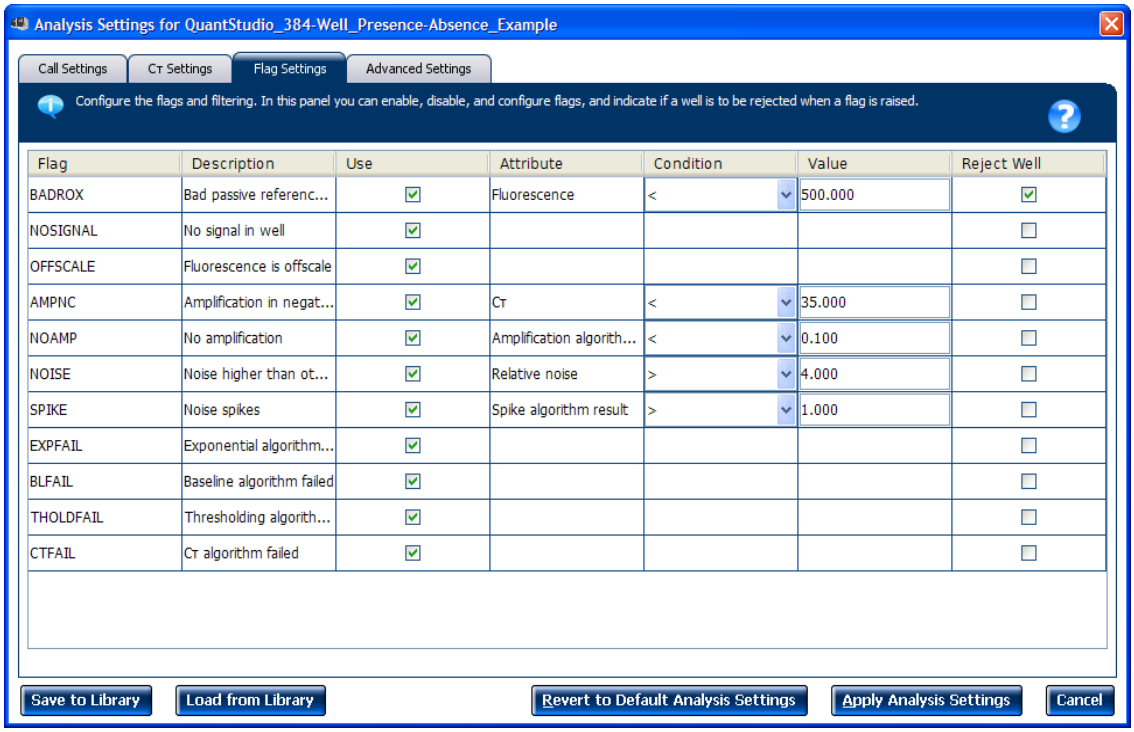
To adjust the flag settings:

1. In the Use column, select the check boxes for flags to apply during analysis.
2. *(Optional)* If an attribute, condition, and value are listed for a flag, specify the setting for applying the flag.

   Note: If you choose to adjust the setting for applying a flag, make minor adjustments as you evaluate the appropriate setting.
3. In the Reject Well column, select the check boxes if you want the software to reject wells with the flag.

   Note: After you have rejected the flagged wells, analysis results depend on factors such as the experiment type and flag type. For example, rejecting wells flagged by HIGHSD in experiments using the Standard Deviation calculations may change the result of C_T SD. For some flags, analysis results calculated before the well is rejected are maintained.
4. Click **Apply Analysis Settings** in the Analysis Settings dialog box. If the run status is complete, the data are reanalyzed.

The following is an image of the Flag Settings tab:



Advanced Settings

Use the Advanced Settings tab to change baseline settings well-by-well.

To use custom baseline settings for a well-target combination:

1. Select one or more well-target combinations in the table.
2. Deselect the **Use C_T Settings Defined for Target** check box.
3. Define the custom baseline settings:
   - For automatic baseline calculations, select the **Automatic Baseline** check box.
   - To define the baseline, deselect the **Automatic Baseline** check box, then enter the baseline start cycle and baseline end cycle.

For more information

For more information on...	Refer to	Publication number
Amplification efficiency	*Amplification Efficiency of TaqMan® Gene Expression Assays Application Note.*	127AP05-03

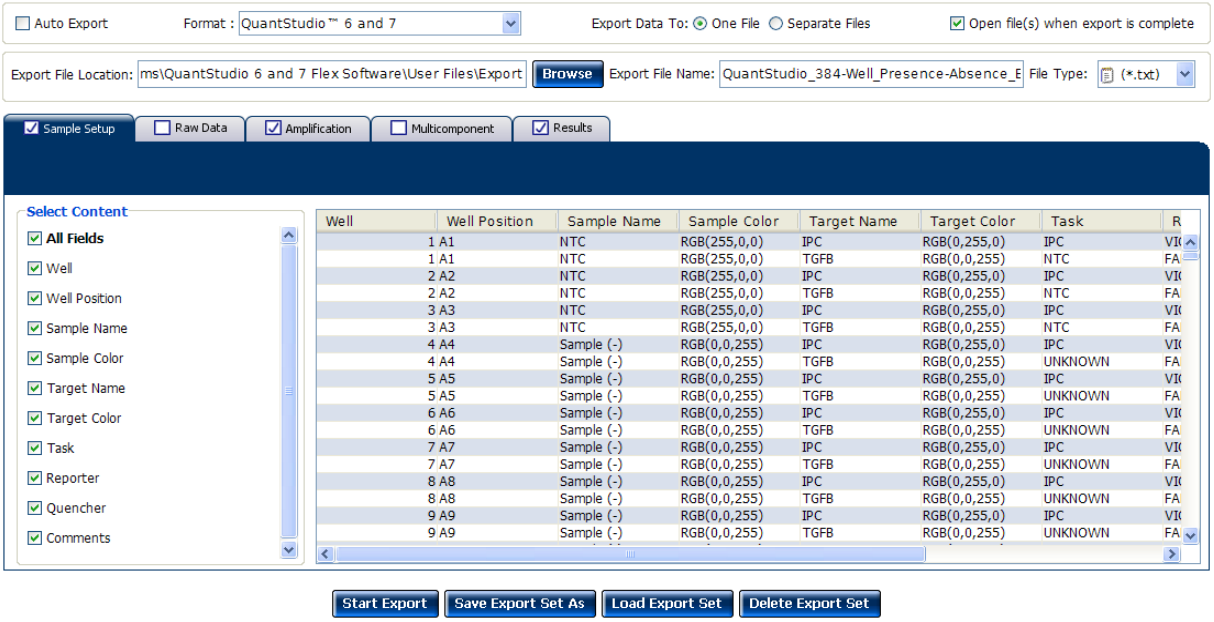
6 Exporting Analysis Results

1. Open the Presence/Absence example experiment file that you analyzed in Chapter 5.
2. In the Experiment Menu, click  **Export.**

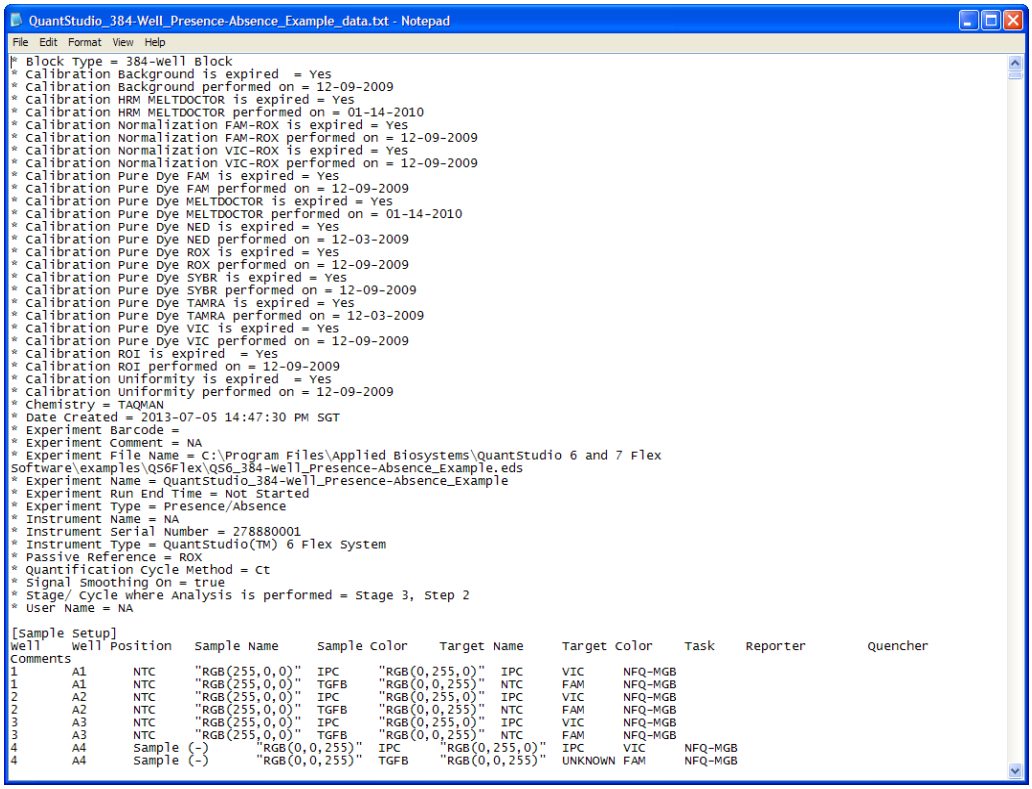
   Note: To export data automatically after analysis, select the **Auto Export** check box during experiment setup or before running the experiment. Auto export is unchecked for the example experiment.
3. Select **QuantStudio™ 6 and 7** format.
4. Complete the Export dialog box as shown below:

Field or Selection	Entry
Select Data to export/ Select Content	Results
Export Data To	One File
Export File Name	QuantStudio_384-Well_Presence-Absence_Example_data
File Type	*.txt
Export File Location	C:\Applied Biosystems\QuantStudio 6 and 7 Flex Software\User Files\Export

Your Export screen should look like this:



Your exported file when opened in Notepad should look like this:



QuantStudio_384-Well_Presence-Absence_Example_data.txt - Notepad

File Edit Format View Help

```
* Block Type = 384-Well Block
* Calibration Background is expired  = Yes
* Calibration Background performed on = 12-09-2009
* Calibration HRM MELTDOCTOR is expired = Yes
* Calibration HRM MELTDOCTOR performed on = 01-14-2010
* Calibration Normalization FAM-ROX is expired = Yes
* Calibration Normalization FAM-ROX performed on = 12-09-2009
* Calibration Normalization VIC-ROX is expired = Yes
* Calibration Normalization VIC-ROX performed on = 12-09-2009
* Calibration Pure Dye FAM is expired = Yes
* Calibration Pure Dye FAM performed on = 12-09-2009
* Calibration Pure Dye MELTDOCTOR is expired = Yes
* Calibration Pure Dye MELTDOCTOR performed on = 01-14-2010
* Calibration Pure Dye NED is expired = Yes
* Calibration Pure Dye NED performed on = 12-03-2009
* Calibration Pure Dye ROX is expired = Yes
* Calibration Pure Dye ROX performed on = 12-09-2009
* Calibration Pure Dye SYBR is expired = Yes
* Calibration Pure Dye SYBR performed on = 12-09-2009
* Calibration Pure Dye TAMRA is expired = Yes
* Calibration Pure Dye TAMRA performed on = 12-03-2009
* Calibration Pure Dye VIC is expired = Yes
* Calibration Pure Dye VIC performed on = 12-09-2009
* Calibration ROI is expired  = Yes
* Calibration ROI performed on = 12-09-2009
* Calibration Uniformity is expired  = Yes
* Calibration Uniformity performed on = 12-09-2009
* Chemistry = TAQMAN
* Date Created = 2013-07-05 14:47:30 PM SGT
* Experiment Barcode =
* Experiment Comment = NA
* Experiment File Name = C:\Program Files\Applied Biosystems\QuantStudio 6 and 7 Flex
Software\examples\QS6Flex\QS6_384-Well_Presence-Absence_Example.eds
* Experiment Name = QuantStudio_384-Well_Presence-Absence_Example
* Experiment Run End Time = Not Started
* Experiment Type = Presence/Absence
* Instrument Name = NA
* Instrument Serial Number = 278880001
* Instrument Type = QuantStudio(TM) 6 Flex System
* Passive Reference = ROX
* Quantification Cycle Method = Ct
* Signal Smoothing On = true
* Stage/ Cycle where Analysis is performed = Stage 3, Step 2
* User Name = NA

[Sample Setup]
Well    Well Position   Sample Name     Sample Color    Target Name     Target Color    Task    Reporter        Quencher
Comments
1       A1      NTC     "RGB(255,0,0)"  IPC     "RGB(0,255,0)"  IPC     VIC     NFQ-MGB
1       A1      NTC     "RGB(255,0,0)"  TGFB    "RGB(0,0,255)"  NTC     FAM     NFQ-MGB
2       A2      NTC     "RGB(255,0,0)"  IPC     "RGB(0,255,0)"  IPC     VIC     NFQ-MGB
2       A2      NTC     "RGB(255,0,0)"  TGFB    "RGB(0,0,255)"  NTC     FAM     NFQ-MGB
3       A3      NTC     "RGB(255,0,0)"  IPC     "RGB(0,255,0)"  IPC     VIC     NFQ-MGB
3       A3      NTC     "RGB(255,0,0)"  TGFB    "RGB(0,0,255)"  NTC     FAM     NFQ-MGB
4       A4      Sample (-)      "RGB(0,0,255)"  IPC     "RGB(0,255,0)"  IPC     VIC     NFQ-MGB
4       A4      Sample (-)      "RGB(0,0,255)"  TGFB    "RGB(0,0,255)"  UNKNOWN FAM     NFQ-MGB
```

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lifetechnologies.com

October 2013

USER GUIDE

applied biosystems®
by life technologies™

QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Getting Started Guide for Melt Curve Experiments

Booklet 6

Publication Number 4489822
Revision A

life technologies™

Contents

1 About Melt Curve Experiments

This chapter covers:

IMPORTANT! First-time users of the QuantStudio™ 6 and 7 Flex Real-Time PCR System Software, please read Booklet 1, *Getting Started with QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Experiments* and Booklet 7, *QuantStudio™ 6 and 7 Flex Real-Time PCR System Software - Appendixes* of this binder thoroughly. The booklet provides information and general instructions that are applicable to all the experiments described in this binder.

Note: For more information about any of the topics discussed in this guide, access the Help from within QuantStudio™ 6 and 7 Flex Real-Time PCR System Software by pressing **F1**, clicking in the toolbar, or selecting **Help ▸ QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Help**.

Overview

A Melt Curve, also known as dissociation curve, is a plot of data collected during the Melt Curve stage of an experiment. Melt Curve experiments are performed to determine the melting temperature (Tm) of a target nucleic acid sequence or to identify nonspecific PCR amplification.

Melting temperature (Tm) is the temperature at which 50% of the target DNA is double-stranded and 50% is dissociated into single-stranded DNA.

The melting temperature and non-specific PCR amplification can be identified as peaks in the melt curve stage of an experiment.

About the Melt Curve reactions

With Melt Curve experiments, the reactions consist of completed PCR reactions that contain amplified products and SYBR® Green dye to detect double-stranded DNA.

The QuantStudio™ 6 and 7 Flex Real-Time PCR System Software detects the number of fluorescence peaks, determines the melting temperature (Tm) for each peak, and plots the results in a melt curve.

The fluorescence data collected during the QuantStudio™ 6 or 7 Instrument run are stored in an experiment data file (*.eds).

There are two types of reactions in a Melt Curve experiment:

- **Unknowns** - Wells containing PCR product with unknown melting temperature(s).
- **Negative controls** - Wells containing buffer or water instead of sample. Negative controls should contain no double-stranded DNA.

About the example experiment

To illustrate how to perform Melt Curve experiments, this guide leads you through the process of designing and performing an example experiment. The example experiment represents a typical setup that you can use to quickly familiarize yourself with a QuantStudio™ 6 and 7 Flex Real-Time PCR System Software.

The objective of the example Melt Curve experiment is to investigate the melting temperature of Target 1, and verify that no extraneous peaks appear. The SYBR Green reagent is used to detect the melting temperature stage.

Note: The example experiment performs a melt curve analysis on PCR products from a PCR performed on the QuantStudio™ 6 or 7 Instrument or on another thermal cycler.

2 Design the Experiment

This chapter explains how to design the example experiment from the Experiment Setup menu.

Note: To automatically export the analyzed data to a specified location, select the Auto Export task from the Export screen, before running the experiment. For more information on Auto Export, refer to Chapter 2 in Booklet 1, *Getting Started with QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Experiments*.

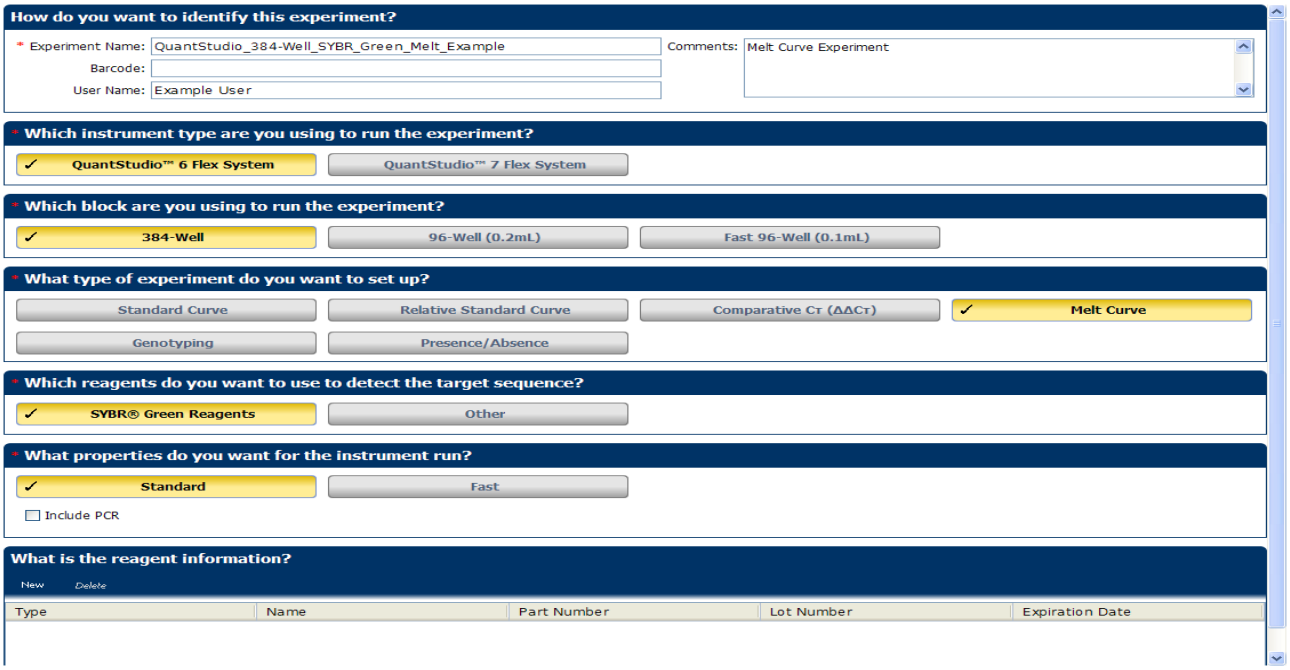
Define the experiment properties

Click **Experiment Setup** ▸ **Experiment Properties** to create a new experiment in the QuantStudio™ 6 and 7 Flex Software. Enter:

Field or Selection	Entry
Experiment Name	QuantStudio_384-Well_SYBR_Green_Melt_Example
Barcode	Leave field empty
User Name	Example User
Comments	Melt Curve example
Instrument type	QuantStudio™ 6 Flex System
Block	384-Well Block
Experiment Type	Melt Curve
Reagents	SYBR® Green Reagents
Ramp speed	Standard
Include PCR	Unchecked
Reagent information	NA

Save the experiment.

Your Experiment Properties screen should look like this:



Define targets and samples

Click **Define** to access the Define screen. Enter:

1. Targets

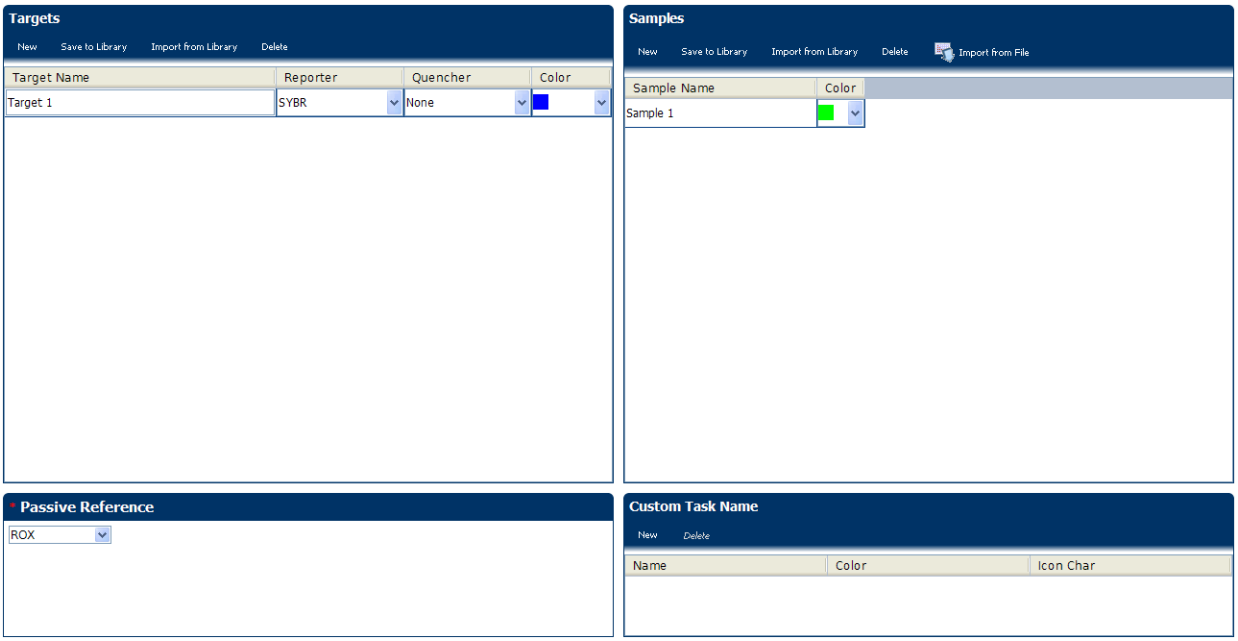
Target Name	Reporter	Quencher	Color
Target 1	SYBR	None	

2. Samples

Sample Name	Color
Sample 1	

3. Dye to be used as a Passive Reference
   ROX
4. Custom Task Name
   Not applicable

Your Define screen should look like this:

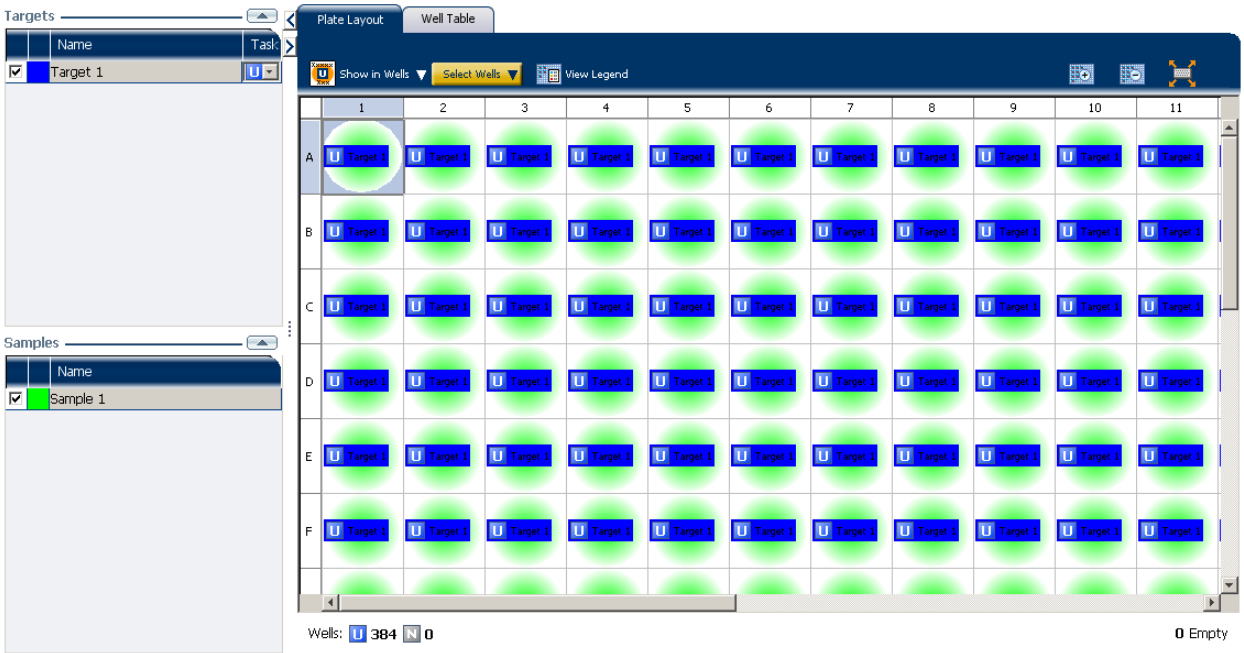


Assign targets and samples

Click **Assign** to access the Assign screen. Enter the targets and samples:

Target Name	Sample	Well Number	Task
SYBR	Sample 1	A1 - P24 (Columns 1 -24)	Unknown

Your Assign screen should look like this:



Set up the run method

Set the thermal profile

Click **Run Method** to access the Run Method screen. Set the thermal profile under the Graphical View tab. Enter:

- Reaction Volume Per Well: 20 µL
- Thermal Profile

Stage	Step	Ramp rate	Temperature	Time
Melt Curve Stage	Step 1	1.6°C/s	95ºC	15 seconds
	Step 2	1.6°C/s	60ºC	1 minute
	Step 3 (Dissociation)	0.05°C/s	95ºC	15 seconds

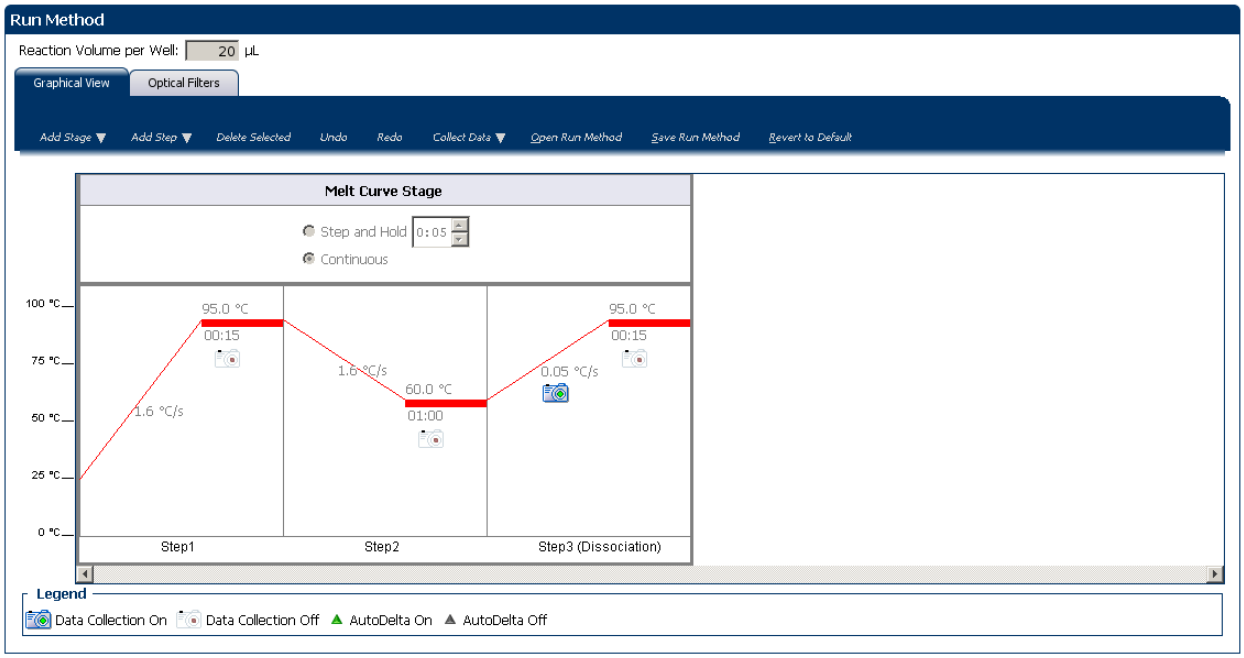
Edit the ramp increment

Edit the ramp increment for a melt curve (dissociation) step.

1. Select a melt curve ramp increment method:
   - **Step and Hold** – Increases or decreases the ramp temperature in 0.1°C increments over the time (duration) for the melt curve ramp.
   - **Continuous (default)** – Increases or decreases the ramp rate in 0.005°C per second increments.
2. If you selected the Step and Hold ramp increment method, edit the melt curve ramp time:
   - To increase or decrease the time in 1-minute or 1-second increments, click the **Step and Hold** field, select the minutes or seconds, then use the up or down arrow keys or click the up or down buttons in the field until you reach the desired time.
   - To enter the desired time, click the **Step and Hold** field, select the minutes or seconds, then enter the desired time.
3. Edit the melt curve ramp increment:
   - To increase or decrease the ramp increment, click the melt curve (dissociation) ramp increment in the thermal profile, then use the up or down arrow keys or click the up or down buttons in the field until you reach the desired value.
   - To enter the desired ramp increment, click the melt curve (dissociation) ramp increment in the thermal profile, select the value in the field, then enter the desired value.

     Note: To view the maximum and minimum allowed values, place the cursor over melt curve (dissociation) ramp increment in the thermal profile and wait for the tooltip to pop up.

Your Run Method screen should look like this:



For more information

For more information on...	Refer to	Publication number
Consumables	Chapter 1 in Booklet 1, *Getting Started with QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Experiments* Appendix A in Booklet 7, *QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Experiments - Appendixes*	4489822
Using Alternative Setup	Chapter 3 in Booklet 1, *Getting Started with QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Experiments*	4489822

3 Prepare the Reactions

This chapter explains how to prepare the reactions for running a PCR prior to running a Melt Curve.

To perform a Melt Curve experiment without running a PCR, use the reaction plate containing the PCR product.

Note: The example experiment performs a melt curve analysis on PCR products from a PCR performed on the QuantStudio™ 6 or 7 Instrument or on another thermal cycler.

This chapter covers:

Assemble required materials

- Items listed in Booklet 1, *Getting Started with QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Experiments*
- Sample 1
- Example experiment reaction mix components:
  - Power SYBR® Master Mix
  - Target - Assay Mix Forward primer (10 µM)
  - Target - Assay Mix Reverse primer (10 µM)

Prepare the sample dilutions

The stock concentration of each sample is 100 ng/µL. After you dilute the sample according to the Sample Dilutions Calculations table, the sample will have a concentration of 10 ng/µL. Add 2 µL to each reaction.

Sample name	Stock concentration (ng/µL)	Sample volume (µL)	Diluent volume (µL)	Total volume of diluted sample (µL)
Sample 1	100.0	2	18	20

Prepare the reaction mix ("cocktail mix")

The following table lists the universal assay conditions [volume and final concentration for using the Power SYBR Master Mix (2X)].

Reaction Component	Volume for 1 reaction (µL)	Volume for 384 reactions + 10% excess (µL) = 424 reactions
Power SYBR®Green PCR Master Mix (2X)	10	4240
Forward primer (10 µM)	0.1	42.4
Reverse primer (10 µM)	0.1	42.4
Water	7.8	3307.2
Total reaction mix volume	18	7632

Procedure

1. Label an appropriately sized tube for the reaction mix: Power SYBR Reaction Mix.
2. Add the required volume of each cocktail mix component to the tube.
3. Mix the cocktail thoroughly by gently pipetting up and down several times, then cap the tube.
4. Centrifuge the tube briefly to remove air bubbles.
5. Place the cocktail mix on ice until you prepare the reaction plate.

Calculations

Determine the quantity of primer to be added to the reaction mix by performing the following calculation:

Concentration (initial) C1 x Volume (primer stock) V1 = Concentration (final) C2 × Volume (final reaction) V2

(10 µM) × (V1) = (0.05 µM) (20 µL)

V1 = (0.05 × 20) / 10 = 0.1

Prepare the reaction plate

1. Add reaction mix and sample to a tube.
   a. To an appropriately sized tube, add the volumes of reaction mix and sample listed below.

Tube	Unknown reaction	Reaction mix	Reaction mix volume (µL)	Sample	Sample volume (µL)
1	Target 1	Power SYBR reaction mix	7632	Sample 1	848

   b. Mix the reactions by gently pipetting up and down, then cap the tubes.
   c. Centrifuge the tubes briefly to remove air bubbles.

2. Pipet 20 µL of the unknown (sample) reaction to each well in the reaction plate.
3. Seal the reaction plate with optical adhesive film.
4. Centrifuge the reaction plate briefly to remove air bubbles.
5. Confirm that the liquid is at the bottom of each well of the reaction plate. If not, centrifuge the reaction plate again at a higher speed and for a longer period of time.
6. Until you are ready to perform the PCR run, place the reaction plate at 4°C, in the dark.
7. Run the PCR.
8. After the PCR is completed, use the same reaction plate containing the PCR product to run the Melt Curve as described in Chapter 4.

For more information

For more information on...	Refer to...	Publication number
Assigning the reaction plate components	Chapter 2 in Booklet 1, *Getting Started with QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Experiments*	4489822
Sealing the reaction plate	Chapter 2 in Booklet 1, *Getting Started with QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Experiments*	4489822

4 Run the Experiment

This chapter explains how run the example experiment on the QuantStudio™ 6 or 7 Instrument.

This chapter covers:

IMPORTANT! Run the experiment at the same ambient temperature at which you calibrated the instrument. Extreme variations in ambient temperature can affect the heating and cooling of the instrument and influence experimental results.

IMPORTANT! Do not attempt to open the access door during the run. The door is locked while the QuantStudio™ 6 or 7 Instrument is in operation.

Start the run

1. Open the Melt Curve example file that you created using instructions in Chapter 2.

   IMPORTANT! The example experiment includes the melt curve analysis of a PCR product from PCR on the QuantStudio™ 6 or 7 Instrument, or another thermal cycler. To run a Melt Curve on the example file you created in Chapter 2, ensure that PCR has already been performed on the reaction plate you load into the instrument. Absence of the PCR product will lead to no results in the Dissociation Step of the Melt Curve Stage.

2. Load the reaction plate, containing the PCR product, into the instrument.
3. Start the run.

Monitor the run

Monitor the example experiment run:

- From the QuantStudio™ 6 and 7 Flex Real-Time PCR System Software using the Run screen, while the experiment is in progress.
- From the Instrument Console of the QuantStudio™ 6 and 7 Flex Real-Time PCR System Software (to monitor an experiment started from another computer or from the QuantStudio™ 6 or 7 Instrument touchscreen).
- From the QuantStudio™ 6 or 7 Instrument touchscreen.

From the Instrument Console of the QuantStudio™ 6 and 7 Flex Real-Time PCR System Software

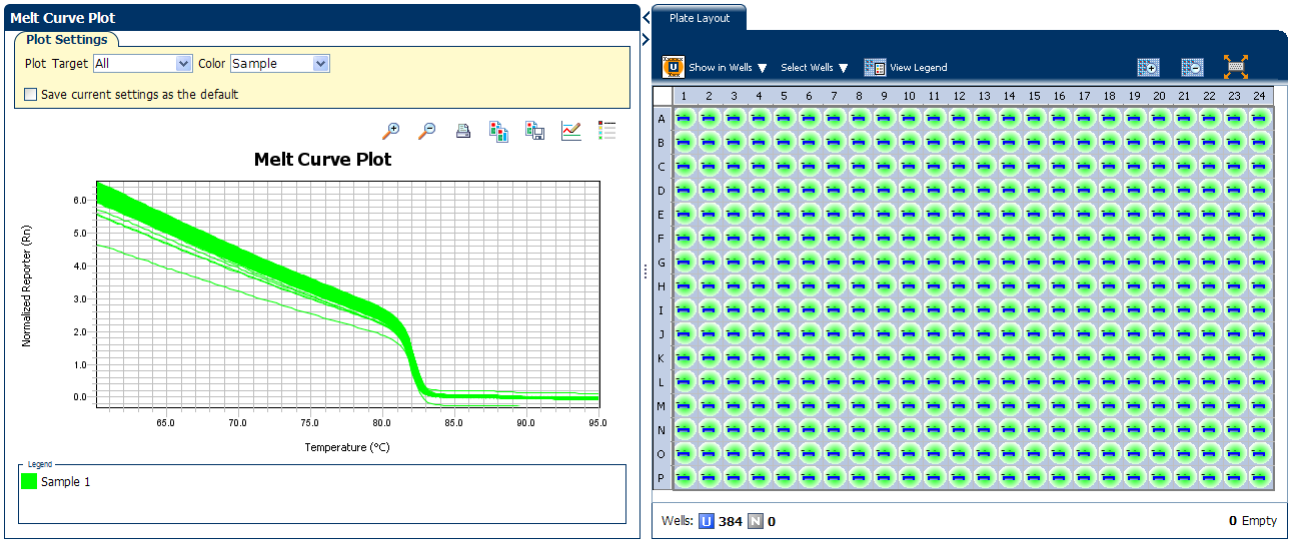
1. In the Instrument Console screen, select the instrument icon.
2. Click **Manage Instrument** or double-click on the instrument icon.
3. On the Manage Instrument screen, click **Monitor Running Instrument** to access the Run screen.

View the Melt Curve

You can view the progress of the run in real time. During the run, periodically view all the three plots available from the QuantStudio™ 6 and 7 Flex Real-Time PCR System Software for potential problems.

Click **Melt Curve** from the Run Experiment Menu, select the Plate Layout tab, then select the wells to view.

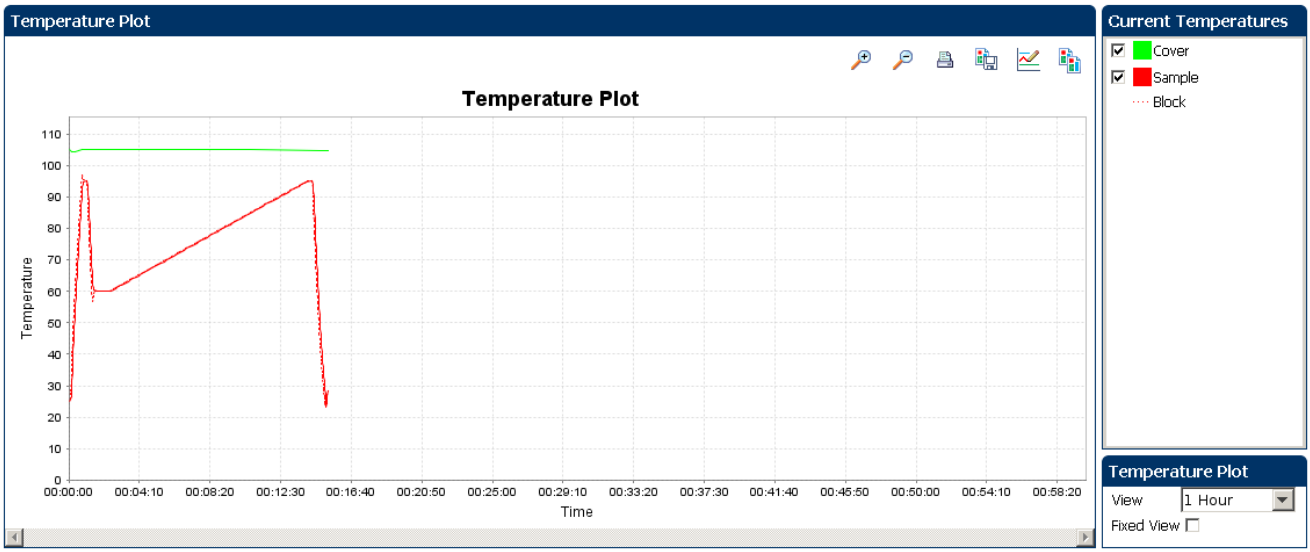
The following is an image of the Melt Curve as it appears at the end of the example experiment.



View the Temperature Plot

Click **Temperature Plot** from the Run Experiment Menu.

The following is an image of the Temperature Plot screen as it appears during the example experiment.

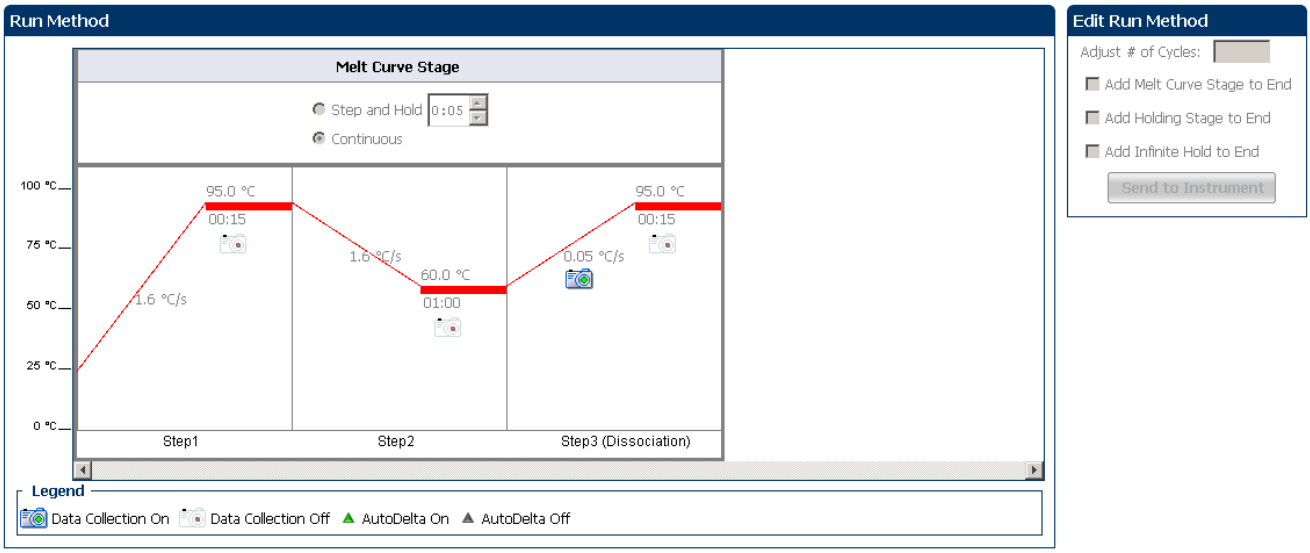


Note: The Sample temperature displayed in the Current Temperatures group is an estimated value.

View the Run Method

Click **Run Method** from the Run Experiment Menu.

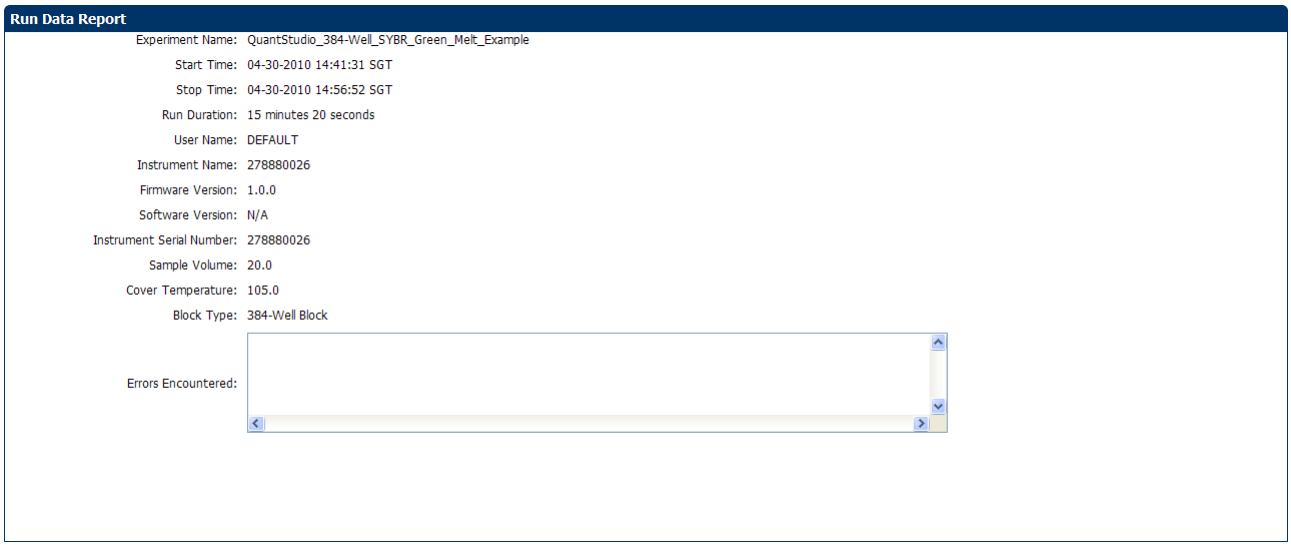
The following is an image of the Run Method screen as it appears in the example experiment.



View run data

Click **View Run Data** from the Run Experiment Menu.

The following is an image of the View Run Data screen as it appears in the example experiment.

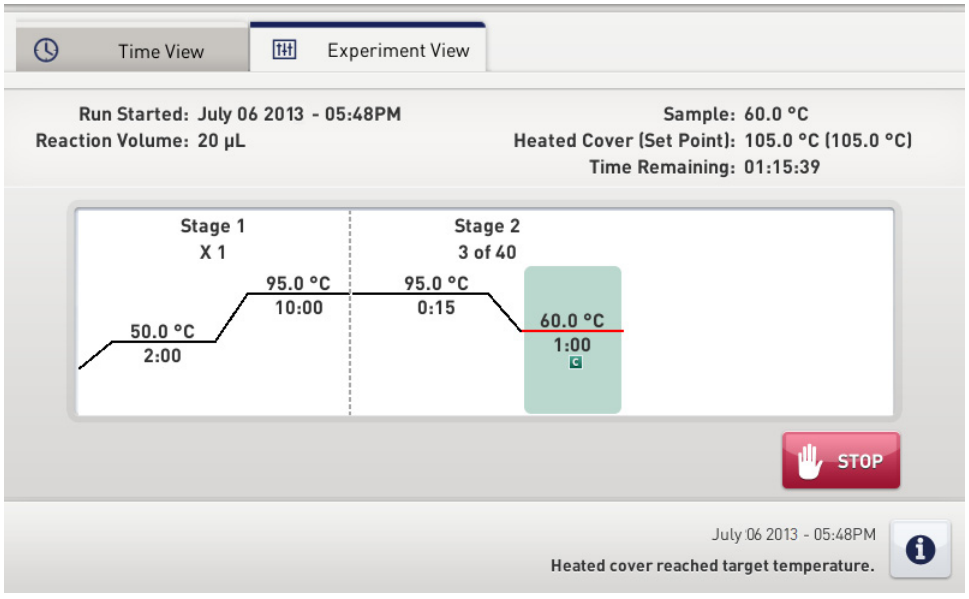


From the QuantStudio™ 6 or 7 Instrument touchscreen

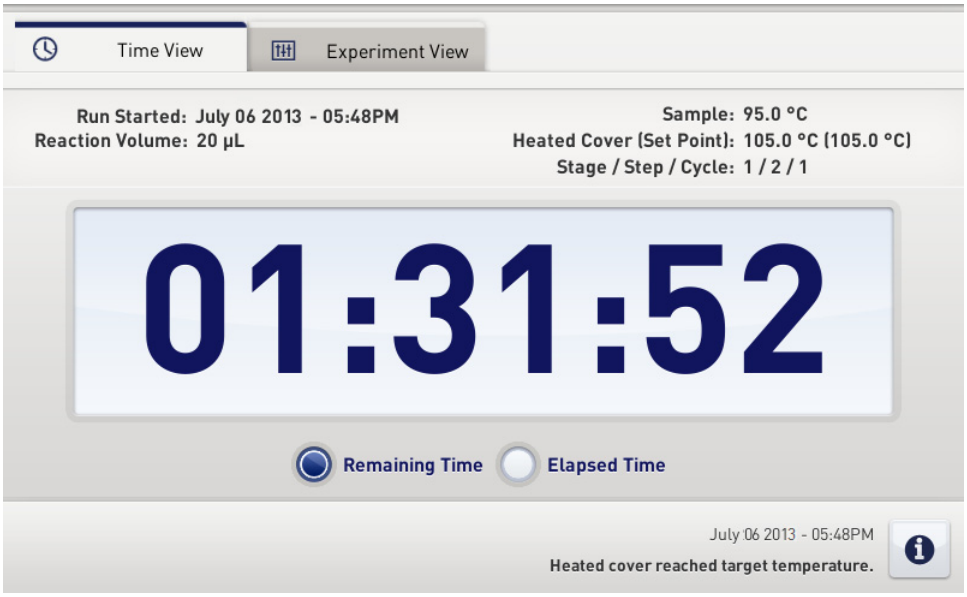
You can also view the progress of the run from the QuantStudio™ 6 or 7 Instrument touchscreen.

The following are images of the Run Method screen on the QuantStudio™ 6 or 7 Instrument touchscreen:

Experiment View



Time View


Time View
Experiment View
Run Started: July 06 2013 - 05:48PM
Reaction Volume: 20 µL
Sample: 95.0 °C
Heated Cover (Set Point): 105.0 °C (105.0 °C)
Stage / Step / Cycle: 1 / 2 / 1
01:31:52
Remaining Time
Elapsed Time
July 06 2013 - 05:48PM
Heated cover reached target temperature.

5 Review Results and Adjust Experiment Parameters

In Section 5.1 of this chapter you review the analyzed data using several of the analysis screens and publish the data. Section 5.2 of this chapter walks you through the process of modifying experiment parameters to troubleshoot problems with experiment results before rerunning an experiment.

This chapter covers:

Section 5.1 Review Results

Analyze the example experiment

1. Open the example experiment file that you ran in Chapter 4.
2. Click **Analyze**. The software analyzes the data using the default analysis settings.

   Note: You can also access the experiment to analyze from the Home screen.

View the Melt Curve Plot

View the Melt Curve Plot as the Derivative Reporter (-Rn) versus the Temperature Plot generated by the target.

The Melt Curve screen displays the melt curve of the targets in the selected wells. Use the Melt Curve plots to confirm the results of the experiment:

- **Normalized Reporter (Rn) vs. Temperature** – This plot displays the fluorescence signal from the reporter dye normalized to the fluorescence signal of the passive reference. You can use this plot to see the change in Rn with change in the temperature. You cannot use this plot to determine the Tm of the target.
- **Derivative Reporter (-Rn) vs. Temperature** – This plot displays the derivative reporter signal in the y-axis. The peaks in the plot indicate significant decrease in SYBR® Green signal, and therefore the Tm of the target.

Purpose

The purpose of viewing the Melt Curve Plot for the example experiment is to review the melting temperature of the target.

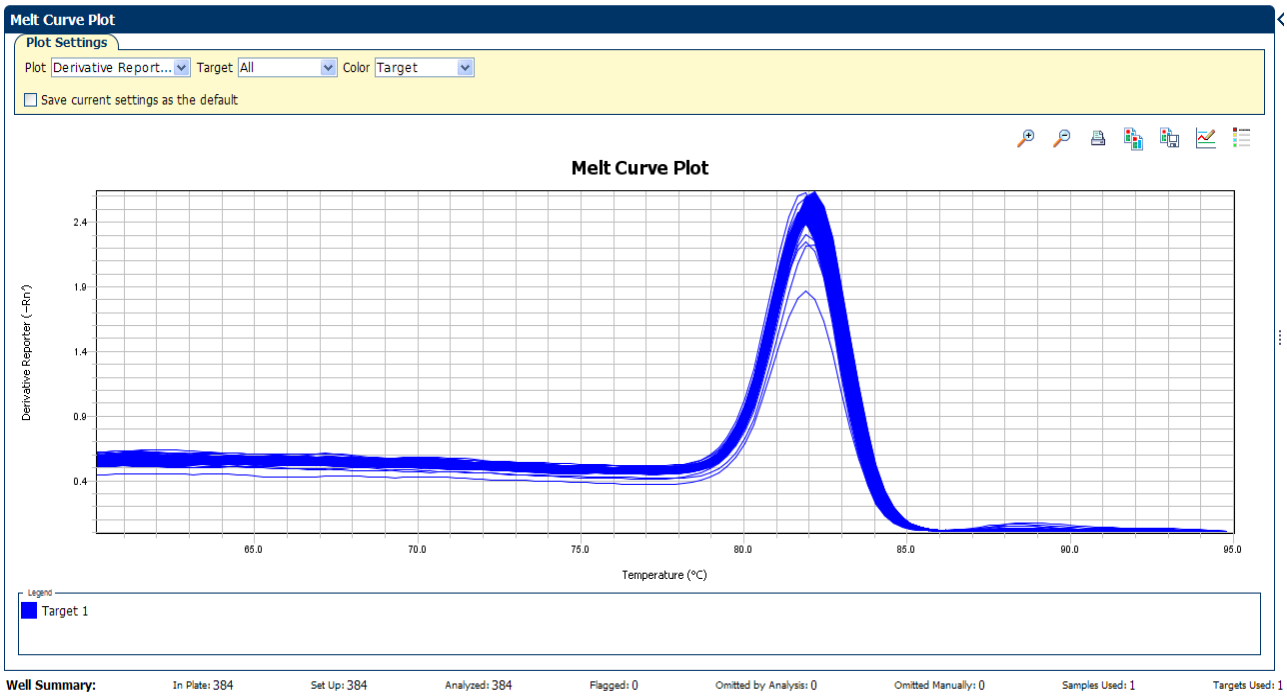
To view and assess the Melt Curve

1. From the Experiment menu pane, select **Analysis ▸ Melt Curve Plot.**

   Note: If no data are displayed, click **Analyze**.
2. Enter the Plot Settings:

Menu	Selection
Plot	Derivative Reporter
Target	All
Plot Color	Target
 (This is a toggle button. When the legend is displayed, the button changes to **Hide the plot legend**.)	Check (default)

The following is an image of the Melt Curve for the example experiment:



Tips for viewing melt curves in your own experiments

When you analyze your own Melt Curve experiment, look for wells with multiple peaks, indicating non-specific amplifications or primer dimer formation.

If your experiment does not amplify properly or indicates non-specific amplification, troubleshoot by manually adjusting the Melt Curve settings (see "Adjust analysis settings" on page 35).

Identify well problems using the Well Table

Review the details of the experiment results in the well table and identify any flagged wells. The well table displays the assay-specific setup and analysis properties for the experiment in a tabular format.

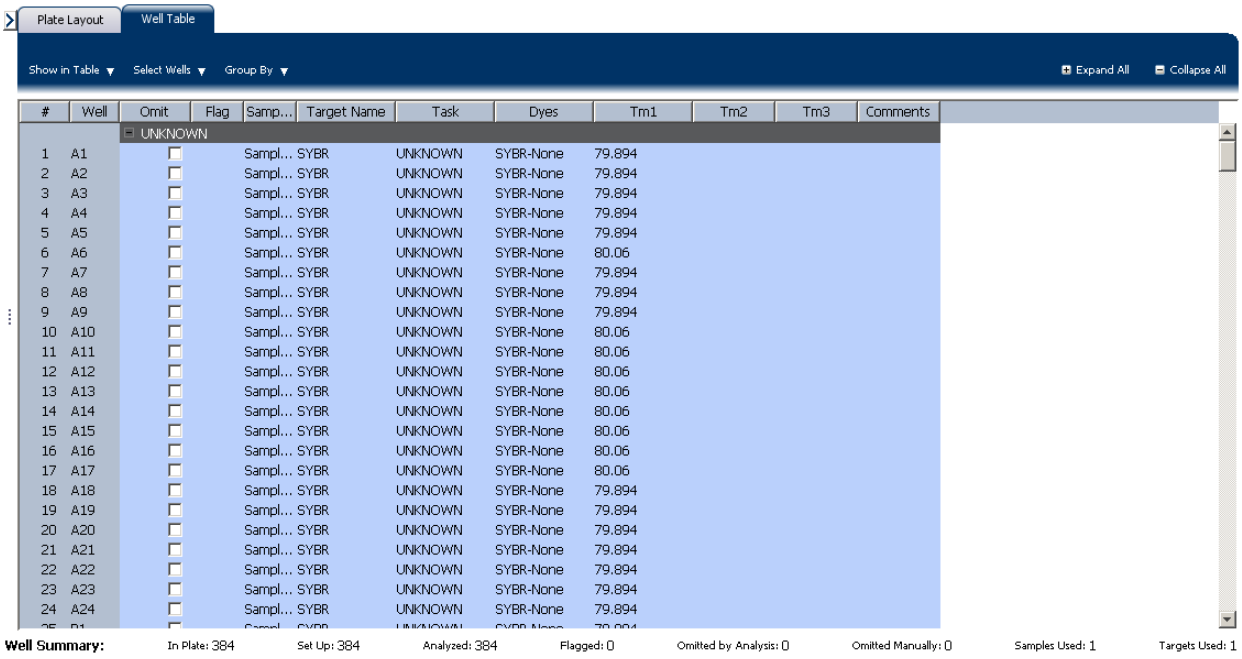
Example experiment values and flags

For the example experiment, confirm that no wells of the reaction plate triggered QC flags ⚠.

View the well table

1. Select the **Well Table** tab.
2. Click the **Flag** column header to sort the data so that the wells that triggered flags appear at the top of the table.
3. Confirm the integrity of the controls:
   a. From the Group By menu, select **Task** to organize the table rows by their function on the reaction plate.
   b. Confirm that each of the controls do not display flags (⚠1).

The following is an image of the well table of the example Melt Curve experiment.



The following table gives the description of each column in the well table.

Column	Description
Well	The position of the well on the reaction plate.
Omit	A check mark indicates that the well has been removed from the analysis.

Column	Description
Flag	A ⚠1 indicates that the well triggered the number of flags listed inside the symbol.
Sample Name	The name of the sample.
Target Name	The name of the target evaluated by the well.
Task	The task assigned to the well (Unknown, Negative Control, or Positive Control).
Dyes	The name of the reporter and quencher dyes of the associated sample for the target evaluated by the well.
Tm1	The melting temperature of the target.
Tm2	The second melting temperature (for targets with multiple melting temperatures).
Tm3	The third melting temperature (for targets with multiple melting temperatures).

Tips for viewing well tables your own experiments

When you analyze your own experiment:

- Review the data for the Unknown samples. For each row that displays ⚠ in the Flag column, note the data and the flag(s) triggered by the associated well.
- Select areas of the table or wells of a specified type by:
  - Left-clicking the mouse and dragging across the area you want to select an area of the table.
  - Selecting **Sample**, **Target**, or **Task** from the Select Items menu in the Well Table tab, then selecting the sample, target, or task name from the second Select Items menu to select wells of a specific type using the well-selection tool.
- Group the rows of the plate layout by selecting an option from the Group By menu. You can then collapse or expand the lists either by clicking the **+/-** icon next to individual lists, or by clicking ⊟ **Collapse All** or ⊞ **Expand All**.
- Omit a well from the analysis by selecting the **Omit** check box for that well. To include the well in the analysis, deselect the **Omit** check box.

  Note: You must reanalyze the experiment each time you omit or include a well.

Confirm accurate dye signal using the Multicomponent Plot

The Multicomponent Plot screen displays the complete spectral contribution of each dye in a selected well over the duration of the PCR run.

Purpose

In the Melt Curve example experiment, you review the Multicomponent Plot screen for:

- ROX™ dye (passive reference)
- SYBR® dye (reporter)
- Spikes, dips, and/or sudden changes

View the Multicomponent Plot

1. From the Experiment Menu pane, select **Analysis ▸ Multicomponent Plot**.

   Note: If no data are displayed, click **Analyze**.

2. Display the unknown wells in the plate layout to display the corresponding data in the Multicomponent Plot screen:

   a. Click the **Plate Layout** tab.

   b. Select one well in the plate layout; the well is shown in the Multicomponent Plot screen.

      Note: If you select multiple wells, the Multicomponent Plot screen displays the data for all selected wells simultaneously.

3. From the Plot Color drop-down menu, select **Dye**.

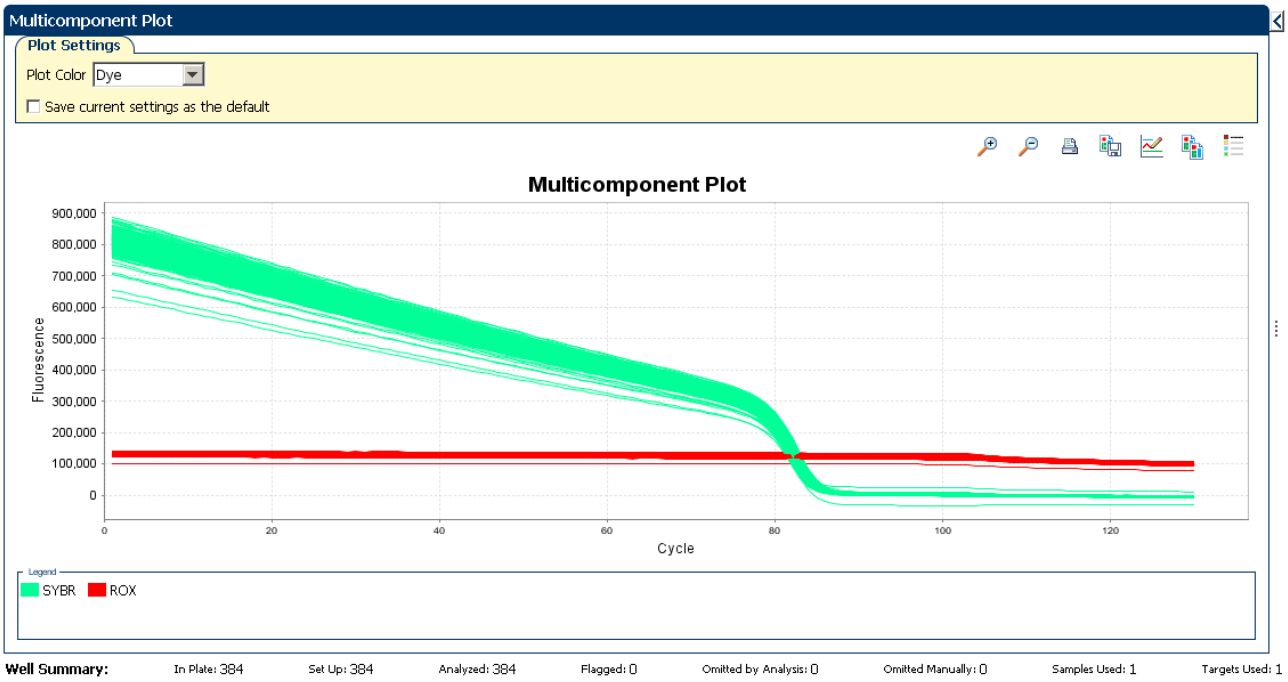
4. Click  **Show a legend for the plot** (default).

   Note: This is a toggle button. When the legend is displayed, the button changes to Hide the plot legend.

5. Check the ROX dye signal. In the example experiment, the ROX dye signal remains constant throughout the PCR process; a constant ROX dye signal indicates typical data.

6. Check the SYBR® dye signal. In the example experiment, because the PCR run has already been completed, the SYBR® dye signal shows gradual decrease throughout the run and a sudden dip in the fluorescence at one point; the sudden drop in the SYBR® dye signal indicates the melting temperature of the target.

The following is an image of the Multicomponent Plot screen for the example experiment:



Tips for confirming dye accuracy in your own experiment

When you analyze your own Melt Curve experiment, look for:

- **Passive reference** – The passive reference dye fluorescence level should remain relatively constant throughout the PCR process.
- **Reporter dye** – The reporter dye fluorescence level should display a flat region corresponding to the baseline, followed by a rapid rise in fluorescence as the amplification proceeds. If the Melt Curve is being performed post-PCR, then there should be a gradual decrease in fluorescence and a sudden dip indicating the melting temperature of the target.
- **Irregularities in the signal** – There should not be any spikes, dips, and/or sudden changes in the fluorescent signal.

Determine signal accuracy using the Raw Data Plot

The Raw Data Plot screen displays the raw fluorescence signal (not normalized) for each optical filter for the selected wells during each cycle of the real-time PCR.

About the example experiment

In the Melt Curve example experiment, you review the Raw Data Plot screen for a stable increase in signal (no abrupt changes or dips) from the appropriate filter.

View the Raw Data Plot

1. From the Experiment Menu pane, select **Analysis** ▸ **Raw Data Plot**.

   Note: If no data are displayed, click **Analyze**.

2. Display all 384 wells in the Raw Data Plot screen by clicking the upper left corner of the plate layout in the Plate Layout tab.

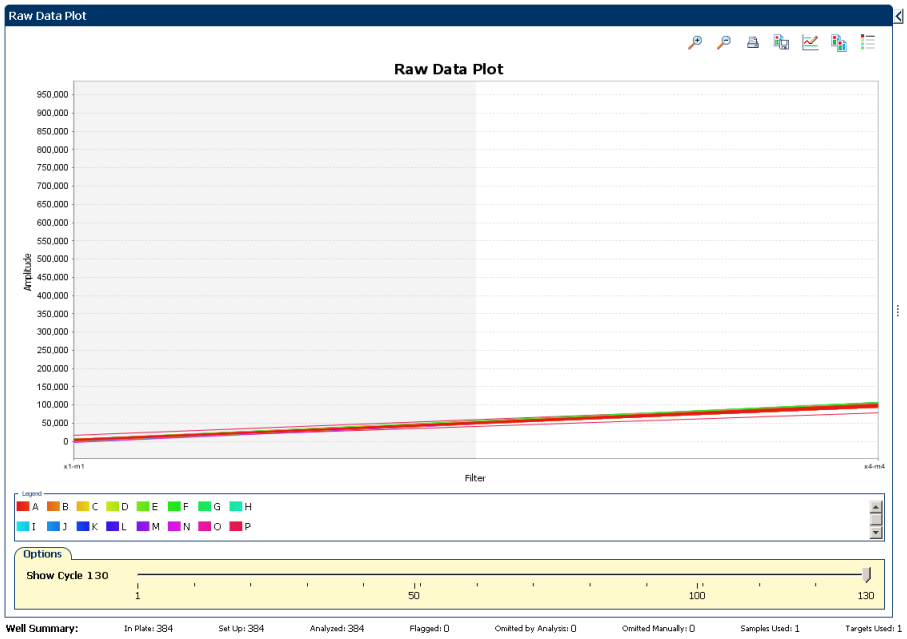
3. Click  **Show a legend for the plot** (default). The legend displays the color code for each row of the reaction plate (see the legend in the Raw Data Plot shown below).

4. Click and drag the Show Cycle pointer from cycle 1 to cycle 130. In the example experiment, the signal from filter 1, which corresponds to the SYBR® dye filter, is stable throughout.

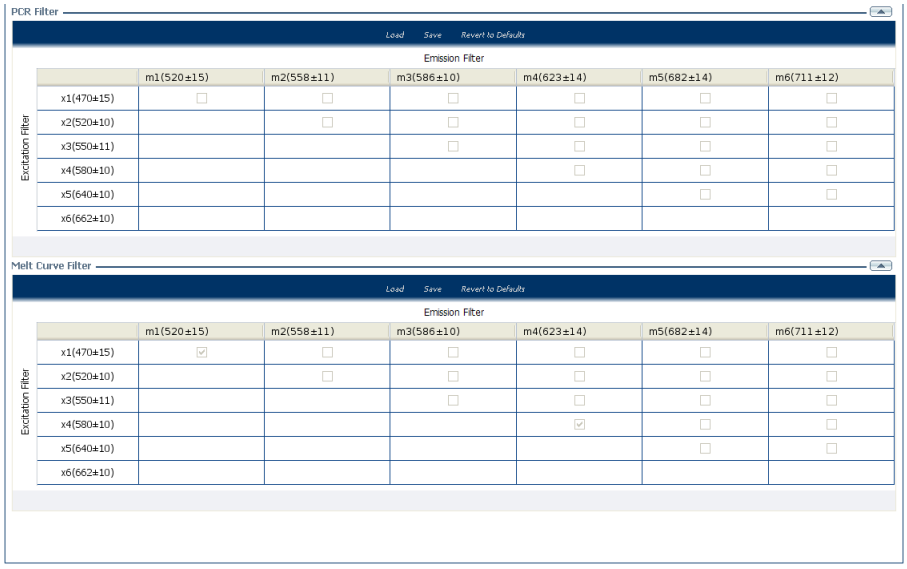
   Note: The readings shown below are from the example experiment. Actual results will vary with individual experiment setup.

   Note: The cycle number in the Melt Curve represents the number of data collection points for that experiment.

The following is an image of the Raw Data plot for the example experiment:



The filters used for the example experiment are:



Tips for determining signal accuracy in your own experiments

When you analyze your own Melt Curve experiment, look for the following in each filter:

- Characteristic signal growth
- No abrupt changes or dips

Review the flags in the QC Summary

The QC Summary screen displays a list of the QuantStudio™ 6 and 7 Flex Software flags, including the flag frequency and location for the open experiment.

View the QC Summary

1. From the Experiment Menu pane, select **Analysis** ▸ **QC Summary**.

   Note: If no data are displayed, click **Analyze**.

2. Review the Flags Summary.

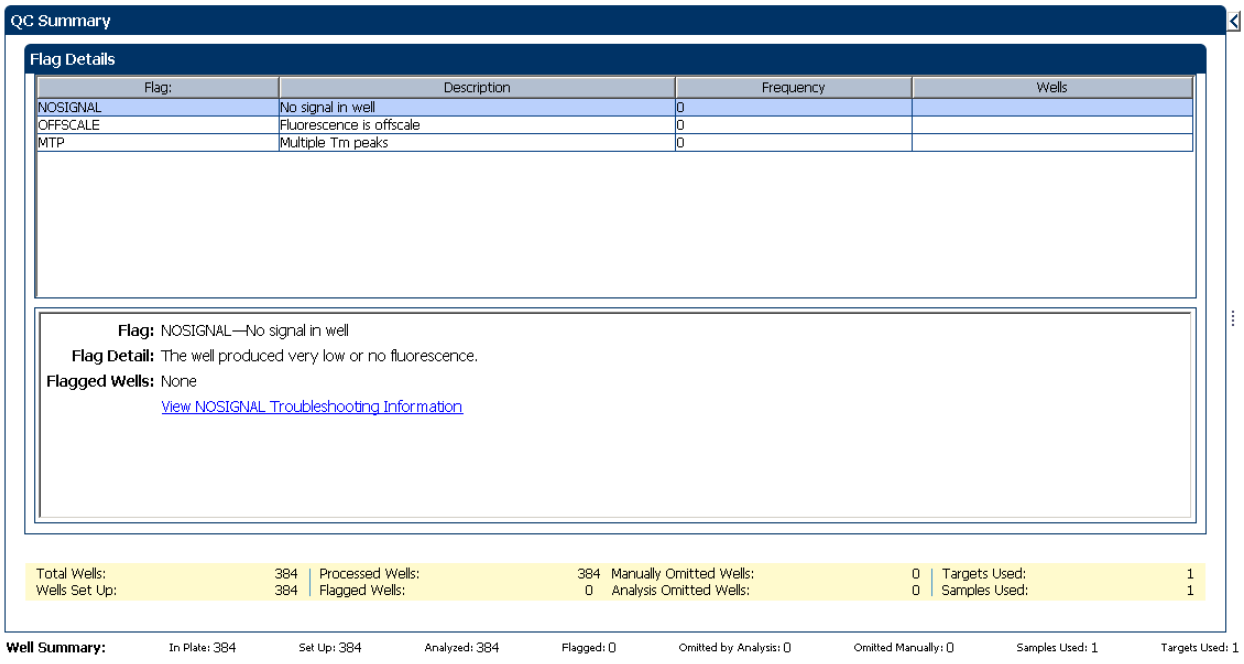
   Note: A 0 displayed in the Frequency column indicates that the flag does not appear in the experiment. If the frequency is >0, the flag appears somewhere in the experiment; the well position is listed in the Wells column.

   In the example experiment, there are no flagged wells.

3. In the Flag Details table, click each flag with a frequency >0 to display detailed information about the flag. In the example experiment, the Frequency column displays 0 for the three flags NOSIGNAL, OFFSCALE, and MTP.

4. *(Optional)* For those flags with frequency >0, click the troubleshooting link to view information on correcting the flag.

   The following is an image of the QC Summary for the example experiment:



Possible flags

For Melt Curve experiments that do not include amplification, the flags listed below may be triggered by the experiment data.

Flag	Description
Pre-processing flag	
OFFSCALE	Fluorescence is offscale
Primary analysis flag	
NOSIGNAL	No signal in well
Secondary analysis flag	
MTP	Multiple Tm peaks

For Melt Curve experiments that include amplification, the flags listed below may be triggered by the experiment data.

Flag	Description
Pre-processing flag	
OFFSCALE	Fluorescence is offscale
Primary analysis flags	
BADROX	Bad passive reference signal
NOAMP	No amplification
NOISE	Noise higher than others in plate
SPIKE	Noise spikes
NOSIGNAL	No signal in well
EXPFAIL	Exponential algorithm failed
BLFAIL	Baseline algorithm failed
THOLDFAIL	Thresholding algorithm failed
CTFAIL	C_T algorithm failed
Secondary analysis flags	
MTP	Multiple Tm peaks
OUTLIERRG	Outlier in replicate group
AMPNC	Amplification in negative control
HIGHSD	High standard deviation in replicate group

For more information

For more information on...	Refer to...	Publication number
Publishing data	Chapter 2 in Booklet 1, *Getting Started with QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Experiments*	4489822

Section 5.2 Adjust parameters for re-analysis of your own experiments

Adjust analysis settings

The Analysis Settings dialog box displays the analysis settings for the Melt Curve and flags.

If the default analysis settings in the QuantStudio™ 6 and 7 Flex Software are not suitable for your own experiment, you can change the settings in the Analysis Settings dialog box, then reanalyze your experiment.

View the analysis settings

1. From the Experiment Menu pane, select **Analysis**.
2. Click **Analysis** ▸ **Analysis Settings** to open the Analysis Settings dialog box.

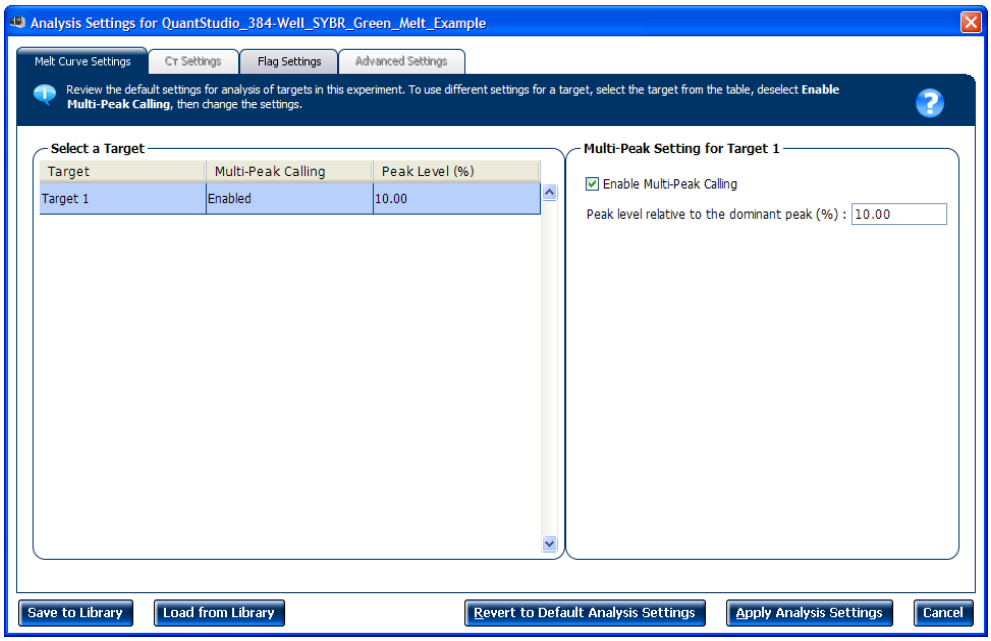
   In the example experiment, the default analysis settings are used for each tab:

   - Melt Curve Settings
   - C_T Settings
   - Flag Settings
   - Advanced Settings

   Note: The C_T Settings and Advanced Settings tabs appear in the Analysis Settings dialog box only if the Melt Curve experiment you are performing includes the PCR process.

   Note: Select the **Include PCR** check box on the Experiment Properties screen to include amplification in your Melt Curve experiment.

   The following is an image of the Analysis Settings dialog box for a Melt Curve experiment:



3. View and, if necessary, change the analysis settings (see "Adjust analysis settings" below).

   Note: You can save the changes to the analysis settings to the Analysis Settings Library for later use. For more information, see About the Analysis Settings Library in Booklet 1, *Getting Started with QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Experiments*.

4. Click **Apply Analysis Settings** to apply the current analysis settings.

   Note: You can go back to the default analysis settings, by clicking **Revert to Default Analysis Settings**.

Adjust analysis settings

You may change the following settings:

Melt Curve Settings

Use this tab to:

- Enable or disable multi-peak calling.
  - Select the **Enable the Multi-Peak Calling** check box if you expect to amplify more than 1 PCR product and you want to determine the Tm for more than one peak.
  - Deselect the **Enable the Multi-Peak Calling** check box if you expect to amplify 1 PCR product and you do not want to determine the Tm for more than one peak.
- Enter a value (in percentage) for the peak level relative to the dominant peak.

  Specify a fractional level value as the peak detection threshold. The detected peaks are measured relative to the height of the tallest peak, which has a perfect fractional level 100%. The default value is initially set at 10%.

  For example, if you set a fractional level detection threshold value at 40, then only peaks above 40% of the tallest peak are reported and the peaks at lower height are regarded as noise.

C_T Settings

- **Data Step Selection**

  Use this feature to select one stage/step combination for C_T analysis when there is more than one data collection point in the run method.
- **Algorithm Settings**

  Use the Baseline Threshold algorithm to determine the C_T values.

  The Baseline Threshold algorithm is an expression estimation algorithm that subtracts a baseline component and sets a fluorescent threshold in the exponential region for gene quantification.
- **Default C_T Settings**

  Use the default C_T settings feature to calculate C_T for the targets that do not have custom settings. To edit the default settings, click **Edit Default Settings**.

- **C_T Settings for Target**

  When you manually set the threshold and baseline, Life Technologies recommends:

Setting	Recommendation
Threshold	Enter a value for the threshold so that the threshold is: • Above the background. • Below the plateau and linear regions of the amplification curve. • Within the exponential phase of the amplification curve.
Baseline	Select the Start Cycle and End Cycle values so that the baseline ends before significant fluorescent signal is detected.

  Note: This setting is applicable only to the Baseline Threshold algorithm.

  Note: Selecting Automatic Threshold implies selection of automatic setting of the baseline. However, if Automatic Threshold is deselected, then you can choose between setting the baseline either automatically or manually.

Flag Settings

Use the Flag Settings tab to:

- Adjust the sensitivity so that more wells or fewer wells are flagged.
- Change the flags that are applied by the QuantStudio™ 6 and 7 Flex Software.

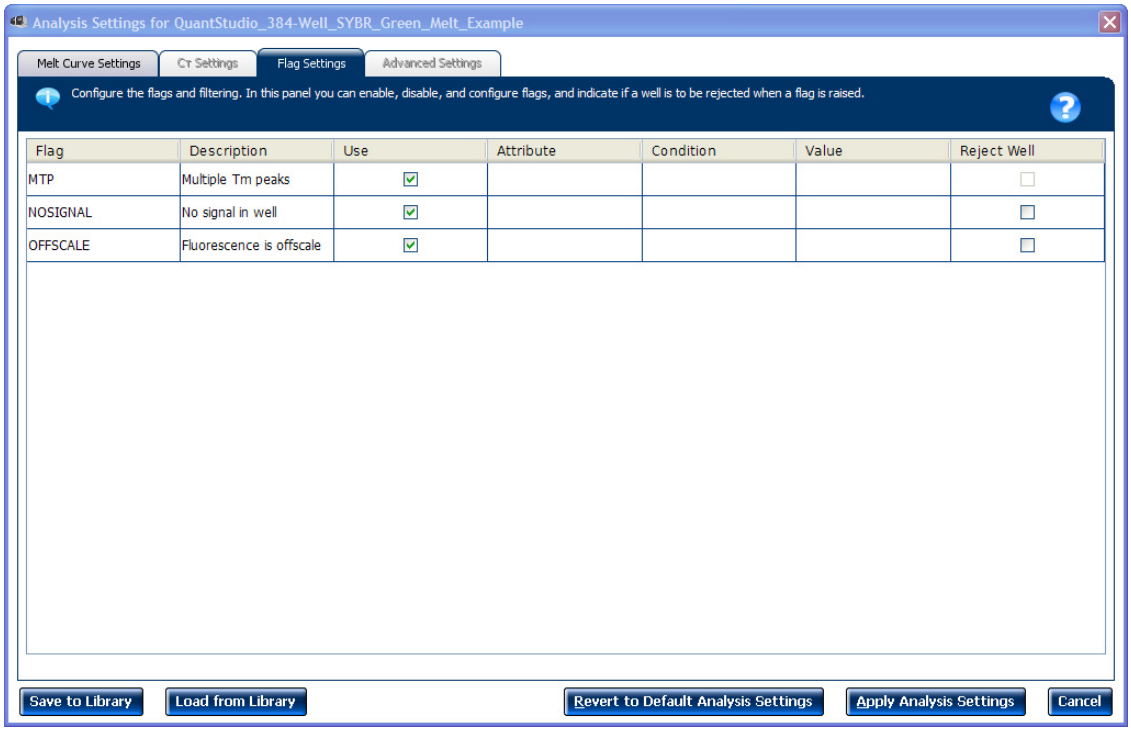
To adjust the flag settings:

1. In the Use column, select the check boxes for flags to apply during analysis.
2. *(Optional)* If an attribute, condition, and value are listed for a flag, specify the setting for applying the flag.

   Note: If you choose to adjust the setting for applying a flag, make minor adjustments as you evaluate the appropriate setting.
3. In the Reject Well column, select the check boxes if you want the software to reject wells with the flag.

   Note: After you have rejected the flagged wells, analysis results depend on factors such as the experiment type and flag type. For example, rejecting wells flagged by HIGHSD in experiments using the Standard Deviation calculations may change the result of C_T SD. For some flags, analysis results calculated before the well is rejected are maintained.
4. Click **Apply Analysis Settings** in the Analysis Settings dialog box. If the run status is complete, the data are reanalyzed.

The following is an image of the Flag Settings tab:



Advanced Settings

Use the Advanced Settings tab to change baseline settings well-by-well.

To use custom baseline settings for a well-target combination:

1. Select one or more well-target combinations in the table.
2. Deselect the **Use C_T Settings Defined for Target** check box.
3. Define the custom baseline settings:
   - For automatic baseline calculations, select the **Automatic Baseline** check box.
   - To define the baseline, deselect the **Automatic Baseline** check box, then enter the baseline start cycle and baseline end cycle.

For more information

For more information on...	Refer to	Publication number
Amplification efficiency	*Amplification Efficiency of TaqMan® Gene Expression Assays Application Note*.	127AP05-03

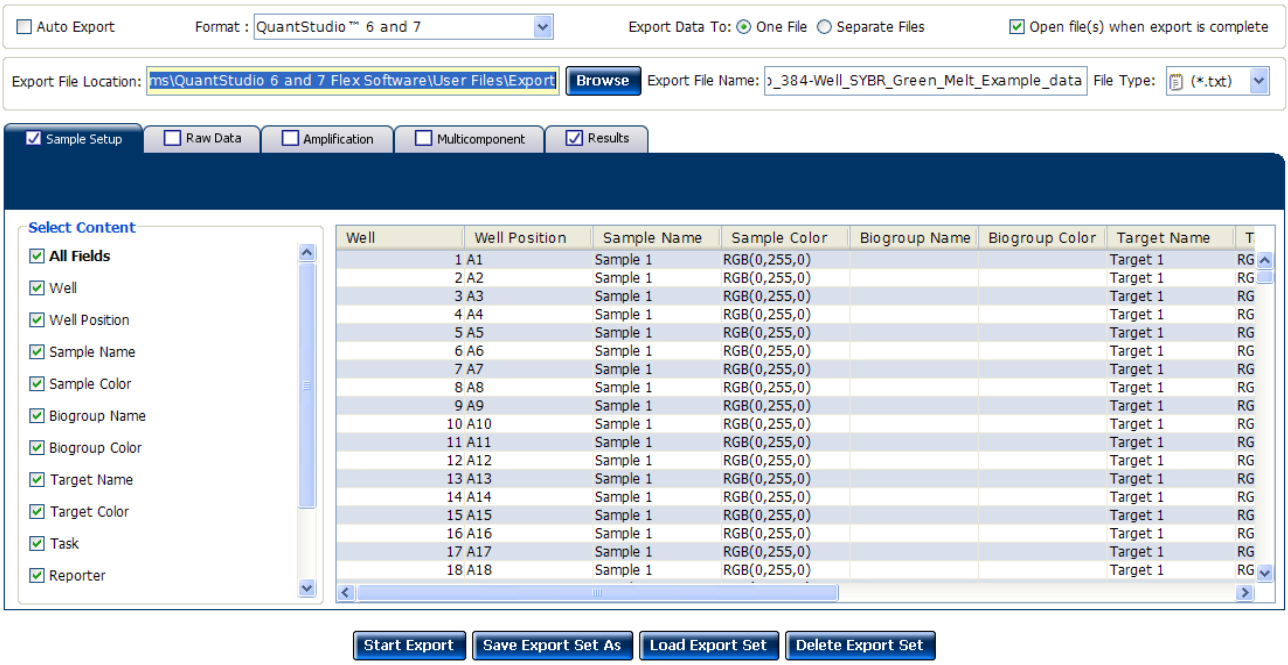
6 Export Experiment Results

1. Open the Melt Curve example experiment file that you analyzed in Chapter 5.
2. In the Experiment Menu, click **Export.**

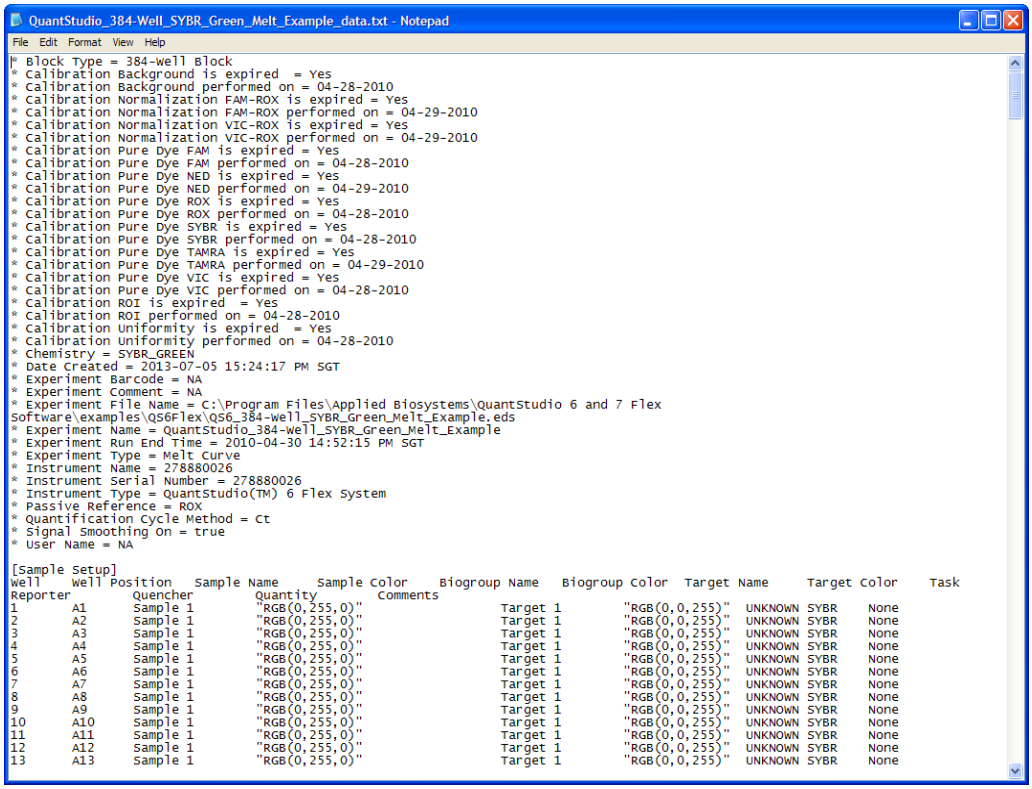
   Note: To export data automatically after analysis, select the **Auto Export** check box during experiment setup or before running the experiment. Auto export is unchecked for the example experiment.
3. Select **QuantStudio™ 6 and 7** format.
4. Complete the Export dialog box as shown below:

Field or Selection	Entry
Select Data to export/ Select Content	Results
Export Data To	One File
Export File Name	QuantStudio_384-Well_SYBR_Green_Melt_Example_data
File Type	*.txt
Export File Location	C:\Applied Biosystems\QuantStudio 6 and 7 Flex Software\User Files\Export

Your Export screen should look like this:



Your exported file when opened in Notepad should look like this:



QuantStudio_384-Well_SYBR_Green_Melt_Example_data.txt - Notepad

File Edit Format View Help

```
* Block Type = 384-Well Block
* Calibration Background is expired  = Yes
* Calibration Background performed on = 04-28-2010
* Calibration Normalization FAM-ROX is expired = Yes
* Calibration Normalization FAM-ROX performed on = 04-29-2010
* Calibration Normalization VIC-ROX is expired = Yes
* Calibration Normalization VIC-ROX performed on = 04-29-2010
* Calibration Pure Dye FAM is expired = Yes
* Calibration Pure Dye FAM performed on = 04-28-2010
* Calibration Pure Dye NED is expired = Yes
* Calibration Pure Dye NED performed on = 04-29-2010
* Calibration Pure Dye ROX is expired = Yes
* Calibration Pure Dye ROX performed on = 04-28-2010
* Calibration Pure Dye SYBR is expired = Yes
* Calibration Pure Dye SYBR performed on = 04-28-2010
* Calibration Pure Dye TAMRA is expired = Yes
* Calibration Pure Dye TAMRA performed on = 04-29-2010
* Calibration Pure Dye VIC is expired = Yes
* Calibration Pure Dye VIC performed on = 04-28-2010
* Calibration ROI is expired  = Yes
* Calibration ROI performed on = 04-28-2010
* Calibration Uniformity is expired  = Yes
* Calibration Uniformity performed on = 04-28-2010
* Chemistry = SYBR_GREEN
* Date Created = 2013-07-05 15:24:17 PM SGT
* Experiment Barcode = NA
* Experiment Comment = NA
* Experiment File Name = C:\Program Files\Applied Biosystems\QuantStudio 6 and 7 Flex
Software\examples\QS6Flex\QS6_384-Well_SYBR_Green_Melt_Example.eds
* Experiment Name = QuantStudio_384-Well_SYBR_Green_Melt_Example
* Experiment Run End Time = 2010-04-30 14:52:15 PM SGT
* Experiment Type = Melt Curve
* Instrument Name = 278880026
* Instrument Serial Number = 278880026
* Instrument Type = QuantStudio(TM) 6 Flex System
* Passive Reference = ROX
* Quantification Cycle Method = Ct
* Signal Smoothing On = true
* User Name = NA

[Sample Setup]
well    Well Position   Sample Name     Sample Color    Biogroup Name   Biogroup Color  Target Name     Target Color    Task
Reporter        Quencher        Quantity        Comments
1       A1      Sample 1        "RGB(0,255,0)"                  Target 1        "RGB(0,0,255)"  UNKNOWN SYBR    None
2       A2      Sample 1        "RGB(0,255,0)"                  Target 1        "RGB(0,0,255)"  UNKNOWN SYBR    None
3       A3      Sample 1        "RGB(0,255,0)"                  Target 1        "RGB(0,0,255)"  UNKNOWN SYBR    None
4       A4      Sample 1        "RGB(0,255,0)"                  Target 1        "RGB(0,0,255)"  UNKNOWN SYBR    None
5       A5      Sample 1        "RGB(0,255,0)"                  Target 1        "RGB(0,0,255)"  UNKNOWN SYBR    None
6       A6      Sample 1        "RGB(0,255,0)"                  Target 1        "RGB(0,0,255)"  UNKNOWN SYBR    None
7       A7      Sample 1        "RGB(0,255,0)"                  Target 1        "RGB(0,0,255)"  UNKNOWN SYBR    None
8       A8      Sample 1        "RGB(0,255,0)"                  Target 1        "RGB(0,0,255)"  UNKNOWN SYBR    None
9       A9      Sample 1        "RGB(0,255,0)"                  Target 1        "RGB(0,0,255)"  UNKNOWN SYBR    None
10      A10     Sample 1        "RGB(0,255,0)"                  Target 1        "RGB(0,0,255)"  UNKNOWN SYBR    None
11      A11     Sample 1        "RGB(0,255,0)"                  Target 1        "RGB(0,0,255)"  UNKNOWN SYBR    None
12      A12     Sample 1        "RGB(0,255,0)"                  Target 1        "RGB(0,0,255)"  UNKNOWN SYBR    None
13      A13     Sample 1        "RGB(0,255,0)"                  Target 1        "RGB(0,0,255)"  UNKNOWN SYBR    None
```

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lifetechnologies.com

October 2013

USER GUIDE

applied biosystems®
by life technologies™

QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Experiments - Appendixes

Booklet 7

Publication Number 4489822
Revision A

life technologies™

Contents

A

Ordering Information

This appendix covers:

How to order from the Life Technologies website

You can order materials accessories directly from the Life Technologies store over the internet.

Note: Product availability and pricing may vary according to your region or country. Online ordering through the Life Technologies store is not available in all countries. Contact your local Life Technologies representative for help.

To order through the website:

- Confirm that your computer has an Internet connection.
- We recommend the following browsers and Adobe® Acrobat® Reader® Software versions to use the Life Technologies website:

Operating system	Microsoft® Internet Explorer®	Apple® Safari®	Mozilla® Firefox®	Adobe® Acrobat® Reader®
Microsoft® Windows®	v6.x or later	None†	v2.x or later	v4.0 or later
Macintosh®	None†	v2.0.4 or later		

† Browser not available for this platform.

Note: Confirm that cookies and Javascript® are turned on for the website to function correctly.

To purchase reagents, accessories, and calibration kits:

1. Go to **www.lifetechnologies.com**
2. Under "I Want to Buy," select the product of interest.

Reagents and consumables

The reagents and consumables listed below are required for calibrating and for performing experiments with the QuantStudio™ 6 and 7 Flex Software.

Note: For reagent or consumable shelf-life expiration date, see the package label.

Calibration reagents and consumables

The following table shows the reagents and consumables required to calibrate and verify the performance of the QuantStudio™ 6 and 7 Instruments when run with the QuantStudio™ 6 and 7 Flex Software.

384-well sample block

Consumable	Part number	Shelf-life at environmental temperature	Storage conditions (°C)
384-Well Spectral Calibration Plate with FAM™ Dye	4432271	Use the consumable by the expiration date mentioned on the package	–15°C to –25°C
384-Well Spectral Calibration Plate with VIC® Dye	4432278		
384-Well Spectral Calibration Plate with ROX™ Dye	4432284		
384-Well Spectral Calibration Plate with SYBR® Green Dye	4432290		
384-Well Spectral Calibration Plate with TAMRA™ Dye	4432296		
384-Well Spectral Calibration Plate with NED™ Dye	4432302		
384-Well Region of Interest (ROI) and Background Plates	4432320		
384-Well Normalization Plates with FAM™/ROX™ and VIC®/ROX™ Dyes	4432308		
TaqMan® RNase P Fast 384-Well Instrument Verification Plate	4455280		

96-well (0.2 mL) sample block

Consumable	Part number	Shelf-life at environmental temperature	Storage conditions (°C)
96-Well Spectral Calibration Plate with FAM™ Dye	4432327	Use the consumable by the expiration date mentioned on the package	–15°C to –25°C
96-Well Spectral Calibration Plate with VIC® Dye	4432334		
96-Well Spectral Calibration Plate with ROX™ Dye	4432340		
96-Well Spectral Calibration Plate with SYBR® Green Dye	4432346		
96-Well Spectral Calibration Plate with TAMRA™ Dye	4432352		
96-Well Spectral Calibration Plate with NED™ Dye	4432358		
TaqMan® RNase P 96-Well Instrument Verification Plate	4432382		
96-Well Region of Interest (ROI) and Background Plates	4432364		
96-Well Normalization Plates with FAM™/ROX™ and VIC®/ROX™ Dyes	4432370		

Fast 96-well (0.1 mL) sample block

Consumable	Part number	Shelf-life at environmental temperature	Storage conditions (°C)
Fast 96-Well Spectral Calibration Plate with FAM™ Dye	4432389	Use the consumable by the expiration date mentioned on the package	–15°C to –25°C
Fast 96-Well Spectral Calibration Plate with VIC® Dye	4432396		
Fast 96-Well Spectral Calibration Plate with ROX™ Dye	4432402		
Fast 96-Well Spectral Calibration Plate with SYBR® Green Dye	4432408		
Fast 96-Well Spectral Calibration Plate with TAMRA™ Dye	4432414		
Fast 96-Well Spectral Calibration Plate with NED™ Dye	4432420		
Fast 96-Well Region of Interest (ROI) and Background Plates	4432426		
Fast 96-Well Normalization Plates with FAM™/ROX™ and VIC®/ROX™ Dyes	4432432		
TaqMan® RNase P Fast 96-Well Instrument Verification Plate	4351979		

Array card sample block

Consumable	Part number	Shelf-life at environmental temperature	Storage conditions (°C)
Array Card Spectral Dye Calibration Kit	4432376	Use the consumable by the expiration date mentioned on the package	–15°C to –25°C
Array Card RNase P Instrument Verification Kit	4432464		

Note: To prepare calibration plates, refer to the instrument user guide.

Note: The Array card sample block is applicable only to the QuantStudio™ 7 Flex System.

Experiment consumables

The following table shows the consumables required to perform experiments with the QuantStudio™ 6 and 7 Flex Software.

96-well (0.2 mL) sample block

Consumable		Part number
MicroAmp® Optical 8-Cap Strip	300 strips	4323032
MicroAmp® Optical 8-Tube Strip (0.2 mL)	125 strips	4316567
MicroAmp® Optical Tube without cap (0.2 mL)	2000 tubes	N8010933
MicroAmp® 96-Well Tray/Retainer Set (Blue) (for 0.2 mL)	10 pairs	4381850
MicroAmp® Optical 96-Well Reaction Plate (0.2 mL)	10 plates	N8010560
MicroAmp® Optical 96-Well Reaction Plate with Barcode (0.2 mL)	10 plates	4306737

Fast 96-well (0.1 mL) sample block

Consumable		Part number
MicroAmp® Fast 8-Tube Strip (0.1 mL)	125 strips	4358293
MicroAmp® Fast Reaction Tube with cap (0.1 mL)	1000 tubes	4358297
MicroAmp® 96-Well Tray (Black) (for 0.1 mL)	10 plates	4379983
MicroAmp® Fast Optical 96-Well Reaction Plate (0.1 mL)	10 plates	4346907
MicroAmp® Fast Optical 96-Well Reaction Plate with Barcode (0.1 mL)	10 plates	4346906

Array card sample block

Consumable		Part number
Array Card	4-pack	4334812
	1-pack	4351471

Note: The Array card sample block is applicable only to the QuantStudio™ 7 Flex System.

Miscellaneous

Consumable		Part number
MicroAmp® Multi-Removal Tool	1 tool	4313950
MicroAmp® Cap Installing Tool (Handle)	1 tool	4330015

Consumable		Part number
MicroAmp® Optical Adhesive Film	25 films	4481193
MicroAmp® Adhesive Film Applicator	5 applicators	4481196
RT-PCR Grade Water	–	AM9935

QuantStudio™ 6 or 7 Instrument accessories

The accessories listed in the following table are for the QuantStudio™ 6 or 7 Instrument when run with the QuantStudio™ 6 and 7 Flex Software.

Accessory		Part number
384-Well Plate/Array Card Heated Cover		4453555
384-Well Plate Sample Block		4453553
96-Well Plate Heated Cover		4453560
96-Well Plate Sample Block		4453556
Fast 96-Well Plate Heated Cover		4459838
Fast 96-Well Plate Sample Block		4453559
Array Card Buckets/Clip Set	1st Generation	4337762
	2nd Generation	4442571
Array Card Sample Block		4453554
Array Card Staker/ Sealer		4331770
Handheld Barcode Scanner		4453271

Experiment reagents

The following table lists the reagents that can be ordered for performing experiments with the QuantStudio™ 6 and 7 Flex Software.

To perform	Recommended reagent kits	Part number
Reverse Transcription	SuperScript® VILO™ cDNA Synthesis Kit	4453650
TaqMan® PCR	TaqMan® Fast Advanced Master Mix	4444557
	TaqMan® GTXpress™ Master Mix	4401892
	TaqMan® Fast Virus 1-Step Master Mix	4444432
	TaqMan® Gene Expression Master Mix	4369016
	TaqMan® Genotyping Master Mix	4371355
	TaqMan® Universal Master Mix II, with UNG	4440038
	TaqMan® RNA-to-CT™ 1-Step Kit	4392938
SYBR® Green PCR	Fast SYBR® Green Master Mix	4385612
	Power SYBR® Green PCR Master Mix	4367659
	Power SYBR® Green RNA-to-CT™ 1-Step Kit	4389986

General-use materials and consumables

The following general-use materials and consumables are required to calibrate, maintain, and perform experiments with the QuantStudio™ 6 and 7 Flex Software. Unless indicated otherwise, all materials shown below are available from major laboratory suppliers (MLS). The materials are applicable to all sample blocks.

Material/Consumable	Source
Bleach, 10% solution	MLS
Lint-free lab tissues	MLS
Cotton or nylon swabs and lint-free cloths	MLS
Centrifuge with buckets appropriate for your consumable type	MLS
Ethanol, 95% solution	MLS
Glasses, safety	MLS
Gloves, powder-free	MLS
Permanent marker or pen	MLS
Pipettors: 100-µL and 200-µL (with pipette tips)	MLS
Screwdriver, flathead	MLS
Optical clear adhesive film for PCR	MLS
Deionized water	MLS

B Supplemental Information

This appendix covers:

(Optional) Libraries for designing your own experiments

The QuantStudio™ 6 and 7 Flex Software allows you to save information to libraries, so you can easily use the information again when setting up an experiment. The libraries include:

- Dye Library
- Targets Library
- Samples Library
- Control Library
- SNP Assay Library (only available for Genotyping experiments)
- Run Method Library
- Analysis settings Library

Dye library

You can access the Dye library from the Tools menu to add new custom dyes, edit existing dyes, and delete dyes.

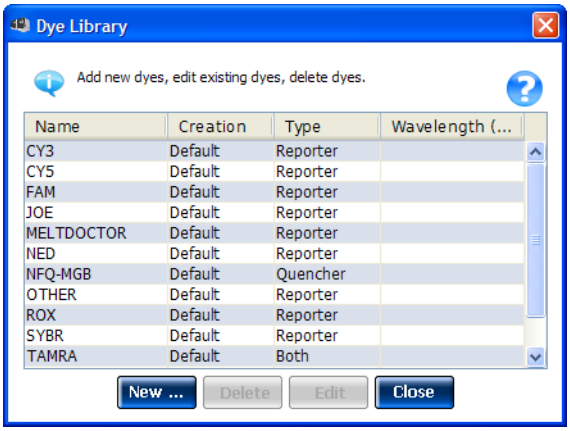
To add a new dye to the Dye Library:

1. Go to **Tools** ▸ **Dye Library...**.
2. Click **New** at the bottom of the Dye Library dialog box.

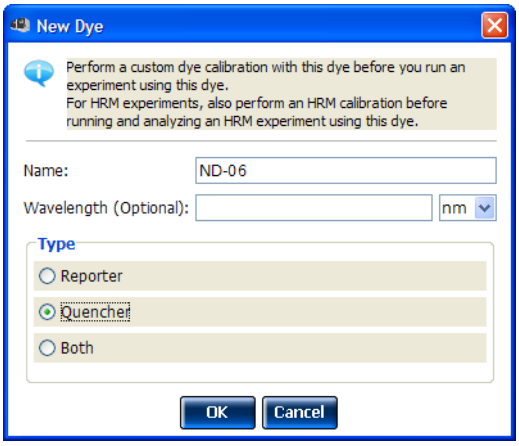
3. In the New Dye dialog box, enter the name and wavelength (optional) of the custom dye in the respective fields.

   IMPORTANT! Ensure that you perform a custom dye calibration with the new dye before you run an experiment using this dye.

4. Select the Type of dye: **Reporter**, **Quencher**, or **Both**.
   The new dye gets added to the Dye Library.



5. Click **OK**.



6. To edit or delete any of the dyes in the Dye Library, select the dye and click **Edit** or **Delete** respectively.

7. Click **Close** to exit the Dye Library.

Target, Sample, Control, and SNP Assay libraries

You can access the Targets, Samples, and SNP Assay libraries from the Tools menu to add, edit, delete, and import or export items. You can also access a library by clicking **Import from Library** in the Define screen when you are setting up an experiment.

Run Method library

You can use the Run Method library from the Run Method screen to:

- Save a new run method for later use.
- To select an existing run method for an experiment.

To add a run method to the Run Method Library:

1. Click **Save Run Method** in the toolbar of the Graphical View tab on the Run Method screen.
2. Enter a name and description (*optional*) for the run method, then click **Save**.

To select a run method from the Run Method Library:

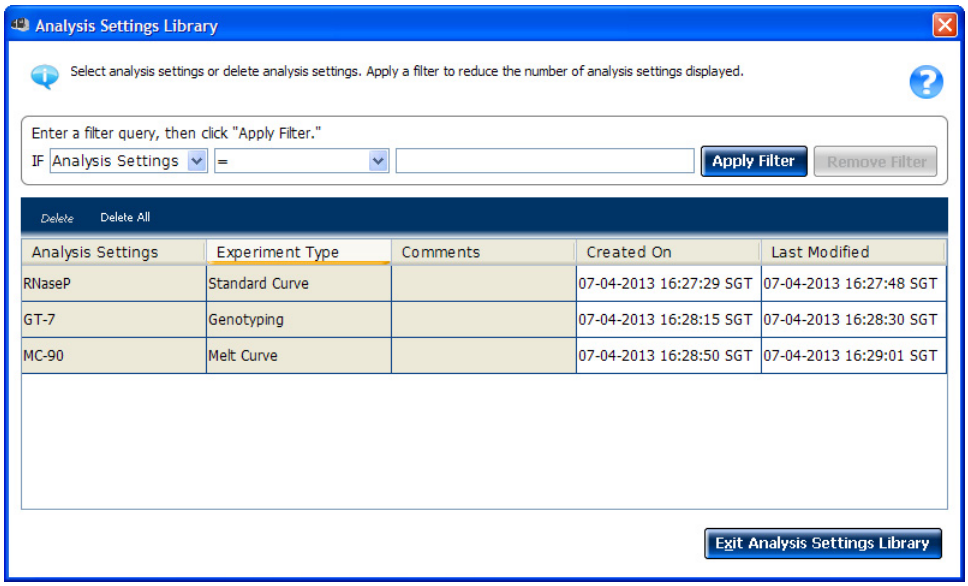
1. Click **Open Run Method** on the Run Method screen.
2. Select one from the saved run methods.
3. Click **OK**.

Analysis Settings Library

Analysis Settings are different for each experiment type. If the default analysis settings in the QuantStudio™ 6 and 7 Flex Software are not suitable for your own experiment, you can change the settings in the Analysis Settings dialog box, then reanalyze your experiment.

You can save the changed analysis settings to the Analysis Settings Library so that you can use them in other experiments.

To access the Analysis Settings Library, go to **Tools** ▸ **Analysis Settings Library**. The Analysis Settings Library dialog box looks like this:



Note: In the Analysis Settings Library dialog box you can apply a filter to reduce the number of settings protocols displayed.

To change the analysis settings and to save them to the Analysis Settings Library:

1. From the Experiment Menu pane, select **Analysis**.
2. On the Analysis screen, click **Analysis Settings** to open the Analysis Settings dialog box.
3. Change the analysis settings according to your requirement.

4. Click **Save to Library** to save the changes you have made to the Analysis Settings Library.

   Note: You can import the analysis settings you have previously saved to the Analysis Settings Library, by clicking **Load from Library** in the Analysis Settings dialog box.

Changing default analysis settings in Preferences

(Optional) For real-time data collection, you can change the default analysis settings in the Preferences for the following:

- Automatic analysis
- Automatic save
- Baseline settings

To change the default analysis settings:

1. Go to **Tools** ▸ **Preferences**.
2. Click the Experiment tab. Select the Auto Analysis and Auto Save check boxes for the QuantStudio™ 6 and 7 Flex Software to automatically analyze and save experiment results.

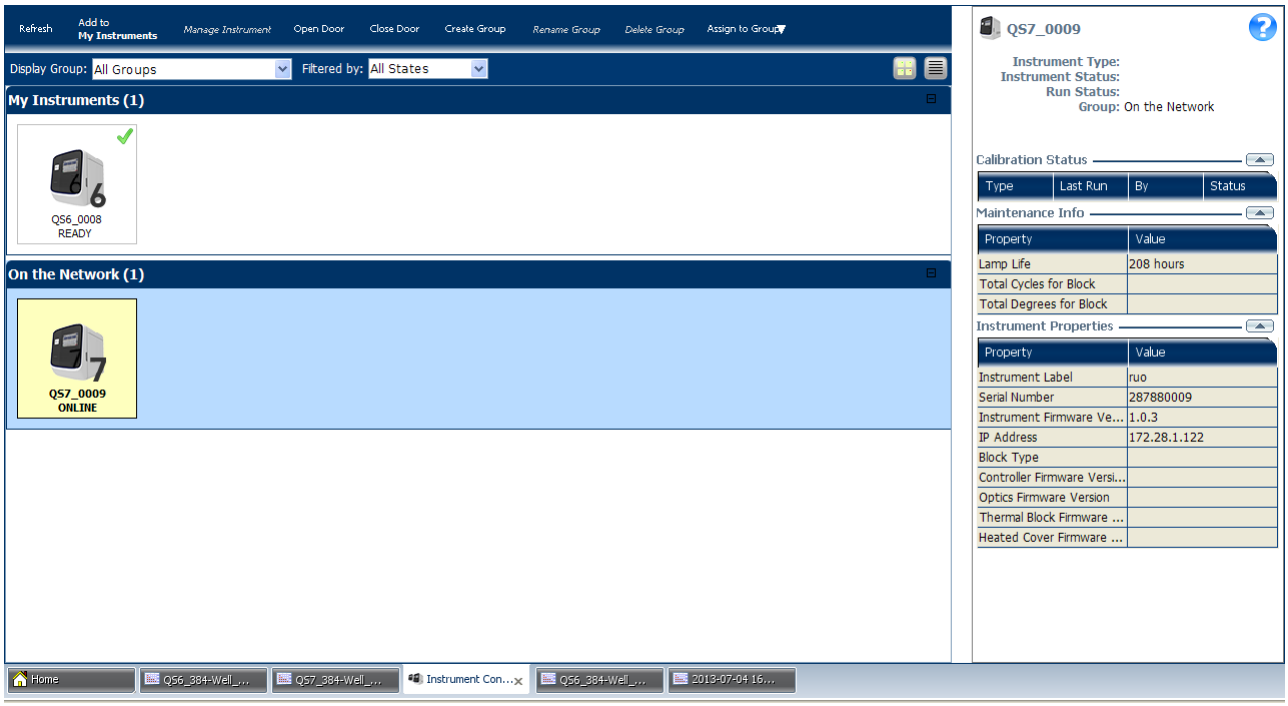
   You can also edit the following default baseline settings:

Field	Entry
Start Cycle Number	3 (default)
End Cycle Number	15 (default)

   Note: By default, the Auto Analysis and Auto Save check boxes are selected.

Instrument Console

The Instrument Console displays all the QuantStudio™ 6 and 7 Instruments discovered on a network.



Left panel

The features on the left panel of the Instrument Console allow:

- Instrument access: Open and close the QuantStudio™ 6 or 7 Instrument door from the QuantStudio™ 6 and 7 Flex Software user interface.
- Group management:
  - Create, rename, and delete groups and assign instruments to the groups.
  - Add and remove instruments to and from My Instruments.

    Note: To add instruments, select the icon of the QuantStudio™ 6 or 7 Instrument that you want to add to the My Instruments list. Then click **Add to My Instruments**. Similarly, click **Remove from My Instruments** to remove an instrument from the My Instruments list. You can also drag and drop the instrument icon into My Instruments or into the group created by you.
  - Display instrument groups from the Display Group drop-down menu. according to their activity. Select the status from the Filtered By drop-down menu. For more information on the status of an instrument, see Monitor the experiment, in Chapter 1, Booklet 1 *Getting Started with QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Experiments*.
- Instrument management:
  - Monitor experiments (check the run status or monitor a temperature plot or amplification plot during a run). For more information on monitoring experiments, see Monitor the experiment, in Chapter 1, Booklet 1 *Getting Started with QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Experiments*.
  - Maintain instruments (check the calibration status of instruments and perform different calibrations). For more information on instrument maintenance, refer to the instrument user guide.
  - Manage files (upload setup files; download completed experiments; and create, rename, and delete experiment files and plate setup folders).

    Note: Completed experiments are downloaded into the default folder **Completed Experiments**.

    Note: To manage files, click **Manage Instrument**.
    Use the File Manager to create, rename or delete folders for holding setup files for starting a run or completed experiments for analysis.
    To move setup files from one folder to the other, click **Move** and select the setup folder you want to shift the setup file into.

    IMPORTANT! To Manage and Monitor, you must move instruments from On the Network to My Instruments or a custom group. You can start a run and calibrate instruments present only in the My Instruments group or the custom group(s) that you created.

Right panel

The right panel of the Instrument Console displays:

- The name of the instrument whose instrument icon is selected.
- The run status of the selected instrument.
- The group the instrument belongs to.
- The calibration status, maintenance reminders and instrument properties of the selected instrument.

The calibration status is indicated by the ⚠ icon. The icon appears in the Status column of the Calibration Status table after the last reminder date before the calibration expires.

Status icons

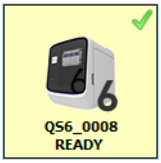
You can monitor the instrument status and view calibration and other information in the Instrument Console.

The status of an instrument is represented by an icon in the top-right corner of the thumbnail representation of the instrument on the Instrument Console. An instrument displays the status when you place the instrument icon under My Instruments or under the Group(s) that you created.

QuantStudio™ 6 or 7 Instrument status icon

To monitor the instrument status:

1. On the Home tab ( ), select **Instrument Console**. If you do not see an instrument, click **Refresh** in the instrument console toolbar.
2. If needed, move the instrument from the On the Network group to a group which can be monitored:
   a. Click the instrument of interest, then click **Assign to Group** in the instrument console toolbar.
   b. Select the **My Instruments** or a personal group in the drop-down list. The instrument is now monitored.

Icon	Instrument status
	Ready
(no icon)	Available on the network but cannot be monitored because that instrument is not under My Instruments or a group you created.
	Run in process (The time remaining for the run is shown to the left of the icon.)
	Unavailable
	Incompatible firmware version
	No longer connected to the network
	Error occurred during run

About the reagents

TaqMan® Reagents

Description

TaqMan® reagents use a fluorogenic probe to enable detection of a specific PCR product as it accumulates during PCR cycles.

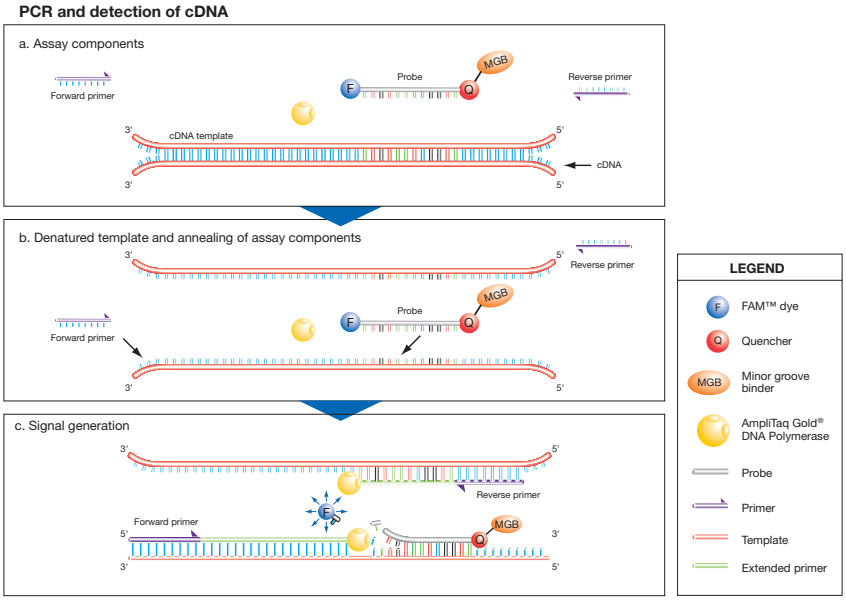
Advantages

- Increased signal specificity with the addition of a fluorogenic probe.
- Multiplex capability.
- Optional preformulated assays, optimized to run under universal thermal cycling conditions, are available.
- Can be used for either 1- or 2-step RT-PCR.

Limitations

Require synthesis of a unique fluorogenic probe.

TaqMan® Reagents detection process



SYBR® Green reagents

Description

SYBR Green reagents use SYBR® Green I dye, a double-stranded DNA binding dye, to detect PCR products as they accumulate during PCR cycles.

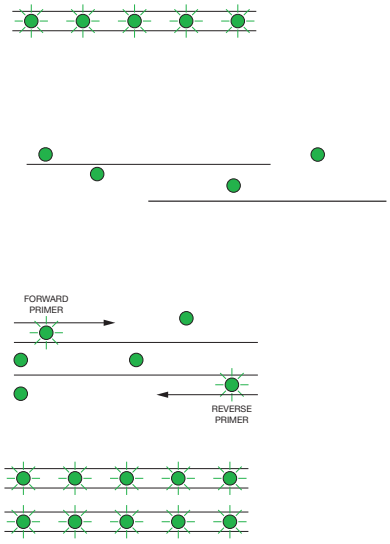
Advantages

- Economical (no probe needed).
- Allow for melt curve analysis to measure the Tm of all PCR products.
- Can be used for either 1- or 2-step RT-PCR.

Limitations

Bind nonspecifically to all double-stranded DNA sequences. To avoid erroneous information signals, check for nonspecific product formation using melt curve or gel analysis.

SYBR® Green detection process



Step 1: Reaction setup
The SYBR® Green I dye fluoresces when bound to double-stranded DNA.

Step 2: Denaturation
When the DNA is denatured into single-stranded DNA, the SYBR® Green I dye is released and the fluorescence is drastically reduced.

Step 3: Polymerization
During extension, primers anneal and PCR product is generated.

Step 4: Polymerization completed
SYBR® Green I dye binds to the double-stranded product, resulting in a net increase in fluorescence detected by the instrument.

Documentation and Support

Related documentation

The following related documents are shipped with the instrument:

Document	Pub. no.	Description
QuantStudio™ 6 and 7 flex Real-Time PCR System Software Getting Started Guide	4489822	Contains seven individual booklets that explain how to perform the six different experiments on the QuantStudio™ 6 and 7 flex Real-Time PCR System Software. The experiments include Standard Curve, Relative Standard Curve and Comparative C_T, Genotyping, Presence/ Absence and Melt Curve. Each Getting Started Guide booklet functions as both: • A tutorial, using example experiment data provided with the QuantStudio™ 6 and 7 flex Real-Time PCR System Software. • A guide for your own experiments. Intended for laboratory staff and principal investigators who perform experiments using the QuantStudio™ 6 and 7 flex Real-Time PCR System Software.
QuantStudio™ 6 and 7 Flex Real-Time PCR Systems Maintenance and Administration Guide	4489821	Explains how to use and maintain the QuantStudio™ 6 and 7 Flex Real-Time PCR Systems Intended for laboratory staff responsible for the use and maintenance of the QuantStudio™ 6 and 7 Instruments.
QuantStudio™ 6 and 7 Flex Real-Time PCR Systems Site Preparation Guide	4489824	Explains how to prepare your site to receive and install the QuantStudio™ 6 and 7 Instruments Intended for personnel who schedule, manage, and perform the tasks required to prepare your site for installation of the QuantStudio™ 6 and 7 Instruments.
QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Programming Supplement	4489825	Provides IT administrative personnel with sufficient information to integrate the instrument and software with a LIS/LIMS. Intended to be used with the *QuantStudio™ 6 and 7 Flex Real-Time PCR System Software Getting Started Guide*.

Document	Pub. no.	Description
Applied Biosystems® Twister® Robot Automation Accessory Quick Reference	4468146	Provides abbreviated instructions for operating an Applied Biosystems® Twister® Robot Automation Accessory that has been installed with the QuantStudio™ 6 or 7 Flex Real-Time PCR System. Intended for laboratory staff and principal investigators who perform experiments using the QuantStudio™ 6 or 7 Flex Real-Time PCR System.
QuantStudio™ 6 and 7 flex Real-Time PCR System Software Help	NA	Explains how to use the QuantStudio™ 6 and 7 flex Real-Time PCR System Software to: • Set up, run, analyze, audit, sign, export, and print experiments. • Monitor networked QuantStudio™ 6 and 7 Instruments. • Calibrate and verify the performance of the QuantStudio™ 6 and 7 Instruments. • Intended for laboratory staff and principal investigators who perform experiments using the QuantStudio™ 6 and 7 flex Real-Time PCR System Software.

Note: For additional documentation, see "How to obtain support" on page 25.

Other related documents

Documents related to Standard Curve experiments

Document	Pub. no.
Amplification Efficiency of TaqMan® Gene Expression Assays Application Note	127AP05
Custom TaqMan® Gene Expression Assays Protocol	4334429
Primer Express® Software Version 3.0 Getting Started Guide	4362460
TaqMan® Gene Expression Assays Protocol	4333458
User Bulletin #2: Relative Quantitation of Gene Expression	4303859

Documents related to Relative Standard Curve and Comparative C_T experiments

Document	Pub. no.
Amplification Efficiency of TaqMan® Gene Expression Assays Application Note	127AP05
Applied Biosystems® High-Capacity cDNA Reverse Transcription Kits Protocol	4375575
Custom TaqMan® Gene Expression Assays Protocol	4334429
Primer Express® Software Version 3.0 Getting Started Guide	4362460
TaqMan® Gene Expression Assays Protocol	4333458
User Bulletin #2: Relative Quantitation of Gene Expression	4303859

Documents related to Genotyping experiments

Document	Pub. no.
Allelic Discrimination Pre-Developed TaqMan® Assay Reagents Quick Reference Card	4312212
Custom TaqMan® Genomic Assays Protocol	4367671
Custom TaqMan® SNP Genotyping Assays Protocol	4334431
Ordering TaqMan® SNP Genotyping Assays Quick Reference Card	4374204
Pre-Developed TaqMan® Assay Reagents Allelic Discrimination Protocol	4312214
TaqMan® Drug Metabolism Genotyping Assays Protocol	4362038
TaqMan® SNP Genotyping Assays Protocol	4332856

Documents related to Presence/ Absence experiments

Document	Pub. no.
DNA Isolation from Fresh and Frozen Blood, Tissue Culture Cells, and Buccal Swabs Protocol	4343586
NucPrep® Chemistry: Isolation of Genomic DNA from Animal and Plant Tissue Protocol	4333959
PrepMan® Ultra Sample Preparation Reagent Protocol	4318925

How to obtain support

For the latest services and support information for all locations, go to:

www.lifetechnologies.com/support

At the Life Technologies web site, you can:

- Access worldwide telephone and fax numbers to contact Life Technologies Technical Support and Sales facilities.
- Search through frequently asked questions (FAQs).
- Submit a question directly to Technical Support.
- Order Life Technologies user documents, SDSs, certificates of analysis, and other related documents.
- Download PDF documents.
- Obtain information about customer training.
- Download software updates and patches.

Obtaining information from the Help system

The QuantStudio™ 6 and 7 flex Real-Time PCR System Software has a Help system that describes how to use each feature of the user interface. Access the Help system by doing one of the following:

- Click in the toolbar of the QuantStudio™ 6 and 7 flex Real-Time PCR System Software window.
- Select **Help ▸ QuantStudio™ 6 and 7 flex Real-Time PCR System Software Help**.
- Press **F1**.

You can use the Help system to find topics of interest by:

- Reviewing the table of contents
- Searching for a specific topic

Limited Product Warranty

Life Technologies Corporation and/or its affiliate(s) warrant their products as set forth in the Life Technologies' General Terms and Conditions of Sale found on Life Technologies' website at **www.lifetechnologies.com/termsandconditions**. If you have any questions, please contact Life Technologies at **www.lifetechnologies.com/support**.

Glossary

AIF	See assay information file (AIF).
AIX	XML version of the assay information file. See also assay information file (AIF).
allele	In a diploid organism, one of two DNA sequences found at the same locus (for example, a particular gene), but located on homologous chromosomes. Two corresponding alleles may have the identical sequence, or they may differ somewhat, often at one or more single-base sites (SNPs).
aligned melt curve plot	Plot of the re-scaled melt curve.
allelic discrimination plot	Display of genotyping data collected during the post-PCR read. The allelic discrimination plot is a graph of the normalized reporter signal from the allele 1 probe, plotted against the normalized reporter signal from the allele 2 probe.
amplicon	A segment of DNA amplified during PCR.
amplification	Part of the instrument run in which PCR amplifies the target. Fluorescence data collected during amplification are displayed in an amplification plot, and the data are used to calculate results. **Note:** Only quantitative real-time PCR experiments, not end-point experiments, take amplification data into account.
amplification efficiency (EFF%)	Calculation of the efficiency of the PCR amplification in an experiment. EFF% is calculated using the slope of the regression line in the standard curve. A slope close to –3.32 indicates optimal, 100% PCR amplification efficiency.
amplification plot	Display of data collected during the cycling stage of PCR amplification. The amplification plot can be viewed as: • Baseline-corrected normalized reporter (ΔRn) vs. cycle • Normalized reporter (Rn) vs. cycle • Threshold cycle (C_T) vs. well
amplification stage	Part of the instrument run in which PCR amplifies the target. The amplification stage, called a cycling stage in the thermal profile, consists of denaturing, primer annealing, and extension steps that are repeated. Fluorescence data collected during the extension stage are displayed in an amplification plot, and the data are used to calculate results. With TaqMan® chemistry, the last two steps of a PCR stage are typically combined. See also cycling stage.

Analysis Settings Library	In the software, a collection of analysis settings to use in experiments. You can save settings and reuse them. You cannot edit or import settings into the library.
assay	In a PCR reaction mix, two target-specific primers or two primers and a probe used to amplify a target.
Assay ID	Identifier assigned by Life Technologies to TaqMan® assays.
assay information file (AIF)	Tab-delimited data file on a CD shipped with each assay order. The AIF contains technical details about all assays in the shipment. It includes information about assay concentrations; reporters and quenchers used; part and lot numbers; and assay, vial, and plate ID numbers. The file name includes the number from the barcode on the plate.
assay mix	PCR reaction component in Applied Biosystems® TaqMan® assays. The assay mix contains primers designed to amplify a target and a TaqMan® probe designed to detect amplification of the target.
AutoDelta	In the run method, a setting to increase or decrease the temperature and/or time for a step with each subsequent cycle in a cycling stage. When AutoDelta is enabled for a cycling stage, the settings are indicated by an icon in the thermal profile: • AutoDelta on:  • AutoDelta off: 
automatic baseline	An analysis setting in which the software calculates the baseline start and end cycles for the amplification plot. See also baseline.
automatic threshold	An analysis setting in which the software calculates the baseline start and end cycles and the threshold in the amplification plot. The software uses the baseline and threshold to calculate the threshold cycle (C_T). See also threshold cycle (C_T).
background calibration	Type of calibration in which the instrument performs reads of a background plate, averages the spectra recorded during the run, and extracts the resulting spectral component to a calibration file. The software then uses the calibration file during subsequent runs to remove the background fluorescence from the run data.
baseline	In the amplification plot, a cycle-to-cycle range that defines background fluorescence. This range can be set manually on an assay-by-assay basis, or automatically to set each individual well.
baseline-corrected normalized reporter (ΔRn)	The magnitude of normalized fluorescence signal generated by the reporter. In experiments that contain data from real-time PCR, the magnitude of normalized fluorescence signal generated by the reporter at each cycle during the PCR amplification. In the ΔRn vs Cycle amplification plot, ΔRn is calculated at each cycle as: ΔRn (cycle) = Rn (cycle) –Rn (baseline), where Rn = normalized reporter

	In genotyping experiments and presence/absence experiments, the difference in normalized fluorescence signal generated by the reporter between the pre-PCR read and the post-PCR read. In the allelic discrimination plot (genotyping experiments) and the presence/absence plot (presence/absence experiments), ΔRn is calculated as: ΔRn = Rn (post-PCR read) –Rn (pre-PCR read), where Rn = normalized reporter See also normalized reporter (Rn).
baseline threshold algorithm	Expression estimation algorithm (C_T) which subtracts a baseline component and sets a fluorescent threshold in the exponential region for gene quantification.
biological replicates	Reactions that contain identical components and volumes, but evaluate separate samples of the same biological source (for example, samples from three different mice of the same strain, or separate extractions of the same cell line or tissue sample). When an experiments uses biological replicate groups in a gene expression study, the values displayed in the Biological Replicates tab are calculated by combining the results of the separate biological samples and treating this collection as a single population (that is, as one sample). For ΔC_T computations (normalizing by the endogenous control) in a singleplex experiment, the software treats separate biological samples as unpaired data when computing variability estimates of the single biological replicate. Individual contributions of the separate biological samples to the single biological replicate results are observed in the Technical Replicates tab. See also technical replicates.
blocked IPC	In presence/absence experiments, a reaction that contains IPC blocking agent, which blocks amplification of the internal positive control (IPC). In QuantStudio™ 6 and 7 Flex Real-Time PCR System Software, also the name of the task for the IPC target in wells that contain IPC blocking agent. See also negative control-blocked IPC wells.
calibrator	See reference sample.
chemistry	See reagents.
comparative C_T ($\Delta\Delta C_T$) method	Method for determining relative target quantity in samples. The software measures amplification of the target and of the endogenous control in samples and in a reference sample. Measurements are normalized using the endogenous control. The software determines the relative quantity of target in each sample by comparing normalized target quantity in each sample to normalized target quantity in the reference sample.
C_T	See threshold cycle (C_T).
C_T algorithm	Algorithm used to determine the threshold cycle. The software provides the Baseline Threshold C_T algorithm.
cycle threshold	See threshold cycle (C_T).
cycling stage	In the thermal profile, a stage that is repeated. A cycling stage is also called an amplification stage. See also amplification stage.
C_q	See quantification cycle (C_q).

data collection	During the instrument run, a process in which an instrument detects fluorescence data from each well of the reaction plate. The instrument transforms the signal to electronic data and saves the data in the experiment file. In the QuantStudio™ 6 and 7 Flex Real-Time PCR System Software, a data collection point is indicated by an icon in the thermal profile: • Data collection on:  • Data collection off: 
delta R_n (ΔR_n)	See baseline-corrected normalized reporter (ΔRn).
derivative reporter (–Rn‘)	The rate of change in fluorescence as a function of temperature. Rn‘ is used to plot a melting curve. Significant decrease of the fluorescent signal generates a positive peak on the derivative view of the melting curve.
diluent	A reagent used to dilute a sample or standard before it is added to the PCR reaction.
dilution factor	See serial factor.
dye calibration	Type of calibration in which the software collects spectral data from a series of dye standards and stores the spectral information for the dye standards in a pure spectra calibration file. This file is used during experiment runs to characterize and distinguish the individual contribution of each dye in the total fluorescence collected by the instrument.
EFF%	See amplification efficiency (EFF%).
efficiency correction	In Comparative C_T experiments, a feature that allows you to manually enter previously-determined amplification efficiencies for each experiment, following the experimental run. The real-time software mathematically compensates for differences in efficiency between each target assay and the endogenous control when calculating sample-to-sample relative quantities. This method can be employed as a substitute for the Relative Standard Curve Method.
endogenous control	A gene that is used to normalize template differences and sample-to-sample or run-to-run variation.
endpoint read	See post-PCR read.
error	The standard error of the slope of the regression line in the standard curve. The error can be used to calculate a confidence interval (CI) for the slope. Because the amplification efficiency (EFF%) is calculated from the slope, knowing the error allows a CI for the amplification efficiency to be calculated.
experiment	Refers to the entire process of performing a run, including setup, run, and analysis. You can perform the following types of experiments: • Quantification - Standard curve • Quantification - Relative standard curve • Quantification - Comparative C_T ($\Delta\Delta C_T$)

- Melt Curve
- Genotyping
- Presence/Absence

experiment name Entered during experiment setup, the name that is used to identify the experiment.

Experiment Setup A software feature that allows you to set up an experiment according to your experiment design. Experiment Setup provides you with maximum flexibility in the design and setup of your experiment.

experiment type The type of experiment to perform:

- Standard curve
- Comparative C_T ($\Delta\Delta C_T$)
- Relative standard curve
- Genotyping
- Presence/Absence
- Melt curve

The experiment type that you select affects setup, run, and analysis.

export A software feature that allows you to export experiment setup files, experiment results, instrument information, and security and auditing settings to spreadsheet, presentation, or text files. You can edit the default location of the exported file.

filter Dye excitation and emission filter combination that you select for an experiment. The QuantStudio™ 6 and 7 Instruments include a five-color filter set and six-color filter set, respectively, that support FAM™, SYBR® Green, VIC®, JOE™, NED™, Cy®3, TAMRA™, ROX™, and Texas Red® dyes.

flag A quality control (QC) indicator which, when applied by the software to a well during analysis, indicates a possible issue with that reaction. For example, a flag may be issued if no amplification is detected in a well. Flags indicating potential problems are displayed in the Quality Control tab of the plate layout, well table, and QC Summary screens.

forward primer Oligonucleotide that flanks the 5’ end of the amplicon. The reverse primer and the forward primer are used together in PCR reactions to amplify the target.

genotyping experiment An experiment used to identify known mutations in a DNA sample. With this experiment type, you can determine if a DNA sample is:

- Homozygous (samples having only allele 1). Also called wild type homozygote.
- Homozygous (samples having only allele 2). Also called variant homozygote.
- Heterozygous (samples having both allele 1 and allele 2).

heterozygote Samples having both allele 1 and allele 2.

See also genotyping experiment.

holding stage	In the thermal profile, the stage that holds the temperature constant for a defined period of time. A stage that includes one or more steps. You can add a holding stage to the thermal profile to activate enzymes, to inactivate enzymes, or to incubate a reaction.
homozygote	Samples having only allele 1 or only allele 2. See also genotyping experiment.
housekeeping gene	A gene that is involved in basic cellular functions and that may be constitutively expressed. Housekeeping genes may be candidates for use as endogenous controls; however, their constancy should always be validated experimentally. See also endogenous control.
import	A software feature that allows you to import plate setup information or security settings before an experiment run. You can also import information into some libraries in the software.
Instrument Console	A software feature that allows you to view information about instruments on the network. In the Instrument Console, you can monitor the status of any instrument on the network; view calibration, maintenance, and instrument properties for a selected instrument; and open and close the instrument drawer.
Instrument Manager	A software feature that allows you to view information about instrument available on the network. In the Instrument Manager, you can monitor the status of an instrument; monitor amplification plots and temperature plots in real time; view the calibration status, perform calibrations and manage files on the instrument, including downloading completed experiments to your computer.
internal positive control (IPC)	In presence/absence experiments, a short synthetic DNA template that is added to PCR reactions. The IPC can be used to distinguish between true negative results (the target is absent in the samples) and negative results caused by PCR inhibitors, incorrect assay setup, or reagent or instrument failure.
inventoried assays	TaqMan® Gene Expression Assays and TaqMan® SNP Genotyping Assays that have been previously manufactured, passed quality control specifications, and stored in inventory.
IPC	See internal positive control (IPC).
IPC blocking agent	Reagent added to PCR reactions to block amplification of the internal positive control (IPC).
IPC+	See negative control-IPC wells.
made-to-order assays	TaqMan® Gene Expression Assays that are manufactured at the time of order. Only assays that pass manufacturing quality control specifications are shipped.
manual baseline	An analysis setting for the Baseline Threshold algorithm. You enter the baseline start and end cycles for the amplification plot. See also baseline.

manual threshold	An analysis setting for the Baseline Threshold algorithm. You enter the threshold value and select whether to use automatic baseline or manual baseline values. The software uses the baseline and the threshold values to calculate the threshold cycle (C_T).
melt curve	A plot of data collected during the melt curve stage. Peaks in the melt curve can indicate the melting temperature (T_m) of the target, or they can identify nonspecific PCR amplification. In the software, you can view the melt curve as normalized reporter (R_n) vs. temperature or as derivative reporter ($-R_n'$) vs. temperature. In a high resolution melting experiment, you can view the melt curve as fluorescence vs. temperature. Also called dissociation curve.
melt curve characteristics	The melt curve shape and the difference in melting temperature (T_m) values.
melt curve stage	In the thermal profile, a stage with a temperature increment to generate a melt curve.
melt curve plot	The default view of the melting curve. It plots the negative derivative data (–Rn‘) vs. temperature.
melting temperature (T_m)	The temperature at which 50% of the DNA is double-stranded and 50% of the DNA is dissociated into single-stranded DNA. In a melt curve experiment, the melt curve plot displays the melting temperature.
multicomponent plot	A plot of the complete spectral contribution of each dye for the selected well(s) over the duration of the PCR run.
negative control (NC)	The task for targets or SNP assays in wells that contain water or buffer instead of sample. No amplification of the target should occur in negative control wells. Previously called no template control (NTC).
negative control-blocked IPC wells	In presence/absence experiments, wells that contain IPC blocking agent instead of sample in the PCR reaction. No amplification should occur in negative control-blocked IPC wells because the reaction contains no sample and amplification of the IPC is blocked. Previously called no amplification control (NAC).
negative control-IPC wells	In presence/absence experiments, wells that contain IPC template and buffer or water instead of sample. Only the IPC template should amplify in negative control-IPC wells because the reaction contains no sample. Previously called IPC+.
no amplification control (NAC)	See negative control-blocked IPC wells.
no template control (NTC)	See negative control (NC).
nonfluorescent quencher-minor groove binder (NFQ-MGB)	Molecules that are attached to the 3’ end of TaqMan® probes. When the probe is intact, the nonfluorescent quencher (NFQ) prevents the reporter dye from emitting fluorescence signal. Because the NFQ does not fluoresce, it produces lower background signals, resulting in improved precision in quantification. The minor groove binder (MGB) increases the melting temperature (T_m) of the probe without increasing its length, allowing for the design of shorter probes.

normalization calibration	Type of calibration in which the software collects data from the normalization standards, then stores it in a normalization calibration file. This file is used in comparisons of data from multiple instruments within a study.
normalized quantity	Either the C_T Avg. of the target gene minus the C_T Avg. of the endogenous control (Comparative C_T experiments), or the Q Avg. of the target divided by the Q Avg. of the endogenous control (Relative Standard Curve experiments).
normalized quantity mean	The relative standard curve equivalent of the ΔC_T mean value found in Comparative C_T experiments (computed as the geometric mean).
normalized quantity SE	The relative standard curve equivalent of the ΔC_T SE value found in Comparative C_T experiments (computed as the geometric standard error of the mean).
normalized reporter (Rn)	Fluorescence signal from the reporter dye normalized to the fluorescence signal of the passive reference dye (usually ROX™ dye on Life Technologies instruments).
omit well	An action that you perform before reanalysis to omit one or more wells from analysis. Because no algorithms are applied to omitted wells, omitted wells contain no results. You can add wells back in to the analysis; no information is permanently discarded.
outlier	A measurement (such as a C_T) that deviates significantly from the measurement of the other replicates for that same sample.
passive reference	A dye that produces fluorescence signal independent of PCR amplification, and that is added to each reaction at a constant concentration. Because the passive reference signal should be consistent across all wells, it is used to normalize the reporter dye signal to account for non-PCR related fluorescence fluctuations caused by minor well-to-well differences in volume. Normalization to the passive reference signal generally results in data with noticeably high precision among technical replicates.
plate layout	An illustration of the grid of wells and assigned content in the reaction plate. The number of rows and columns in the grid depends on the sample block that you use. In the software, you can use the plate layout as a selection tool to assign well contents, to view well assignments, and to view results. The plate layout can be printed, included in a report, exported, and saved as a slide for a presentation.
plate setup file	A file (.txt, .csv, .xml, or .sds) that contains setup information such as the well number, sample name, sample color, target name, dyes, and other reaction plate contents.
point	One standard in a standard curve. The standard quantity for each point in a standard curve is calculated based on the starting quantity and serial factor.
positive control	In genotyping and presence/absence experiments, a DNA sample with a known genotype, homozygous or heterozygous. In the software, the task for the SNP assay in wells that contain a sample with a known genotype.

post-PCR read	In genotyping and presence/absence experiments, the part of the instrument run that occurs after amplification. In genotyping experiments, fluorescence data collected during the post-PCR read are displayed in the allelic discrimination plot and used to make allele calls. In presence/absence experiments, fluorescence data collected during the post-PCR read are displayed in the presence/absence plot and used to make detection calls. Also called endpoint read.
pre-PCR read	In genotyping and presence/absence experiments, the part of the instrument run that occurs before amplification. The pre-PCR read is optional but recommended. Fluorescence data collected during the pre-PCR read can be used to normalize fluorescence data collected during the post-PCR read.
primer mix	PCR reaction component that contains the forward primer and reverse primer designed to amplify the target.
primer/probe mix	PCR reaction component that contains the primers designed to amplify the target and a TaqMan® probe designed to detect amplification of the target.
pure dye	Fluorescent compound used to calibrate the instrument. See system dye.
quantification cycle (C_q)	The fractional PCR cycle used for quantification, according to the Real-time PCR Data Markup Language (RDML) data standard. C_T is the algorithm-specific calculations of C_q.
quantification method	In quantification experiments, the method used to determine the quantity of target in the samples.
quantity	In quantification experiments, the amount of target in the samples. Absolute quantity can refer to copy number, mass, molarity, or viral load. Relative quantity refers to the fold-difference between normalized quantity of target in the sample and normalized quantity of target in the reference sample.
quencher	A molecule attached to the 3’ end of TaqMan® probes to prevent the reporter from emitting fluorescence signal while the probe is intact. With TaqMan® reagents, a nonfluorescent quencher-minor groove binder (NFQ-MGB) can be used as the quencher.
QuickStart	A feature that allows you to run an experiment without entering plate setup information, if your instrument and computer are in the same network. QuickStart requires an experiment template file.
R^2 value	Regression coefficient calculated from the regression line in the standard curve. An important quality value, the R^2 value indicates the closeness of fit between the standard curve regression line and the individual C_T data points from the standard reactions. A value of 1.00 indicates a perfect fit between the regression line and the data points.
ramp	The step at which the temperature changes during the instrument run. The ramp rate is defined as °C per second. In the graphical view of the thermal profile, the ramp rate is indicated by a diagonal line.

ramp speed	Speed at which the temperature ramp occurs during the instrument run. Available ramp speeds include fast and standard.
Raw data plot	A plot of raw fluorescent signal as detected through each emission filter, used to view raw data for individual wells and at individual cycles.
reaction mix	A solution that contains all components to run the PCR reaction, except for the template (sample, standard, or control). Also called a "PCR cocktail".
reagents	The PCR reaction components used to amplify the target and to detect amplification.
real-time PCR	Process of collecting fluorescence data during PCR. Data from the real-time PCR are used to calculate results for quantification experiments or to troubleshoot results for genotyping or presence/absence experiments.
Real-time PCR Data Markup Language (RDML)	A reporting format that is compliant with the Minimum Information for Publication for Quantitative Real Time Experiments (MIQE) guidelines.
reference sample	In relative standard curve and Comparative C_T ($\Delta\Delta C_T$) experiments, the sample used as the basis for relative quantification results. Also called the calibrator.
refSNP ID	The reference SNP (refSNP) cluster ID. Generated by the Single Nucleotide Polymorphism Database of Nucleotide Sequence Variation (dbSNP) at the National Center for Biotechnology Information (NCBI). The refSNP ID can be used to search the Life Technologies Store for an Applied Biosystems® SNP Genotyping Assay. Also called an rs number.
region of interest (ROI) calibration	Type of calibration in which the software maps the positions of the wells on the sample block of the instrument. The software uses the ROI calibration data to associate increases in fluorescence during a run with specific wells of the plate. A calibration image for each individual filter must be generated to account for minor differences in the optical path.
regression coefficients	Values calculated from the regression line in standard curves, including the R^2 value, slope, and y-intercept. You can use the regression coefficients to evaluate the quality of results from the standards. See also standard curve.
regression line	In standard curve and relative standard curve experiments, the best-fit line from the standard curve. Regression line formula: $C_T = m [\log (Qty)] + b$ where m is the slope, b is the y-intercept, and Qty is the standard quantity. See also regression coefficients.
reject well	An action that the software performs during analysis to remove one or more wells from further analysis if a specific flag is applied to the well.

relative standard curve method	An experimental method to determine relative quantities. This method compensates for target and endogenous control efficiency differences within each run. In all experiments, unknown samples and dilution series of template (such as cDNA) are amplified. Following a run, the instrument software interpolates relative quantities for each unknown sample from the appropriate dilution curve, then normalizes the data for each sample (or set of replicates) as follows: target $Q_{Avg.}$ ÷ endogenous control Q_{Avg}.
replicate group	A user-defined biological grouping. A replicate group may be a set of identical reactions in an experiment.
replicates	Total number of identical reactions containing identical components and identical volumes.
reporter	A fluorescent dye used to detect amplification. With TaqMan® reagents, the reporter dye is attached to the 5′ end. With SYBR® Green reagents, the reporter dye is SYBR® Green dye. SYBR® dyes are DNA-binding dyes.
reverse primer	An oligonucleotide that flanks the 3′ end of the amplicon. The reverse primer and the forward primer are used together in PCR reactions to amplify the target.
reverse transcriptase	An enzyme that converts RNA to cDNA.
R_n	See normalized reporter (Rn).
ROX™ dye	A dye supplied by Life Technologies, which is used as a passive reference in some experiments.
rs number	See refSNP ID.
run method	Definition of the reaction volume and the thermal profile for the instrument run. The run method specifies the temperature, time, ramp, and data collection points for all steps and stages of the instrument run.
sample	The biological tissue or specimen that you are testing for a target gene.
sample definition file	A tab-delimited file (*.txt or *.csv) that contains the following setup information: well number, sample name, and custom sample properties.
Sample Library	In the software, an editable collection of sample names to use in experiments. The samples in the library contain the sample name and the sample color. The samples in the library may also contain comments about the sample.
sample/SNP assay reaction	In genotyping experiments, the combination of the sample to test and the SNP assay to perform in one PCR reaction. Each PCR reaction can contain only one sample and one SNP assay.
sample/target reaction	In quantification experiments, the combination of the sample to test and the target to detect and quantify in one PCR reaction.

security, auditing and eSignature

An optional software module that provides:

- **System Security** – Controls user access to the software. Provides a default Administrator user account. You can define additional user accounts and permissions.
- **Auditing** – Tracks changes made to library items, actions performed by users, and changes to the Security and Audit settings. The software automatically audits some actions silently. You can select other items for auditing and specify the audit mode. Provides reports for audited library items, Security and Audit changes, and actions.
- **Electronic Signature (eSignature) –** Controls whether users are permitted, prompted, or required to provide a user name and password when accessing certain software features. You can select which features are controlled and the number of signatures required for access. When authorized persons use this feature, they are creating a legally binding signature.

serial factor

In the software, a numeric value that defines the sequence of quantities in the standard curve. The serial factor and the starting quantity are used to calculate the standard quantity for each point in the standard curve. For example, if the standard curve is defined with a serial factor of 1:10 or 10✷, the difference between any 2 adjacent points in the curve is 10-fold.

slope

Regression coefficient calculated from the regression line in the standard curve. The slope indicates the PCR amplification efficiency for the assay. A slope of –3.32 indicates 100% amplification efficiency.

See also amplification efficiency (EFF%) and regression line.

SNP

Single nucleotide polymorphism. The SNP can consist of a base difference or an insertion or deletion of one base.

SNP assay

Used in genotyping experiments, a PCR reaction that contains primers to amplify the SNP and two probes to detect different alleles.

SNP Assay Library

In the software, an editable collection of SNP assays to add to genotyping experiments. The SNP assays in the library contain the SNP assay name; SNP assay color; and for each allele, the allele name or base(s), reporter, quencher, and allele colors. The SNP assays in the library may also contain the assay ID and comments about the SNP assay.

stage

In the thermal profile, a group of one or more steps. Examples: PCR stage, cycling stage (also called amplification stage), and hold stage.

standard

A sample that you dilute and amplify along with unknown samples. This dilution series can contain known starting quantities of the target of interest (absolute standard curve) or it can be of known dilution factor (relative standard curve). Following the run, the software interpolates the C_T values of the unknowns to this curve, yielding either specific quantities of the target (for absolute curves) or relative quantities (for relative dilution curves).

See also standard curve.

standard curve

In standard curve and relative standard curve experiments:

- The best-fit line in a plot of the C_T values from the standard reactions plotted against standard quantities. See also regression line.
- A set of standards containing a range of known quantities. Results from the standard curve reactions are used to generate the standard curve. The standard curve is defined by the number of points in the dilution series, the number of standard replicates, the starting quantity, and the serial factor.

standard curve method

Method for determining absolute target quantity in samples. With the standard curve method, the software measures amplification of the target in samples and in a standard dilution series. Data from the standard dilution series are used to generate the standard curve. Using the standard curve, the software interpolates the absolute quantity of target in the samples.

See also standard and standard curve.

standard dilution series

In standard curve and relative standard curve experiments, a set of standards containing a range of known quantities. The standard dilution series is prepared by serially diluting standards. For example, the standard stock is used to prepare the first dilution point, the first dilution point is used to prepare the second dilution point, and so on. In the software, the volumes needed to prepare a standard dilution series are calculated by the number of dilution points, the number of standard replicates, the starting quantity, the serial factor, and the standard concentration in the stock. See also standard curve.

standard quantity

In the PCR reaction, a known quantity. In standard curve experiments, the quantity of target in the standard. In the software, the units for standard quantity can be for mass, copy number, viral load, or other units for measuring the quantity of target. Standard quantity can also refer to dilution factor.

starting quantity

When defining a standard curve in the software, the highest quantity.

step

A component of the thermal profile. For each step in the thermal profile, you can set the ramp rate (ramp increment for melt curve steps), hold temperature, and hold time (duration).You can turn data collection on or off for the ramp or the hold parts of the step. For cycling stages, a step is also defined by the AutoDelta status.

SYBR® Green reagents

PCR reaction components that consist of two primers designed to amplify the target and SYBR® Green dye to facilitate detection of the PCR product.

system dye

Dye supplied by Life Technologies. Before you use system dyes in your experiments, make sure the system dye calibration is current in the Instrument Console.

The system dyes are:

- FAM™ dye
- JOE™ dye
- ROX™ dye
- NED™ dye
- SYBR® Green dye
- TAMRA™ dye

	• VIC® dye • Cy®3 dye • Texas Red® dye
TaqMan® reagents	PCR reaction components that consist of primers designed to amplify the target and a TaqMan® probe designed to detect amplification of the target.
target	The nucleic acid sequence to amplify and detect.
target color	In the software, a color assigned to a target to identify the target in the plate layout and analysis plots.
Target Library	In the software, an editable collection of targets to use in experiments. Targets in the library contain the target name, reporter, quencher, and target color. The targets in the library may also contain comments about the target.
task	In the software, the type of reaction performed in the well for the target or SNP assay. Available tasks: • Unknown • Negative Control • Standard (standard curve and relative standard curve experiments) • Positive control (genotyping experiments) • IPC (presence/absence experiments) • Blocked IPC (presence/absence experiments)
technical replicates	Wells containing identical reaction components, including sample; important for evaluating precision.
temperature plot	In the software, a display of temperatures for the instrument cover and instrument block during the instrument run.
template	The type of nucleic acid to add to the PCR reaction.
template file	A user-created file that contains experiment setup information (experiment type, sample names, target name, and thermal conditions) to be used as a starting point for new experiment setup. Template files have an .edt extension.
thermal profile	Part of the run method that specifies the temperature, time, ramp, and data collection points for all steps and stages of the instrument run.
threshold	• In amplification plots, the level of fluorescence above the baseline and within the exponential growth region. For the Baseline Threshold algorithm, the threshold can be determined automatically (see automatic threshold) or can be set manually (see manual threshold). • In presence/absence experiments, the level of fluorescence above which the software assigns a presence call.

threshold cycle (C_T)	The PCR cycle number at which the fluorescence meets the threshold in the amplification plot.
T_m	See melting temperature (T_m).
touchscreen	Instrument display that you touch to control the instrument.
uniformity calibration	Type of calibration in which the software measures sample block uniformity. The calibration generates data that compensate for the physical effects of the QuantStudio™ 6 and 7 Instruments' filters on data collected during an experiment.
unknown	In the software, the task for the target or SNP assay in wells that contain the sample being tested. In quantification experiments, the task for the target in wells that contain a sample with unknown target quantities. In genotyping experiments, the task for the SNP assay in wells that contain a sample with an unknown genotype. In presence/absence experiments, the task for the target in wells that contain a sample in which the presence of the target is not known. In melt curve experiments, the task for the target in wells that contain a sample with an unknown melt curve profile.
unknown-IPC wells	In presence/absence experiments, wells that contain a sample and internal positive control (IPC).
variant	A sample (or group of samples) with a unique melt curve (that is, the melt curve is different from the melt curves of other samples or controls used in the experiment). The software determines melt curve differences by the melting temperature (Tm) and the shape of the melt curve.
y-intercept	In the standard curve, the value of y where the regression line crosses the y-axis. The y-intercept indicates the expected threshold cycle (C_T) for a sample with quantity equal to 1.

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